

Research on the effect of Formononetin on photodynamic therapy in K562 cells

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Abstract: At the present time, many cancer patients combine some forms of complementary and alternative medicine therapies with their conventional therapies. The most common choice of these therapies is the use of antioxidants. Formononetin (FORM) is presented in different foods. It has a variety of biological activities including antioxidant and anti-cancer properties. On account of its antioxidant activity, FORM might protect cancer cells from free radical damage in photodynamic therapy (PDT) during which reactive oxygen species (ROS) production was stimulated leading to irreversible tumor cell injury. In this study, the influence of FORM on K562 cells in PDT was demonstrated. The results showed that FORM supplementation alone did not affect the lipid peroxidation, DNA damage and apoptosis in K562 cells. It increases the lipid peroxidation, DNA damage and apoptosis in K562 cells induced by PDT. The singlet oxygen quencher sodium azide suppresses the apoptosis induced by PDT with FORM. In conclusion, FORM consumption during PDT increase the effectiveness of cancer therapy on malignant cells. The effect of antioxidants on PDT maybe was determined by its sensitization ability to singlet oxygen.

Keywords: Formononetin; Reactive oxygen species; Photodynamic therapy; Sodium azide; Leukemia K562

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1. Introduction

Photodynamic therapy (PDT) is a treatment for cancer, and for certain non-cancerous diseases that are generally characterized by overgrowth of unwanted or abnormal cells [1]. Clinical trials continue to expand the role of PDT in cancer and in the treatment of localized microbial infections, as reviewed in [2, 3]. Protoporphyrin IX (PpIX) is an endogenous photosensitizer used for PDT [4], accumulated in the tumor tissue after the exogenous application of 5-aminolevulinic acid (ALA). Molecular mechanisms of photodynamic action result from generation of oxidative stress. In the presence of molecular oxygen, light of appropriate wavelength excites the photosensitizer that undergoes two types of reactions [5, 6]. In type 1 reactions, excited photosensitizer reacts directly with organic cellular substrates to form radical anions or radical cations that may react further with molecular oxygen to produce reactive oxygen species (ROS) [7, 8], resulting in target cell death either through necrosis or apoptosis [9]. In type 2 reactions, the excited photosensitizer transfers its energy directly to oxygen (already a triplet in its ground state) to form highly reactive but short-lived singlet oxygen (1O_2) [10, 11]. Both pathways can occur simultaneously and the ratio between them depends on the photosensitizer and the nature of the substrate. However, direct and indirect evidence supports a prevalent role for 1O_2 in the molecular processes initiated by PDT [3, 12].

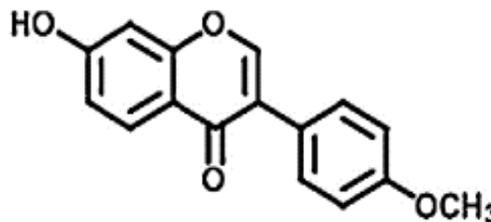


Fig.1. Chemical structures of FORM used in this study.

Consumption of fruits and vegetables is known to lower the risk of several diseases. Formononetin (FORM) [7-hydroxy-3(4-methoxyphenyl) chromone or 4-methoxy daidzein, Fig. 1] is a soy isoflavonoid that is found abundantly in traditional Chinese medicine *Astragalus mongholicus* (Bunge) and *Trifolium pratense* L. (red clover) [13]. They belong to the family Leguminosae. The extract of these herbs has been used clinically to treat different diseases including cardiovascular diseases in China for a long time [14, 15]. In addition, formononetin possesses hypolipidemic properties [16], mammary gland proliferation function [17] and antioxidative and estrogenic effects [18].

The mechanism of PDT exerted on tumor cell killing is the production of ROS. These oxidative actions may be modified by the phytochemicals present in food.

Although studies involving use of antioxidants during cancer therapy are promising, research on this topic is still scarce and controversial [19-21]. Studies indicate that supplementation with dietary antioxidants may improve the efficacy of radiation therapy by increasing tumor response and decreasing some of its toxicity on normal cells [21]. The other suggests that dietary antioxidants should not be used during radiation therapy, because they would protect cancer cells against radiation damage [23]. Each of these is based on different conceptual frameworks that are derived from results obtained from specific experimental designs, and thus, each may be correct within its parameters.

FORM has antioxidant properties and it was popular in diet and medicine [13-16]. It should be noticed that any antioxidant found to reduce toxicity of tumor therapy on healthy tissue has the potential to decrease effectiveness of cancer therapy on malignant cells. To assess whether FORM interferes with PDT treatment, the present study investigated the antioxidant activity of FORM and demonstrated the influence of FORM on PDT. This study could lead to a better understanding of their mechanisms of antioxidants applicability in PDT and found which food consumption during PDT did not decrease the effectiveness of cancer therapy.

2. Materials and Methods

2.1. Chemical reagents

L-glutamine, trypan blue, 5-aminolevulinic acids and FORM were purchased from Sigma-Aldrich Co., Ltd. (USA), while RPMI-1640 medium was from Gibco Co., Ltd. (USA). Dimethyl sulfoxides (DMSO), penicillin and streptomycin were obtained from Solarbio (China). Newborn calf serum (NCS) was from Sijiqing (China). Annexin V-FITC-PI Apoptosis Detection Kit was obtained from centre-Bio Co., Ltd. (China). All of the chemicals used in the present study were of analytical grade and purified water used throughout the experiments.

2.2. Equipments

Apoptosis was analyzed on a four-color fluorescence capability FACScalibur flow cytometer (BD Biosciences, USA). The fluorescence spectra were measured with the 970CRT fluorophotometer (Shanghai SANCO Instrument Co., Ltd, China). The fluorescence images of comet assay were examined with a fluorescence microscope (Olympus, Japan). The radiation source was a xenon lamp (USHIO, Japan).

2.3. Cell treatment

K562 cells were cultured in RPMI 1640 medium supplemented with 10% (v/v) newborn calf serum, 1% (v/v) L-glutamine and 100 units/ml antibiotics (penicillin and streptomycin) at 37 °C in 5% CO₂ and 95% air. FORM was dissolved in 100% DMSO and stored at -20 °C. For the cell growth assay, cells were seeded into six-well plates at a density of 1×10⁵ cells/ml and were treated with FORM or with DMSO only (as control) in triplicates. The final concentration of DMSO was kept at less than 0.05%. For the dose-dependent experiment, cells were treated with FORM for 24 h, respectively. For the time-dependent experiment, cells were treated with FORM and detected at 12, 24, 36 and 48 h.

2.4. Photodynamic treatment

Photodynamic treatment was carried out according to the previous report [24]. PDT was performed by the administration of ALA with light fluencies. Additional samples were treated with light only (light control). Dark control samples contained neither ALA nor light. K562 cells were stained in the dark at room temperature incubated for 4 h with 1mM ALA at 37 °C. Light device (xenon lamp, 400–800 nm) was then used for whole face irradiation at the light intensity of 350 mW/cm² and a light doses of 105 J/cm² at 37 °C. Cells were incubated with FORM for 20 min at 37 °C before PDT irradiation.

2.5. Cell viability assay

K562 cells (1×10⁵ cells/ml) were seeded into 6-well plates at 37 °C, 5% CO₂. After incubation, cells were stained by 0.2% trypan blue solution and monitored on a hemacytometer by a light microscopy. The percent of cell survival was calculated as follows:

$$\% \text{ Survival} = (\text{survival} / \text{control}) \times 100 \%$$

2.6. Determination of MDA

The measurement of lipid peroxidation was validated by the well-known TBA assay with some modifications as report of Jentsch *et al.* [25]. To assay the lipid peroxidation, the colored MDA-TBA adduct, formed in the reaction of MDA with TBA and acidic conditions was measured by a fluorophotometer, which was obtained and the fluorescence intensity at 546 nm was determined excited at 530 nm. The standards of

MDA were prepared by acid hydrolysis of 1,1,3,3-tetramethoxypropane. A calibration curve was used to calculate MDA concentration, and results were expressed in nanograms of MDA per 10^6 cells.

2.7. Flow cytometry

Cell apoptosis was assayed by using Annexin V-FITC-PI apoptosis detection kit from centre-Bio Co., Ltd. (China) according to the manufacturer's instructions. Cells (1×10^6 cells/ml) were washed with PBS and resuspended in binding buffer. Cells solution (400 μ l) was incubated with 10 μ l of Annexin V-FITC (20 μ g/ml) for 15 min and 10 μ l of PI (50 μ g/ml) for another 5 min at 4°C in the dark, and finally analyzed by flow cytometry at room temperature.

