

**Title: Infrared spectroscopic investigation on erythrocyte membrane-smoke interactions due to chronic cigarette smoking**

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**Abstract**

Cigarette smoking is a serious health problem throughout the world, with a complicated and not totally clear bio-effect. In this study, erythrocytes were obtained from healthy male volunteers aged  $22\pm 2$  years and, the possible effects of three cigarette smoking rates namely 10, 15 and 20 cigarette/day on erythrocytes membrane characteristics were examined by Fourier transform infrared spectroscopy (FTIR). The results of this study indicate many smoking-dependent variations on erythrocytes membrane without an obvious dose-response relationship. There was disruption in the acyl chain packing; changes in membrane order and phases as well as membrane proteins becoming more folded. These physico-chemical changes should have an impact on the function of erythrocytes and may explain the complex interaction of cigarette smoke mainstream with erythrocyte membrane and to some extent clarify the pathological processes associated with cigarette smoking.

Keywords: Cigarette smoke; Erythrocyte membrane; FTIR; Protein; Lipid

**Changelog**

Dear Editor

The authors of this manuscript took all the comments and suggestions of both reviewers 1 and 2 on board and we made all the required corrections. Also, we added two citations as both reviewers ask for. All these changes are yellow-highlighted throughout the manuscript. Finally, the reference list is also updated.

Thank you for your efforts with this manuscript

Corresponding author

Sherif S. Mahmoud

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1 **Infrared spectroscopic investigation on erythrocyte membrane-smoke interactions due**  
2 **to chronic cigarette smoking**

3

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14

15 **Abstract**

16 Cigarette smoking is a serious health problem throughout the world, with a complicated and  
17 not totally clear bio-effect. In this study, erythrocytes were obtained from healthy male  
18 volunteers aged 22±2 years and, the possible effects of three cigarette smoking rates namely  
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26 clarify the pathological processes associated with cigarette smoking.

27 **Key words:** Cigarette smoke, Erythrocyte membrane, FTIR, Protein, Lipid.

28

## 29 **Introduction**

30 **Cigarette smoking - a worldwide social habit** - is associated with well documented numerous  
31 health hazards that include lung cancer, stroke, respiratory, cardiovascular diseases and  
32 osteoporosis (Mello 2010). These health risks result from the entrance of the mainstream  
33 smoke into the blood which leads to alterations in the blood constituents, plasma,  
34 **erythrocytes**, platelets and **leukocytes** (Freeman et al. 2005), and have been attributed to the  
35 abundance of reactive oxygen species and reactive nitrogen species (Barua et al. 2003). More  
36 smoking health hazards that should be addressed are the ocular ones aggravated by cigarette  
37 smoke which include dry eye (**Arffa** 1991), conjunctival irritation, age-related macular  
38 degeneration, cataract, ocular inflammation and change in tear characteristics (**Satici** et al.  
39 2003, Sprague et al. 2006, Thorne et al. 2008, Roesel et al. **2011**, Lin et al. 2010). In addition,  
40 the retina is known to have the highest rate of oxygen consumption of any organ in the body  
41 (**Kewal 2011**). Therefore any alterations in the structure and/or function of the erythrocyte  
42 membranes should have an impact on retinal function.

43 Padmavathi et al. (2010) found that chronic cigarette smoking alters the content of individual  
44 phospholipids of erythrocytes membrane that was associated with increased membrane lipid  
45 peroxidation. The erythrocytes membrane-cigarette smoke interactions appear to be more  
46 complex and the exact mechanism that enhance these effects remain uncertain. Therefore, the  
47 present study is aimed to understand the causation of chronic cigarette smoke on the  
48 erythrocyte membrane structural and conformational characteristics that were studied by  
49 Fourier transform infrared spectroscopy (**FTIR**) while **taking** into account the smoking rate  
50 (number of cigarette smoked/day) and the dose-response relationship if any.