2.8. Comet assay

DNA damage was quantified by the comet assay as described previously [26]. Cell solution (3×10^5 cells/ml) mixed with same volume of 1% low melting point agarose in PBS was pipetted on ice precoated with 2% normal melting point agarose for 10 min. The slides were incubated in lysis buffer for other 1 h (0 °C) and then incubated in alkaline unwinding buffer for 20 min. Following electrophoresis for 20 min at 6.28 V in unwinding buffer, nuclei were stained with 0.02% (v/v) ethidium bromide for 20 min at room temperature. Digital images of 30 cells were randomly captured for analysis by Casp-2.2 analysis software. “% DNA in tail” was calculated as the extent of DNA damage.

2.9. Statistical analysis

Data were processed using EXCEL analysis software (Microsoft Co., Washington, USA) and expressed as means \pm SD. Differences between the groups were assessed by the two-tailed Student's t-test for unpaired samples. Statistical differences with $P < 0.05$ were considered significant.

3. Results

3.1. Cell survival

To analyze the effect of FORM on K562 cell growth/proliferation, we treated K562 cells cultures with different concentrations of FORM (0-100 μ M). For the cell growth assay, cells were seeded into six-well plates at a density of 1×10^5 cells/ml and were treated with FORM or with DMSO only (as control) in triplicates. The final concentration

of DMSO was kept at less than 0.05%. Cells were treated with FORM and detected at 12, 24, 36 and 48 h. Cells at each time interval were stained using 0.2 % trypan blue solution and viable cells were counted using a hemacytometer under light microscopy. Cell apoptosis was assayed by using Annexin V-FITC-PI apoptosis detection kit. As showed in Fig. 1A, FORM did not affect cell survival. After 48 h incubation, the viable cells in the culture treated with FORM were same as those in control ($P>0.1$). While the concentration of FORM was 100 μM , the highest concentration used in this study, also not affect cell survival ($P>0.1$). Cell apoptosis was examined by flow cytometry using Annexin V-FITC-PI Apoptosis Detection Kit. The result indicates that FORM also could not induce apoptosis in K562 cells 24h after incubation compare with control (Fig. 1B, $P>0.1$)

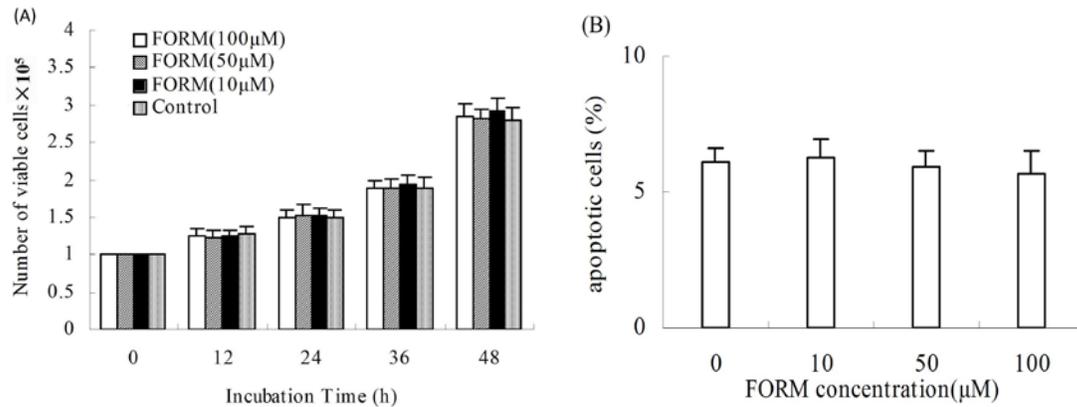


Fig.2. Effects of FORM on cell growth and apoptotic. Equal amounts of inoculants (1.0×10^5 viable cells) were seeded in the cultures containing different concentration of FORM. Cells at each time interval were counted. The data were assessed 24 h incubation. Calculation of apoptotic was by flow cytometry analysis. Results represent means $T \pm SD$ of three independent experiments.

3.2. PDT treatment

To investigate the effect of FORM on PDT we measured cell survival and cell apoptosis after FORM and PDT treatment. K562 cells in the exponential phase of growth were harvested at a density of 1×10^5 cells/ml. Cells were incubated for 4 h with 1mM ALA at 37 $^\circ\text{C}$. Then, cells were illuminated with a light intensity of 350 mW/cm^2 and a light doses of 105 J/cm^2 at 37 $^\circ\text{C}$. K562 cells were incubated with FORM for 20 min at 37 $^\circ\text{C}$ before PDT irradiation. The fraction of live cells in the control approach was set as 100%. Calculation of apoptotic was performed after staining the cells with Annexin

V-FITC-PI by flow cytometry analysis. The ALA added in the cell culture fluid which pH was 7.2. We know the pH of the ALA solution strongly depends on PpIX production in the cells [27]. The PpIX formation increases with the pH value of ALA. This pH=7.2 can ensure that PpIX production in the cells according to the previous report [24].

As shown in Fig. 3, incubation cell with light alone or ALA alone in the dark did not decrease cell viability. After cell with ALA and light, cell survival was significantly decreased. Approximately 35% cells survived in the PDT. In comparison with untreated cells (control) the increase in apoptosis (approximately 20%) was observed after PDT treatment. When cells were exposed to FORM and PDT, a dose-dependent decrease in cell survival was observed compared to cells exposed to PDT alone (Fig. 3, $P<0.05$). Cell apoptosis were examined by flow cytometry. In comparison with control, the increase in apoptosis (22.87%) was observed after ALA-PDT treatment. FORM enhanced the apoptosis induced by ALA-PDT in a dose-dependent manner (Fig. 3, $P<0.05$). When FORM supplemented together with the singlet oxygen quencher sodium azide (NaN_3 , 100 μM) under the same condition, FORM didn't enhanced the apoptosis in K562 cells induced by ALA-PDT (Fig. 4, $P>0.1$). In a word, under the same condition, compared with FORM supplemented alone (Fig. 3), FORM supplemented together with the sodium azide suppresses apoptosis (Fig. 4). Thus it can be seen that singlet oxygen quencher sodium azide suppresses apoptosis by the comparison of Fig. 3 and Fig. 4. FORM reduced the cell survival induced by ALA-PDT in a dose-dependent manner ($P<0.05$). When FORM supplemented together with the NaN_3 (100 μM) at same condition, FORM could not reduce the cell survival in K562 cells induced by ALA-PDT ($P>0.1$). It can be seen that singlet oxygen quencher sodium azide suppresses apoptosis and enhances cell survival by the comparison of Fig. 3 and Fig. 4.

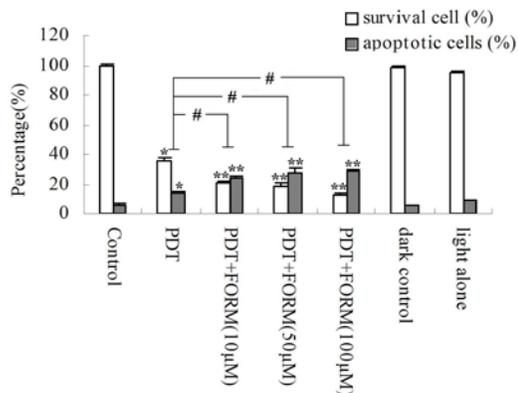


Fig.3. Effects of FORM on cell survival and apoptotic in PDT treatment. K562 cells were incubated 20 min prior PDT irradiation with different concentrations of FORM. The fraction of live cells in the control approach was set as 100%. Calculation of apoptotic was by flow cytometry analysis. Results represent means $T \pm SD$ of three independent experiments. # $p < 0.05$; * $p < 0.05$ vs. control; ** $p < 0.05$ vs. ALA-PDT.

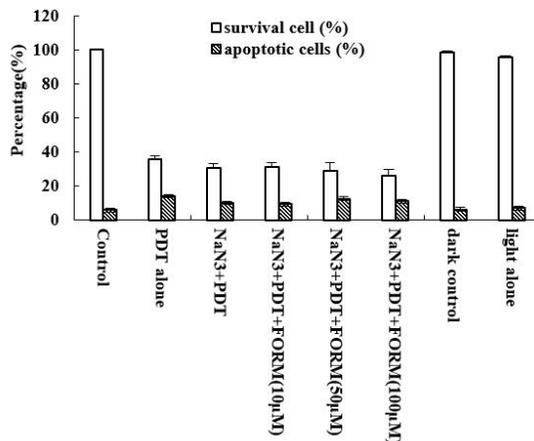


Fig.4. Effects of NaN_3 combined with FORM on apoptotic in PDT treatment. K562 cells were incubated 20 min prior ALA-PDT irradiation with different concentrations of FORM and NaN_3 . The fraction of live cells in the control approach was set as 100%. Calculation of apoptotic was by flow cytometry analysis. Results represent means $T \pm SD$ of three independent experiments.

3.3. Lipid peroxidation and DNA damage

In parallel with measurement of the influence of FORM and PDT on cell survival, we examined the Oxidative damage and DNA damage after treatments. Oxidative damage induced by PDT was investigated by measurement of malondialdehyde, a marker of lipid peroxidation. Peroxidation of lipids is particularly destructive because the formation of lipoperoxidation products leads to a facile propagation of free radicals and membrane disintegration. Comet assay used to evaluate the potential genotoxic effect induced by PDT on the cells. It has been widely used in toxicology, radiation biology, and was introduced into the field of PDT. Comet assay is a useful technique for the detection of DNA single- and double-stranded breaks, and alkali-labile sites in individual cells after treatment with genotoxins. The major advantages of the comet assay over other methods of measuring DNA damage is that information is acquired about the distribution of DNA damage and repair in individual cells within the population, providing an intracellular distribution of damage.

Result was assessed 24h after PDT irradiation. MDA produced was measured to evaluate the lipid peroxidation. MDA amount was determined by measuring the change

of fluorescence intensity by the fluorophotometry. DNA damage was quantified with the comet assay. “% DNA in tail” was calculated as the extent of DNA damage. K562 cells were incubated with FORM for 20 min at 37 °C before PDT irradiation. FORM supplementation alone did not affect the MDA concentration and percentage DNA in tail compared to control cells as shown in Fig. 5 ($P>0.1$). Significant increase in the amounts of MDA and level of DNA damage was found in K562 cells after PDT treatment compared with control cells or cells treated with ALA dark (dark control) or light alone (Fig. 6, $P<0.05$). FORM enhanced the lipid peroxidation and DNA damage in K562 cells induced by PDT (Fig. 6, $P<0.05$).

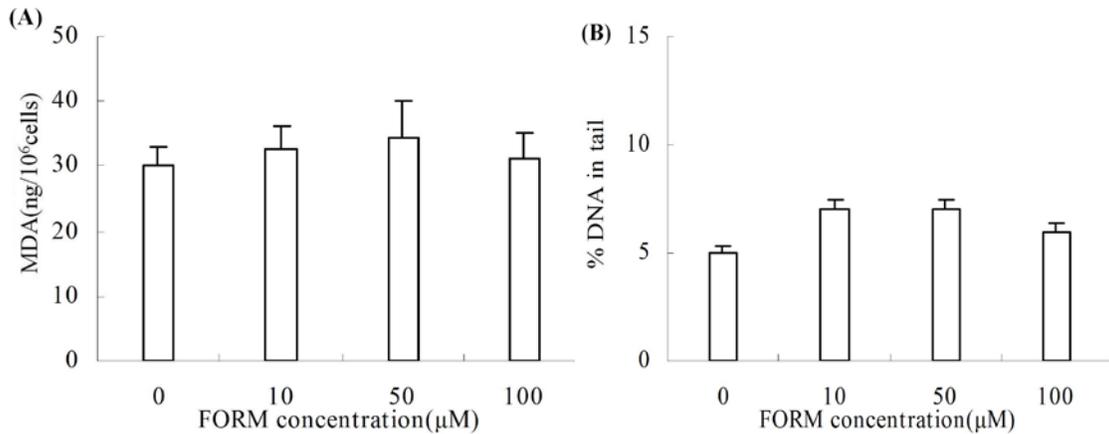


Fig.5. Influence of FORM on lipid peroxidation (A) and DNA damage (B) in K562 cells. Results represent means $T\pm SD$ of three independent experiments.

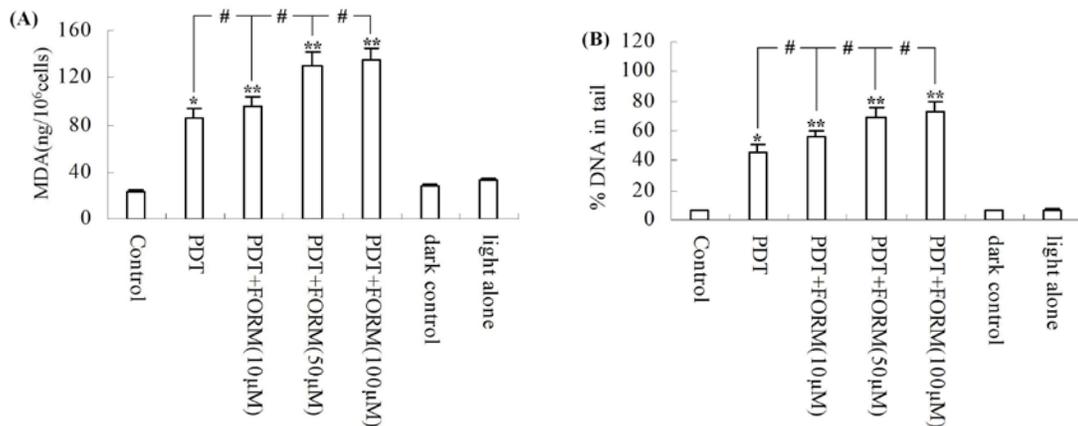


Fig.6. Influence of FORM on lipid peroxidation (A) and DNA damage (B) in K562 cells with PDT treatment. Results represent means $T\pm SD$ of three independent experiments. # $p < 0.05$; * $p < 0.05$ vs. control; ** $p < 0.05$ vs. ALA-PDT.

4. Discussion

The isoflavone FORM possesses antioxidant, antiproliferative, and ROS regulating activities, and have been shown to have protective effects against a number of diseases in humans, including cancer and heart disease [13-18]. The beneficial effects of FORM are credited to their antioxidant activities and inhibition of cellular mediators of cell death, protein kinases and eicosanoids has also been postulated [28]. The present study focuses on the effect of FORM on PDT in K562 cell. The cell line K562 which is derived from the patient with chronic myelogenous leukaemia (CML) is commonly employed as the “*in vitro*” model of the blast phase of this disease. PDT includes loading of the target cells with a photosensitizer and subsequent illumination with visible light. Because of cell need illumination with visible light at treatment, PDT is more convenient use in leukemia cell lines compare with solid cancer.

The use of antioxidants during cancer therapy is currently a debated topic because of some contradictory findings [19, 20, 29-35]. Some data indicate that antioxidants can protect healthy cells and tissues from the damage of free radical without affecting treatment efficacy [33]. On the other hand, other researchers suggested that antioxidant decrease the effectiveness of cancer therapy on malignant cells [34, 35]. Combining these results, further research on antioxidants and chemotherapy are now warranted. The role of antioxidants in PDT is only marginally examined. Few studies have been published to date concerning antioxidants in photodynamic treatment. One research data indicate that discovers that increasing intracellular concentrations of vitamin C contribute to the resistance of cultured cancer cells to prooxidant treatment modalities as PDT [29]. Other researchers found that high concentrations of vitamin E enhance the PDT action against HeLa cervical cancer cell line [30]; pre-treatment with tyrosine kinase inhibitor genistein may significantly improve the effectiveness of PDT with hypericin in MCF-7 and MDA-MB-231 breast cancer cells [32]; in PDT-treated cells a p38MAPK-regulated pathway coordinates the p62/NBR1-mediated clearance of cytosolic aggregates and mitigates PDT-induced proteotoxicity [36]. Our previous study [37] found the soyabean isoflavones genistein and daidzein did not decrease the effectiveness of cancer therapy on malignant cells. The daidzein was metabolized from FORM. Despite the various reports linking many of the beneficial properties of FORM to their antioxidant properties, no

comprehensive studies have been conducted the effect of FORM on PDT, which kill tumor cell by production of ROS. The aim of this study was to determine the effect of FORM on PDT in K562 cell.