## 51 **Participants and Methods**

52 **Subjects**

53 Four groups of healthy male subjects were involved in this study, where each group  
54 composed of 25 age-matched subjects ( $22\pm 2$  years). Three groups were cigarette smokers for  
55  $3\pm 1$  years but differ in the daily smoking rate as 10, 15 and 20 cigarette/day, and the last  
56 group served as the control one. All study subjects had normal renal and hepatic functions  
57 with no evidence of any acute infection or active inflammatory process. None of the subjects  
58 had hypertension, were taking any medications or receiving vitamin supplementation. There  
59 was no significant obesity was observed between all subjects ( $BMI=22.4\pm 3.6$ ), as well as  
60 cholesterol ( $155\pm 15$  mg/dl) and triglycerides ( $125\pm 8$  mg/dl), with normal fasting blood  
61 glycemic level ( $90\pm 5$  mg/dl). All subjects signed an informed consent with tenets of the  
62 Declaration of Helsinki.

#### 63 Preparation of erythrocyte membranes

64 Blood was obtained by veinpuncture, collected into heparinized syringes and immediately  
65 centrifuged at 500 g at  $4^{\circ}\text{C}$  for 10 min. The plasma and buffy coat were discarded, and  
66 erythrocytes were re-suspended and washed three times in physiological saline (0.9% NaCl).  
67 Erythrocyte membranes were prepared as previously mentioned by Sprague et al. 2006. That  
68 is washed packed erythrocytes were added to 200 ml of hypotonic buffer ( $5\text{ mmol/l}$  Tris-HCl,  
69  $2\text{ mmol/}$ EDTA, pH 7.4) and stirred vigorously for 20 min. The mixture was centrifuged at  
70 23,000 g for 15 min, and the supernatant was discarded. After repeating the last step one  
71 more time, membranes were pooled, resuspended in buffer and centrifuged. Finally, after  
72 discarding the supernatant, membranes were freeze-dried and kept under  $\text{N}_2$  gas atmosphere  
73 for further analysis.

#### 74 Fourier transform infrared spectroscopy

75 Erythrocytes membrane was mixed with finely ground powder KBr to prepare the KBr-disks  
76 that were used in analysis. This mixture was placed in a special holder provided by the  
77 manufacturer and pressed under vacuum at 3 tons (13.8 MPa) for 2 min to produce IR-KBr

78 disks (Sherif 2010). FTIR measurements were carried out using Nicolet-iS5 infrared  
79 spectrometer (ThermoFisher Scientific Inc, Madison,USA) with effective resolution of  $2\text{ cm}^{-1}$   
80 <sup>1</sup>. Each spectrum was derived from 100 sample interferograms. The spectrometer was  
81 operated under a continuous dry N<sub>2</sub> gas purge to remove interference from atmospheric CO<sub>2</sub>  
82 and H<sub>2</sub>O vapor. The spectra were baseline corrected and then smoothed with Savitsky–Golay  
83 to remove the noise before Fourier transformation. Three spectra from each sample were  
84 obtained and averaged using OriginPro 7.5 software to obtain the final average group  
85 spectrum that was normalized and analyzed for the following spectral regions: 4000-3000  $\text{cm}^{-1}$   
86 <sup>1</sup> (NH-OH region), 3000-2800  $\text{cm}^{-1}$  (CH stretching region), 1800-1600  $\text{cm}^{-1}$  (amide I region)  
87 and 1600 -900  $\text{cm}^{-1}$  (fingerprint region), and then subjected to curve enhancement analysis  
88 using a combination of Fourier deconvolution and non-linear curve fitting. The second  
89 derivative of the group spectrum was employed to confirm the number of the estimated  
90 peaks.

#### 91 Statistical analysis

92 Results were calculated and expressed as mean±standard deviation (SD). Comparison  
93 between multiple groups was performed using analysis of variance (ANOVA); commercially  
94 available statistical software package (SPSS-11 for **Windows**) was used where the  
95 significance level was set at  $p<0.05$ . All spectral analyses were performed with OriginPro 7.5  
96 software package (Origin Lab Corporation, Northampton, MA, USA).