FORM can likewise induce cancer cell death in different tumor cell type. Auyeung *et al.* [38] found FORM induces apoptosis in human colon cancer HCT 116 cells by activating caspases and down regulating Bcl-2 and Bcl-xL. Ye and colleagues [39] demonstrate that FORM also induces apoptosis in human prostate cancer cells by inactivating MAPK-Bax signaling pathway. Chen and colleagues [40] has reported that FORM inhibits the proliferation of MCF-7 cells and effectively induces cell cycle arrest in the G₀/G₁ phase by inactivating IGF1/PI3K/Akt pathways and decreasing cyclin D1 expression in human breast cancer cells in vitro. The same research group also found that FORM triggers Ras-p38 MAPK signaling pathway, and thus provokes apoptosis on estrogen-positive MCF-7 cells [41]. As shown in Fig. 1A, after 48 h of incubation, the viable cells in the cultures treated with FORM. It did not affect the proliferation of K562 cells. This was contradictory to previous reports [38] that FORM inhibits the growth of cancer cells. The opposite results may because the inhibit cancer cells growth effects of isoflavones is often challenging within the experimental model used due to important metabolic phenotype differences observed between cancer cells [42].

The present study found that FORM supplementation alone did not affect the cell apoptosis (Fig. 1B), but it increased the apoptosis in K562 cells induced by PDT (Fig. 3). FORM supplementation alone did not affect the MDA concentration and the level of DNA damage compared to control cells (Fig. 5), but it supplementation increased the amounts of MDA and the level of DNA damage in PDT treatment compared to PDT treatment alone (Fig. 6). These results imply that the direct effect of FORM may not contribute to the enhancement of PDT induced cytotoxicity in K562 cells and FORM exhibits a synergistic effect on PDT. The exact reasons for the FORM enhancement of PDT damage on cancer cells are unknown. Recently, FORM has been demonstrated to have effects on various cancer cells. Yu reports that FORM can use as an adjuvant to combine with epirubicin, a well-known anthracycline anticancer drug in HeLa cells to enhance the efficacy of epirubicin [43]. These results were similar to our findings that FORM enhanced the effectiveness of PDT.

FORM enhances the PDT-induced cytotoxicity in K562 cells are contradictory to the findings that it is an antioxidant that can scavenge ROS. The contradictory results may be caused by the result that FORM increases the amounts of singlet oxygen. It is known that although both superoxide anion and hydroxyl radical are potentially cytotoxic, most of the oxidative damage in PDT is caused by the singlet oxygen [3]. The different ROS have different signaling and damaging capabilities. Since singlet oxygen is highly reactive and cannot interconvert with endogenous ROS species, it is more likely to cause damage than to elicit signal transduction. Yu found that FORM increased ROS production (H_2O_2 and $^1\text{O}_2$) but decreased MRP1 and MRP2 expressions, thus supporting the negative correlation between ROS levels and MDR transporter expression [43]. However, there is no study on the relationship among the effects of FORM on ROS levels, MDR transporter expression, and apoptosis modulation. Moreover, in this study, the singlet oxygen quencher sodium azide suppresses apoptosis (Fig 4) in K562 cells induced by PDT with FORM support this hypothesis. These results imply that singlet oxygen could be involved in the enhance cytotoxicity effects of the FORM in PDT.

The finding of singlet oxygen involved in the enhance cytotoxicity effects of the FORM in PDT implied that the contradicted findings on the use of antioxidants during cancer therapy may be resulted from the different sensitization activity of antioxidants on singlet oxygen. The antioxidants which can scavenge singlet oxygen may decrease the effectiveness of PDT (such as vitamin C [29]). The antioxidants which increase the amounts of singlet oxygen may exhibit a synergistic effect on PDT (such as genistein [37], FORM (in this research)).

In conclusion, the results proved that FORM enhanced the cell death in K562 cells induced by PDT. FORM supplementation alone did not affect the lipid peroxidation, DNA damage and apoptosis in K562 cells. It increases the lipid peroxidation, DNA damage, and apoptosis in K562 cells induced by PDT. The singlet oxygen quencher sodium azide suppresses apoptosis in K562 cells induced by FORM in PDT. Consequently, these results implied that FORM consumption during PDT did not decrease the effectiveness of cancer therapy on malignant cells and singlet oxygen could be involved in this process. This study could lead to a better understanding of their mechanisms of antioxidants applicability in PDT. Further studies could found the effect

of antioxidants during cancer therapy to guide the patient's diet.

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References

- [1] Milla S.L., Rodriguez M.E., Cogno I.S., Rumie N.B. (2012): Vittar, M.F. Pansa, M.J. Lamberti, V.A. Rivarola, Direct and indirect photodynamic therapy effects on the cellular and molecular components of the tumor microenvironment, *Biochim. Biophys. Acta.* **1835**, 36-45.
- [2] Brown S.B., Brown E.A., Walker I. (2000): The present and future role of photodynamic therapy in cancer treatment, *Lancet. Oncol.* **5(8)** 497-508.
- [3] Agostinis P., Berg K., Cengel K.A., Foster T.H., Girotti A.W., Gollnick S.O., Hahn S.M., Hamblin M.R., Juzeniene A., Kessel D., Korbelik M., Moan J., Mroz P., Nowis D., Piette J., Wilson B.C., Golab J. (2011): Photodynamic therapy of cancer: an update, *CA. Cancer. J. Clin.* **61** 250-281.
- [4] Kennedy J.C., Pottier R.H. (1992): Endogenous protoporphyrin IX, a clinically useful photosensitizer for photodynamic therapy, *J Photochem. Photobiol. B.* **14** 275-92.
- [5] Garland M.J., Cassidy C.M., Woolfson D., Donnelly R.F. (2009): Designing photosensitizers for photodynamic therapy: strategies, challenges and promising developments, *Future. Med. Chem.* **1(4)** 667-691.
- [6] Castano A.P., Pawel M., Hamblin M.R. (2006): Photodynamic therapy and anti-tumor immunity, *Nat. Rev. Cancer.* **6(7)** 535-545.
- [7] Ion R.M. (2010): Derivative UV-Vis spectrophotometry for porphyrins interactions in photodynamic therapy, *Anal. Lett.* **43(7)** 1277-1286.
- [8] Glasauer A., Chandel N.S. (2014): Targeting antioxidants for cancer therapy, *Biochem. Pharmacol.* **92(1)** 90-101.
- [9] Woods J.A., Traynor N.J., Brancalion L., Moseley H. (2004): The Effect of Photofrin on DNA Strand Breaks and Base Oxidation in HaCaT Keratinocytes: A Comet Assay Study, *Photochem. Photobiol.* **79(1)** 105-113.
- [10] Castano A.P., Demidova T.N., Hamblin M.R. (2005): Mechanisms in photodynamic therapy: part two-cellular signaling, cell metabolism and modes of cell death, *Photodiagn. Photodyn. Ther.* **2(1)** 1-23.
- [11] Ogilby P.R. (2010): Singlet oxygen: there is indeed some thing new under the sun, *Chem. Soc. Rev.* **39(8)** 3181-3209.
- [12] Niedre M., Patterson M.S., Wilson B.C. (2002): Direct near-infrared luminescence detection of singlet oxygen generated by photodynamic therapy in cells in vitro and tissues in vivo, *Photochem. Photobiol.* **75(4)** 382-391.
- [13] Wu J.H., Li Q., Wu M. Y., Guo D.J., Chen H.L., Chen S.L., Seto S.W., Au A.L.S., Poon C.C.W., Leung G.P.H., Lee S.M.Y., Kwan Y.W., Chan S.W. (2010): Formononetin, an isoflavone, relaxes rat isolated aorta through endothelium-dependent and endotheliumin dependent pathways. *J. Nutr Biochem.* **21(7)** 613-620
- [14] Wang Y.P., Li X.Y., Song C.Q., Hu Z.B. (2002): Effect of astragaloside IV on T, B lymphocyte proliferation and peritoneal macrophage function in mice, *Acta. Pharmacol. Sin.* **23** 263-266.
- [15] Lei H., Wang B., Li W.P., Yang Y., Zhou A.W., Chen M.Z. (2003): Anti-ageing effect of astragalosides and its mechanism of action, *Acta. Pharmacol. Sin.* **24** 230-234.
- [16] Siddiqui M.T., Siddiqui M. (1976): Hypolipidemic principles of *Cicer arietinum*: biochanin-A and formononetin. *Lipids.* **11** 243-246.
- [17] Wang W., Tanaka Y., Han Z., Higuchi C.M. (1995): Proliferative response of mammary glandular tissue to formononetin. *Nutr. Cancer.* **23** 131-140.
- [18] Mu H., Bai Y.H., Wang S.T., Zhu Z.M., Zhang Y.W. (2009): Research on antioxidant effects and estrogenic effect of formononetin from *Trifolium pratense* (red clover). *Phytomedicine.* **16(4)** 314-319
- [19] González M.J., Miranda-Massari J.R., Mora E.M., Guzmán A., Riordan N.H., Riordan H.D., Casciari J.J., Jackson J.A., Román-Franco A. (2005): Orthomolecular oncology review: ascorbic acid and cancer 25 years later, *Integr. Cancer. Ther.* **4** 32-44.
- [20] Gonzalez M.J., Miranda-Massari J.R. (2006): Advances in vitamin C research, *Integr. Cancer. Ther.* **5(1)** 7-8.
- [21] Ozben T. (2007): Oxidative stress and apoptosis: impact on cancer therapy, *J. Pharm. Sci.* **96(6)** 2181-2196.

- [22] Prasad K., Cole W., Kumar B., Prasad K. (2001): Scientific rationale for using high-dose multiple micronutrients as an adjunct to standard and experimental therapies, *J. Am. Coll. Nutr.* **20(5)** 450S-463S.
- [23] Salganik R.I. (2001): The benefits and hazards of antioxidants: controlling apoptosis and other protective mechanisms in cancer patients and the human population, *J. Am. Coll. Nutr.* **20(5)** 464S-472S.
- [24] Zhang S.J., Zhang Z.X. (2004): 5-Aminolevulinic acid based photodynamic therapy in leukemia cell HL60, *Photochem. Photobiol.* **6** 545-50
- [25] Jentsch A.M., Bachmann H., Fürst P., Biesalski H.K. (1996): Improved analysis of malondialdehyde in human body fluids, *Free. Radical. Bio. Med.* **20(2)** 251-256.
- [26] Singh N.P., McCoy M.T., Tice R.R., Schneider E.L. (1988): A single technique for quantification of low levels of DNA damage in individual cell, *Exp. Cell. Res.* **175(1)** 184-191.
- [27] Krammer B., Uberriegler K. (1996): In-vitro investigation of ALA-induced protoporphyrin IX, *J. Photochem. Photobiol. B.* **36** 121-126
- [28] Gopalakrishna R., Gandimeda U. (2002): Antioxidant regulation of protein kinase C in cancer prevention, *J. Nutr.* **132(12)** 3819-3823.
- [29] Frank J., Flaccus A., Schwarz C. (2006): Ascorbic acid suppresses cell death in rat DS-sarcoma cancer cells induced by 5-aminolevulinic acid-based photodynamic therapy, *Free. Radical. Bio. Med.* **40(5)** 827-836.
- [30] Al-Sherbini A.M., ElNoury A. H., ElRouby M.N., Ibrahim T. (2009): VitaminE (a-tocopherol) enhances the PDT action of hematoporphyrin derivatives on cervical cancer cells, *Med. Laser. Appl.* **24(1)** 65-73.
- [31] Sun D., Zhang S.J., Wei Y.F., Yin L.F. (2009): Study on antioxidants' activities of Mangostin and its effect to K562 leukemia cell in photodynamic therapy, *Acta. Bioch. Bioph. Sin.* **41(12)** 1033-1043.
- [32] Ferenc P., Solár P., Kleban J. (2010): Down-regulation of Bcl-2 and Akt induced by combination of photoactivated hypericin and genistein in human breast cancer cells, *J. Photochem. Photobiol. B.* **98(1)** 25-34.
- [33] Akbas H.S., Timur M., Ozben T. (2006): Concurrent use of antioxidants in cancer therapy: an update, *Expert, Rev. Clin. Immunol.* **2(6)** 931-939.
- [34] Richardson M.A., Sanders T., Palmer J.L., Greisinger A., Singletary S.E. (2000): Complementary/alternative medicine use in a comprehensive cancer center and the implications for oncology, *J. Clin. Oncol.* **18(13)** 2505-2514.
- [35] Seifried H.E., Nald S.S., Anderson D.E., Peter G., Milner J.A. (2003): The antioxidant conundrum in cancer, *Cancer. Res.* **63(15)** 4295-4298.
- [36] Rubio N., Verrax J., Dewaele M., Verfaillie T., Johansen T., Piette J., Agostinis P. (2014): p38MAPK-regulated induction of p62 and NBR1 after photodynamic therapy promotes autophagic clearance of ubiquitin aggregates and reduces reactive oxygen species levels by supporting Nrf2-antioxidant signaling, *Free. Radical. Bio. Med.* **67** 292-303.
- [37] Zhang S.j., Sun D., Hao J.b., Wei Y.F., Yin L.f., Liu X. (2012): The effect of dietary soyabean isoflavones on photodynamic therapy in K562 leukemia cells, *J. Photochem. Photobiol. B.* **110** 28-33.
- [38] Auyeung K.K., Ko J.K. (2010): Novel herbal flavonoids promote apoptosis but differentially induce cell cycle arrest in human colon cancer cell, *Invest. New. Drugs.* **28** 1-13.
- [39] Ye Y., Hou R., Chen J., Mo L., Zhang J., Huang Y., Mo Z. (2012): Formononetin-induced apoptosis of human prostate cancer cells through ERK1/2 mitogen-activated protein kinase inactivation, *Horm. Metab. Res.* **44** 263-267.
- [40] Chen J., Zeng J., Xin M., Huang W., Chen X. (2011): Formononetin induces cell cycle arrest of human breast cancer cells via IGF1/PI3K/Akt pathways in vitro and in vivo, *Horm. Metab. Res.* **43** 681-686.
- [41] Chen J., Sun L. (2012): Formononetin-induced apoptosis by activation of Ras/p38 mitogen-activated protein kinase in estrogen receptor-positive human breast cancer cells, *Horm. Metab. Res.* **44** 943-948.
- [42] Gu L., House S.E., Prior R.L., Fang N., Ronis M.J., Clarkson T.B. (2006): Metabolic phenotype

of isoflavones differ among female rats, pigs, monkeys, and women, *J. Nutr.* **136** 1215-21.
[43] Lo Y.L., Wang W.J. (2013): Formononetin potentiates epirubicin-induced apoptosis via ROS production in HeLa cells in vitro, *Chem-Biol. Interact.* **205** 188-197.

TABLE AND FIGURE CAPTIONS

Fig.1. Chemical structures of FORM used in this study.

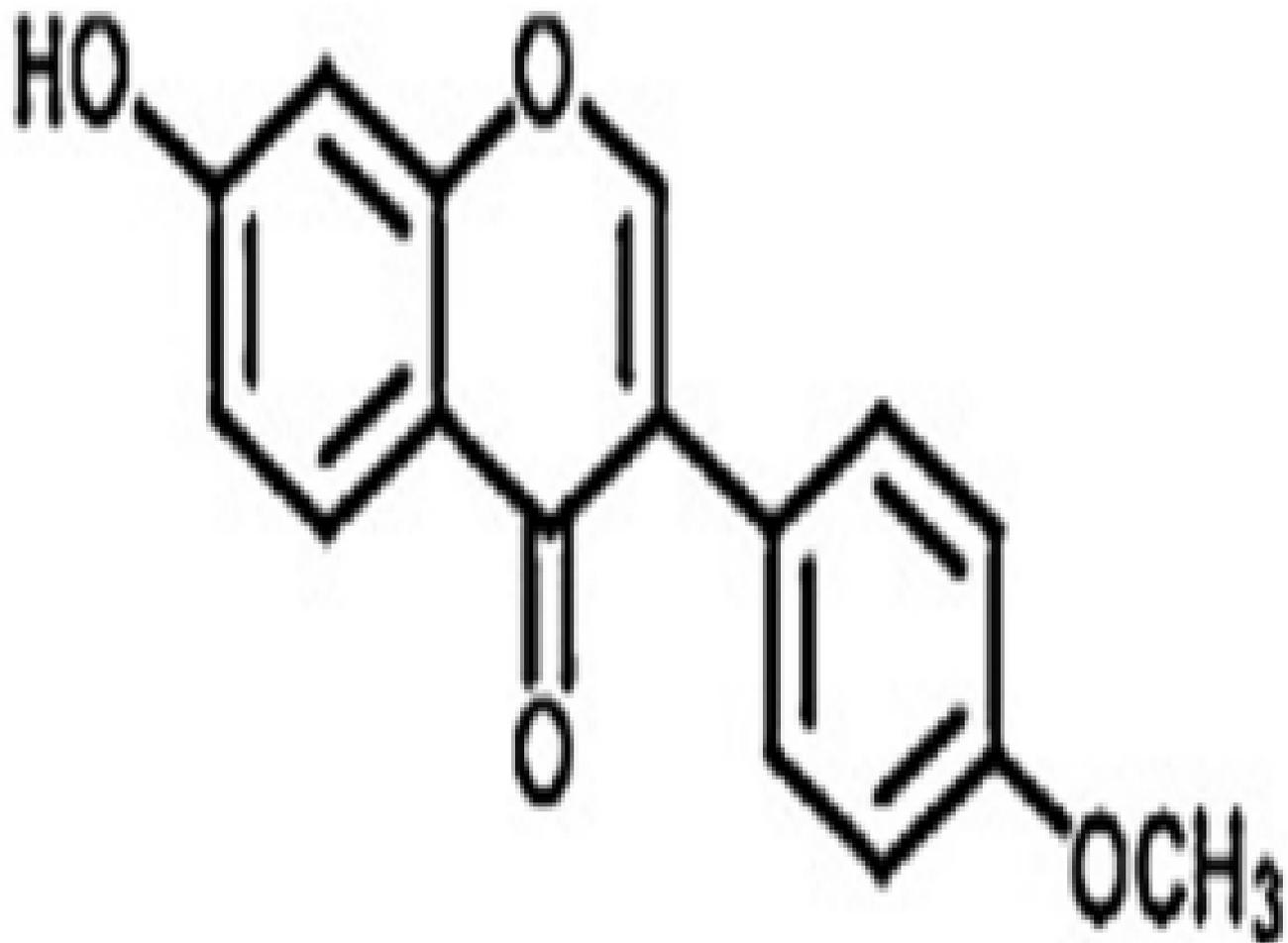
Fig.2. Effects of FORM on cell growth and apoptotic. Equal amounts of inoculants (1.0×10^5 viable cells) were seeded in the cultures containing different concentration of FORM. Cells at each time interval were counted. The data were assessed 24 h incubation. Calculation of apoptotic was by flow cytometry analysis. Results represent means $T \pm SD$ of three independent experiments.