#### 97 **Results**

98 The erythrocytes infrared spectra were analyzed according to the following regions:

#### 99 NH-OH region

100 In figure (1), the contour of the control erythrocytes membrane indicates the presence of  
101 several absorption bands that were resolved into 8 structural components using the curve  
102 enhancement procedure as given in table (1). In the same context, the contour of smokers'

103 erythrocytes membrane was also resolved into 8 structural peaks that differed in band  
104 position, band area and bandwidth as well as their assignment (table 1). The stretching OH  
105 (<sub>str</sub>OH) compositional bands were reduced from four components in the control and 10  
106 cig/day groups to three structural components as the number of smoked cigarettes increased  
107 to 15/20 daily.

108 The two structural components of the asymmetric OH (<sub>asym</sub>OH) bond were reduced to one  
109 component in two groups only; 10 cig/day and 20 cig/day that were characterized by  
110 significantly increased bandwidth and band area. At a smoking rate of 15 cig/day, the two  
111 detected structural component show paradox characteristics; where the higher frequency band  
112 has an increased band area and bandwidth while the lower frequency one has decreased band  
113 area and bandwidth. On the other hand, symmetric OH (<sub>sym</sub>OH) band showed a different  
114 behavior; increased vibrational frequency and band area that were associated to different  
115 smoking rates, this is in addition to its splitting into 2 structural components at a daily  
116 smoking rate of 10 and 20 cigarette compared to control one as shown in table 1.

117 For all smoking groups, the symmetric NH (<sub>sym</sub>NH) stretching vibrational mode was  
118 detected. Comparing the three smoking groups, it is noticed that this vibrational mode was  
119 characterized by increased vibrational frequency, band area as well as bandwidth as the  
120 number of cigarette smoked increased from 10 to 15/20 daily while, the asymmetric  
121 vibrational mode of the NH band (<sub>asym</sub>NH) was restricted at 10 cig/day smoking group, then  
122 detected again as the smoking rate increased to 15/20 daily and is characterized by increased  
123 vibrational frequency and bandwidth as well. The band area was reduced at 20 cig/day  
124 smoking group.

125 CH stretching region

126 The CH stretching pattern of the control erythrocytes indicated the presence of four  
127 absorption bands that were centered at 2962 ± 3, 2920 ± 2, 2870 ± 2 and 2846 ± 3 cm<sup>-1</sup> and

128 assigned to asymmetric  $\text{CH}_3$  ( $\text{asymCH}_3$ ), asymmetric  $\text{CH}_2$  ( $\text{asymCH}_2$ ), symmetric  $\text{CH}_3$  ( $\text{symCH}_3$ )  
129 and symmetric  $\text{CH}_2$  ( $\text{symCH}_2$ ) respectively (figure 2). This pattern was changed in all smoking  
130 subjects. The number of absorption bands was decreased at 10 and 15 cig/day smoking  
131 groups with restricted  $\text{symCH}_2$  vibrational mode. Subjects with smoking rate of 20 cig/day  
132 showed a different CH vibrational pattern; in addition to the restricted  $\text{symCH}_2$  mode, there is  
133 an additional stretching mode of vibration that was detected at  $2904 \pm 2 \text{ cm}^{-1}$  and attributed to  
134 aliphatic CH ( $\text{CH}_{\text{aliph}}$ ). There is no change in band position or bandwidth of smoking  
135 subjects'  $\text{asymCH}_3$  or  $\text{symCH}_3$  while, the band position of smoking  $\text{asymCH}_2$  was found to be  
136 increased relative to the control pattern as given in table 2. This table also shows undetectable  
137 changes in band area of  $\text{asymCH}_3$ ,  $\text{asymCH}_2$  and  $\text{symCH}_3$  bands relative to their corresponding  
138 control ones.

#### 139 Amide I region

140 Analysis of amide I band by using the curve enhancement procedure (figure 3) resolved the  
141 control band into three structural components that are centered at  $1688 \pm 3$ ,  $1655 \pm 2$  and  
142  $1623 \pm 2 \text{ cm}^{-1}$  and can be attributed to  $\beta$ -turn structure,  $\alpha$ -helix and  $\beta$ -sheet respectively (Lin  
143 et al. 1998). Due to cigarette smoking, the contour of amide I can be resolved into five  
144 structural components with additional structural components that were centered on  $1663 \pm 3$   
145 and  $1612 \pm 2 \text{ cm}^{-1}$  and can be assigned to turns and antiparallel  $\beta$ -sheet. The significance of  
146 these results is presented in table 3, which shows the area percentage of each structural  
147 component relative to the total band area. Alpha-helix content is significantly decreased in all  
148 smoking groups concomitant with a significant increase in  $\beta$ -turn and  $\beta$ -sheet structures. The  
149 interesting finding is the detection of Turns-structure that was associated to all smoking rates  
150 (Rose et al. 1985).