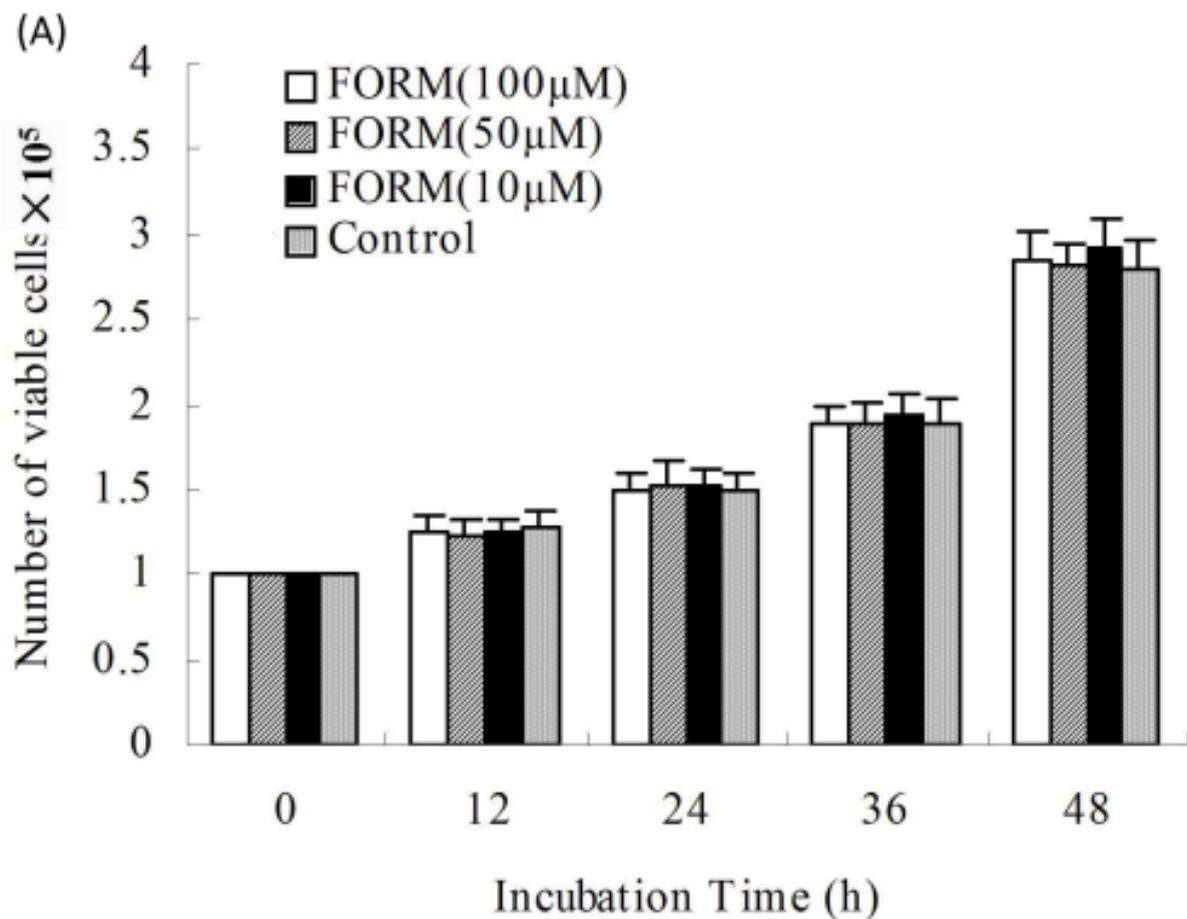
Fig.3. Effects of FORM on cell survival and apoptotic in PDT treatment. K562 cells were incubated 20 min prior PDT irradiation with different concentrations of FORM. The fraction of live cells in the control approach was set as 100%. Calculation of apoptotic was by flow cytometry analysis. Results represent means $T \pm SD$ of three independent experiments. # $p < 0.05$; * $p < 0.05$ vs. control; ** $p < 0.05$ vs. ALA-PDT.

Fig.4. Effects of NaN_3 combined with FORM on apoptotic in PDT treatment. K562 cells were incubated 20 min prior ALA-PDT irradiation with different concentrations of FORM and NaN_3 . The fraction of live cells in the control approach was set as 100%. Calculation of apoptotic was by flow cytometry analysis. Results represent means $T \pm SD$ of three independent experiments.

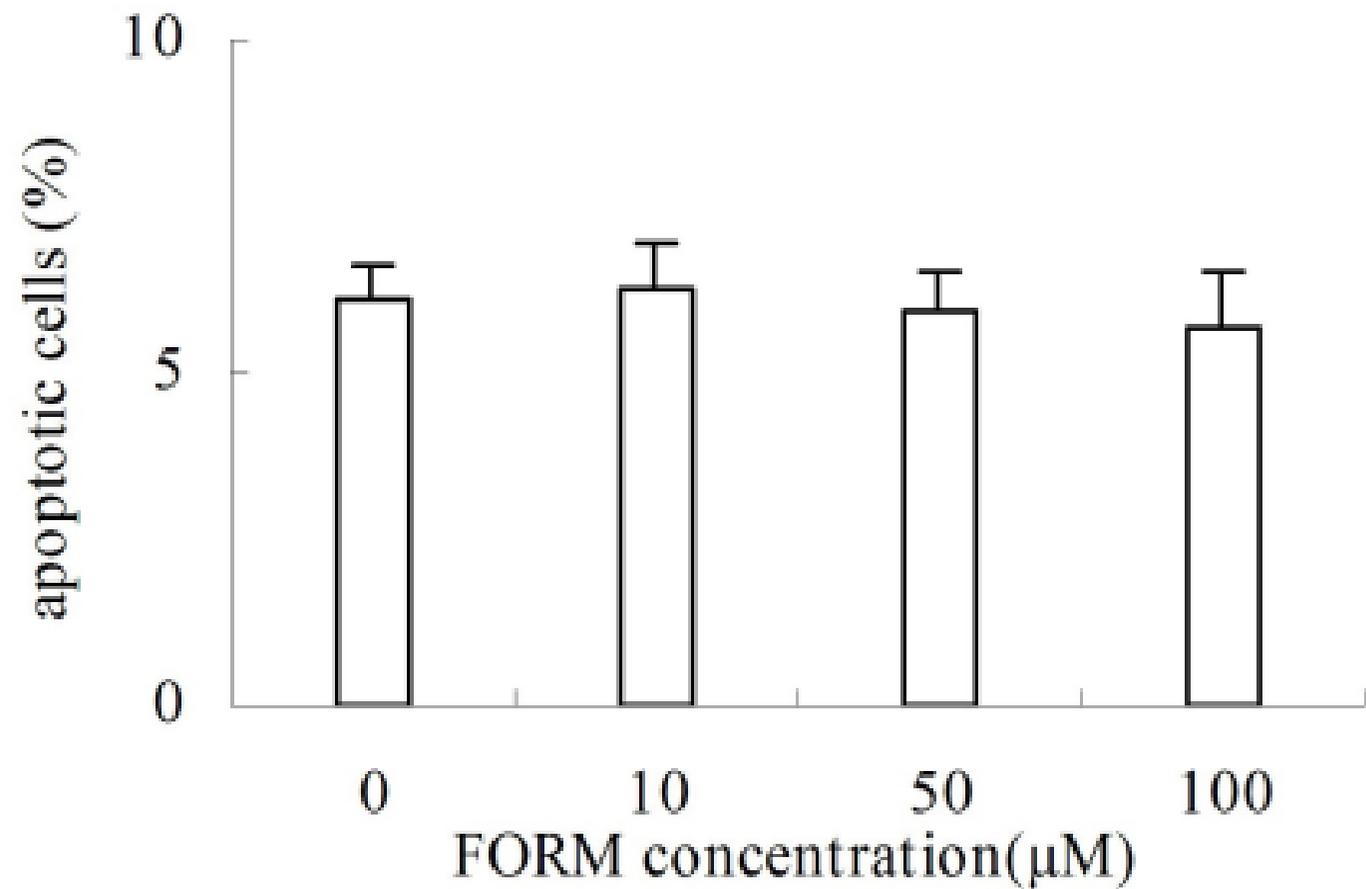
Fig.5. Influence of FORM on lipid peroxidation (A) and DNA damage (B) in K562 cells. Results represent means $T \pm SD$ of three independent experiments.

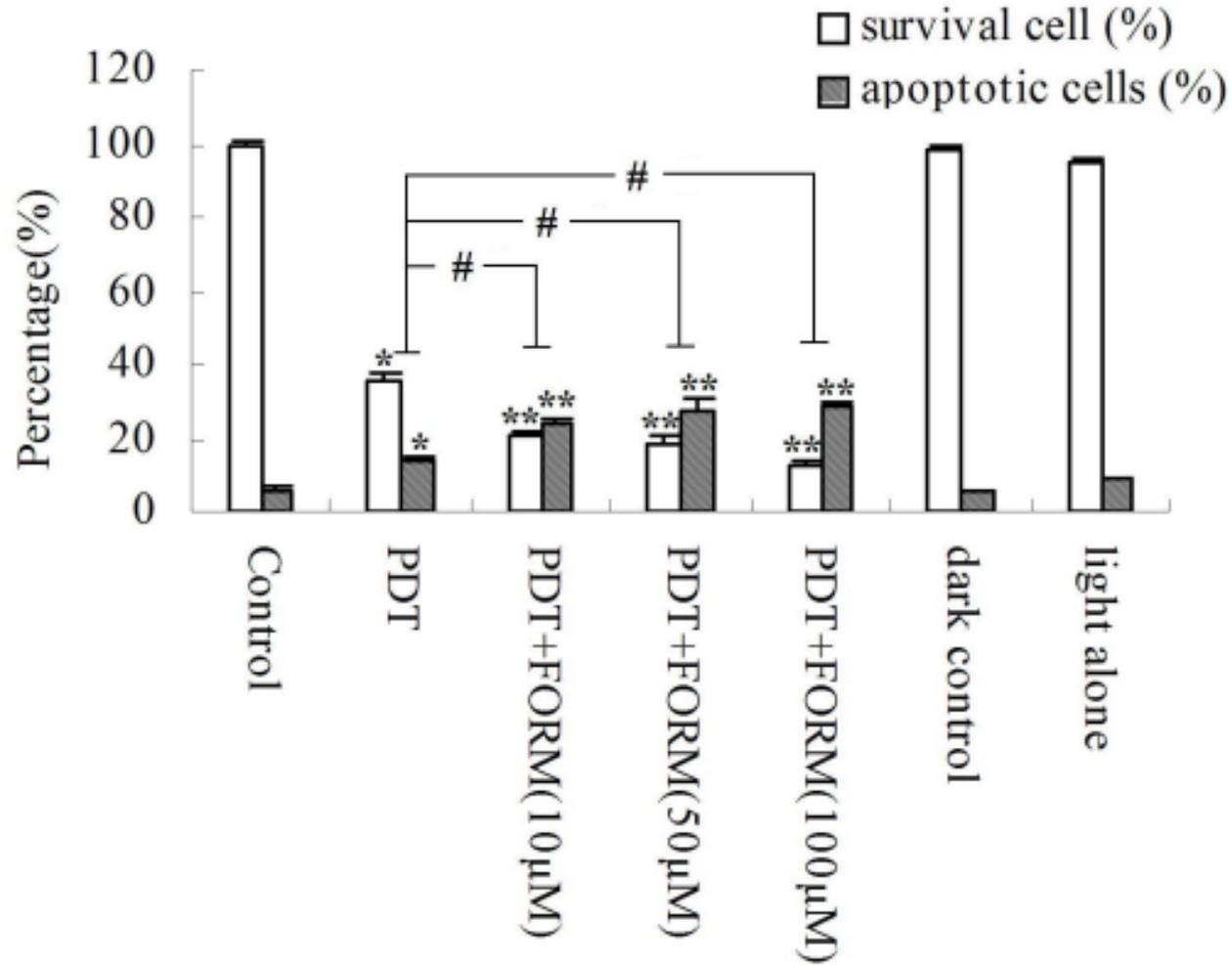
Fig.6. Influence of FORM on lipid peroxidation (A) and DNA damage (B) in K562 cells with PDT treatment. Results represent means $T \pm SD$ of three independent experiments. # $p < 0.05$; * $p < 0.05$ vs. control; ** $p < 0.05$ vs. ALA-PDT.