#### 151 Fingerprint region

152 The other frequency range under consideration is  $1600-900\text{ cm}^{-1}$ , which is shown in figure 4.  
153 There is broadening in the contour of smoking groups with reduction in the absorption  
154 intensities; concomitant to reduced number of the estimated peaks. The control pattern shows  
155 seven absorption bands with eleven estimated structural peaks as given in table 4. No change  
156 in the band characteristics (frequency, bandwidth or band area) of amide II while, reduced  
157 band area of asymmetric COOC ( $_{\text{asym}}\text{COOC}$ ) band was found. The splitting noticed in both  
158 scissoring  $\text{CH}_2$  ( $_{\text{sciss}}\text{CH}_2$ ) and symmetric  $\text{PO}_2$  ( $_{\text{sym}}\text{PO}_2$ ) bands of control group was restricted  
159 in all smoking ones. Another common feature noticed in these two bands is the increased  
160 bandwidth that was associated to different smoking rates. The band position of  $_{\text{sym}}\text{PO}_2$  was  
161 sensitive to all smoking rates (decreased), while that of  $_{\text{sciss}}\text{CH}_2$  was increased in 10 and 15  
162 cig/day groups only. The symmetric COO ( $_{\text{sym}}\text{COO}$ ) higher frequency estimated component  
163 was characterized by decreased vibrational frequency in all smoking subjects, and this is  
164 contrary to the lower frequency one where its vibrational frequency was increased while, both  
165 of them were associated with reduced band area and vibrational motion (bandwidth).  
166 Reduced band position and band area was detected for the asymmetric  $\text{PO}_2$  ( $_{\text{asym}}\text{PO}_2$ ) band  
167 without any change in its vibrational motion. Also, no splitting in the rocking  $\text{CH}_3$  ( $_{\text{rock}}\text{CH}_3$ )  
168 band in all smoking groups with fluctuations in its vibrational frequency that was associated  
169 with increasing vibrational motion. In the same context, increased band area was also  
170 detected for the higher frequency band relative to its corresponding control band.

## 171 Discussion

172 Cigarette mainstream is a complex and heterogeneous mixture that contains volatile  
173 chemicals (e.g. formaldehyde), particles (e.g. nicotine) and gases (e.g. carbon monoxide).  
174 Moreover, a puff of cigarette smoke can contain about 300 million to 3.5 billion particles  
175 (Satici et al. 2003, Galor and Lee 2011). It also contains more than  $10^{14}$  low-molecular

176 carbon and oxygen centered radicals per puff with almost 500 ppm nitric oxide and other  
177 reactive nitrogen oxides (Pryor et al. 1993).

178 The obtained results from NH-OH region showed several fluctuated changes associated to  
179 different smoking rates, and only two structural components –namely  $_{str}OH$  that detected at  
180 an average vibrational frequency of  $3535 \pm 10 \text{ cm}^{-1}$  and  $_{sym}NH$  one – can be directly related to  
181 cigarette smoking, meaning that can be used to monitor/probe the impact of cigarette  
182 smoking on the erythrocytes membrane. This region also indicates that the detected  $_{sym}NH$   
183 mode should had another impact on the function of erythrocytes membranes since it is an  
184 indicator that the membrane become more ordered as previously reported by Schultz et al.  
185 1998 that symmetric stretching vibrations are functioning as a marker for membrane order.  
186 In spectroscopy, the area under the peak is used as an estimate of the concentration of the  
187 compound. Regarding this issue, the increased membrane order is also evident by the  
188 increased concentration of all structural components of  $_{sym}OH$  band and to clarify this last  
189 finding, the ratio of total band areas of  $_{sym}OH/_{asym}OH$  in control erythrocytes was 0.2 while it  
190 increased as the smoking rate increased to 15 and 20 cigarette/day to be 0.4 and 0.6  
191 respectively. In the same context, the relative-total concentration of the individual structural  
192 components of  $_{str}OH$  bands was also found to be increased from  $93 \pm 4$  in the control  
193 erythrocytes and reaches  $187 \pm 5$ ,  $111 \pm 2$  and  $118 \pm 2$  that corresponds to smoking rates of  
194 10, 15 and 20 cigarette/day. The OH bond can be found in many membrane constituents and  
195 one of these possibilities is cholesterol, which may support this finding about increased  
196 erythrocytes membrane order.