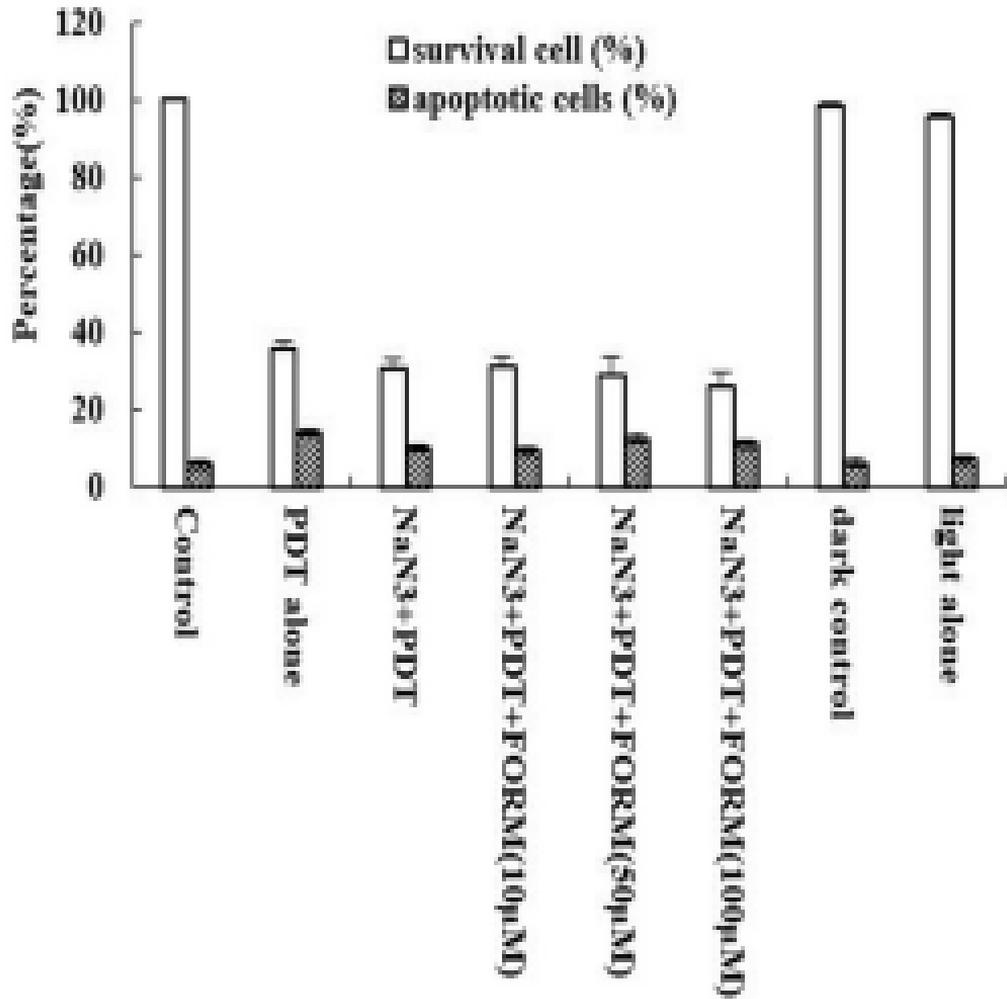




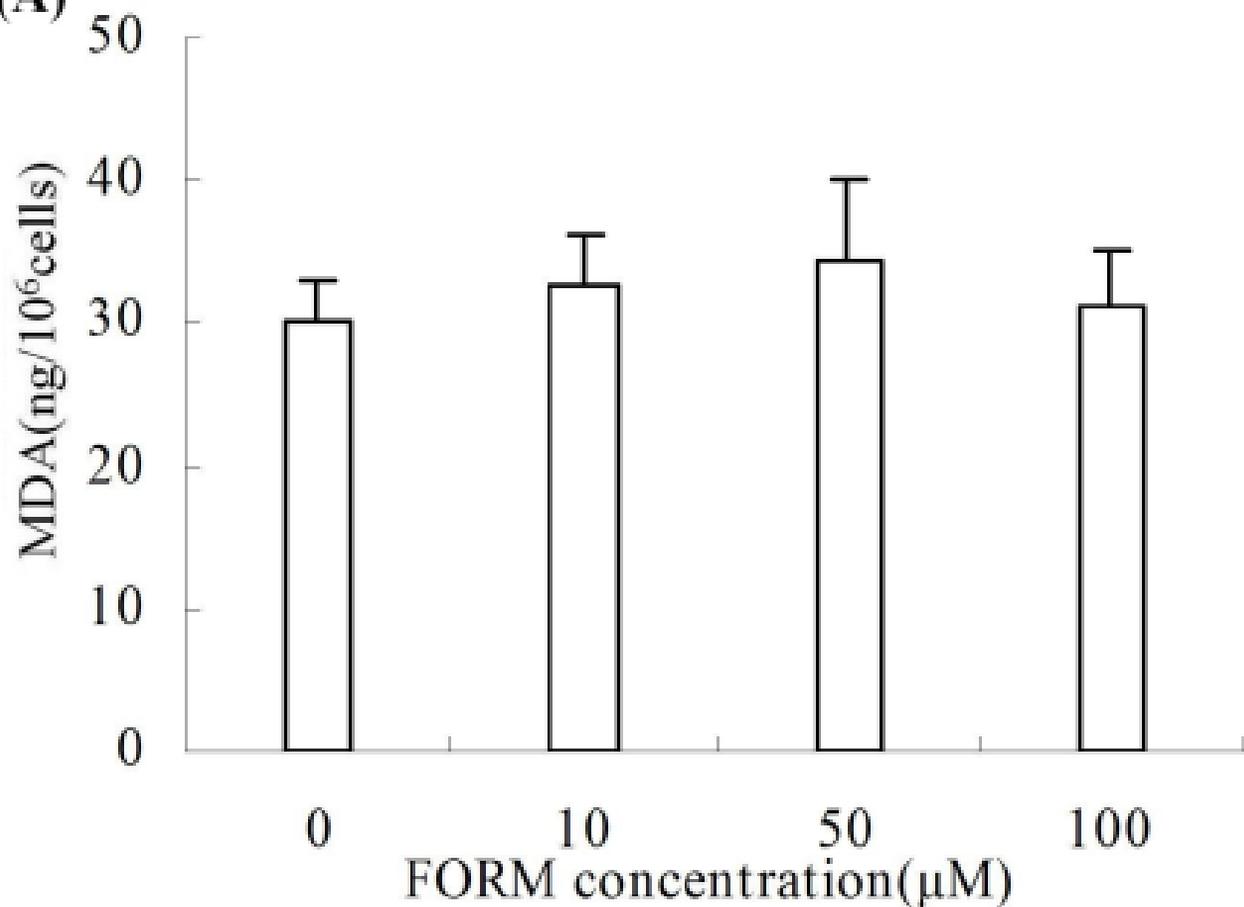
(B)

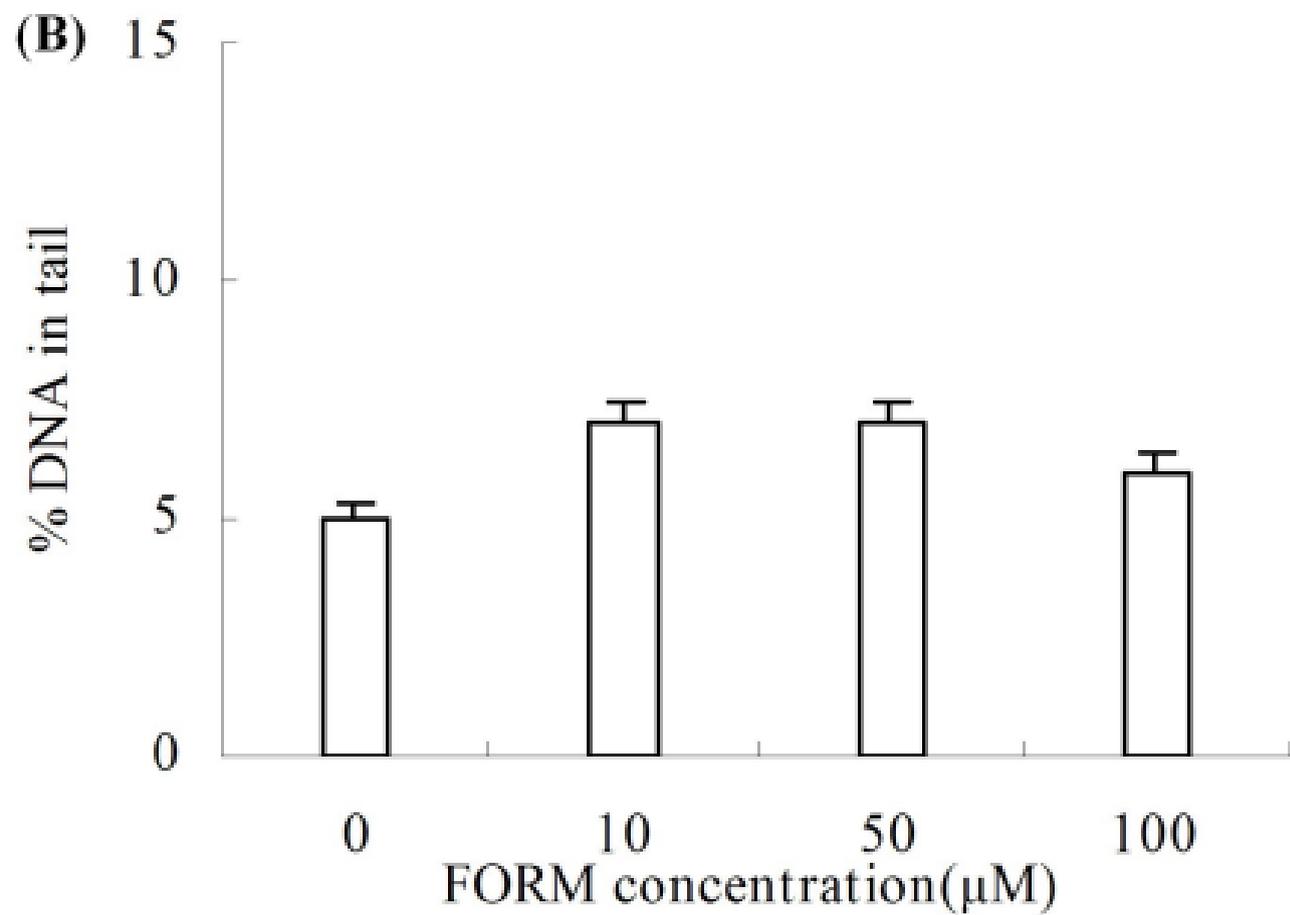






(A)





(A)