197 On the other hand, the CH region was greatly affected by all smoking rates involved in this  
198 study and, in particular symmetric and asymmetric  $CH_2$  bands which again give the impetus  
199 about the possibility of using these two bands to probe cigarette smoking-induced changes.

200 This vibrational region is used generally to characterize the lipid molecules within the cell

201 membrane. As shown in figure 2 and table 2, there are changes in the molecular environment  
202 of the  $_{\text{asym}}\text{CH}_2$  as reflected by increased band position. The disappeared  $_{\text{sym}}\text{CH}_2$  mode of  
203 vibration was previously reported by Szalontai et al. (2003) and Sherif (2010) and referred to  
204 it as a lipid-related phenomenon correlated with changes in protein secondary structure. This  
205 vibrational region can also be used to monitor the compositional changes of the erythrocytes  
206 membrane by considering the concentration ratio  $_{\text{asym}}\text{CH}_3$  (lipids)/ $_{\text{sym}}\text{CH}_3$  (protein). For  
207 control erythrocyte membrane this ratio was found to be 0.9, while it decreased for smoking  
208 rates of 10 and 20 cigarette/day to 0.8 and 0.7 respectively; and mimicking the control value  
209 for 15 cigarette/day group. This reveals that erythrocyte membranes, and due to different  
210 smoking rates are suffer from fluctuating lipid/protein ratio and it may be a buffering  
211 response for the induced cigarette stress.

212 Proteins in biological membranes can perturb the lipid environment and, depending on their  
213 nature and concentration, influence membrane fluidity (Chapman et al. 1979, Szalontai et al.  
214 2003).The erythrocyte proteins are essential for the linkage connecting the membrane  
215 skeleton to the lipid bilayer, which is also essential for membrane stability (Lux et al. 1978).  
216 The skeletal protein network appears to play a key role in the maintenance of the membrane's  
217 discoid shape and in restriction of the lateral mobility of its molecules (Goodman and  
218 Branton 1978, Low et al. 1991). Amide I mode of vibration in the  $1800\text{-}1600\text{ cm}^{-1}$  spectral  
219 region was used to study the protein secondary structure in infrared spectroscopy since this  
220 absorption is mainly associated with C=O stretching vibrations and it is suitable as a probe to  
221 determine the different secondary structures and polypeptides (Lux 1979, Susi et al. 1967).  
222 The obtained results (figure 3 and table 3) show that the protein secondary structure was  
223 affected by all smoking rates. It has been suggested that protein insolubility is functioning in  
224 the content of  $\beta$ -sheet structure: more  $\beta$ -sheet structure means more insoluble protein and, the  
225 formation of  $\beta$ -sheet can be deduced by firstly increasing the disordered structure of the

226 helical structure, and then, the disordered chains aggregate to form  $\beta$ -sheet structure (Lin et  
227 al. 1998, Zagorski and Barrow 1992). Accordingly, smoking rates of 10, 15 and 20 cig/day  
228 strongly affect the solubility of erythrocytes membrane protein and resulted in insoluble  
229 protein as indicated by both reduced  $\alpha$ -helix content and the associated increased  $\beta$ -sheet  
230 content, as well as protein becomes more folded due to increased  $\beta$ -turns and turns structure.  
231 These complex changes in erythrocytes membrane protein secondary structure explain the  
232 disappearance of  ${}_{\text{sym}}\text{CH}_2$  mode of vibrations as previously mentioned. Maintenance of  
233 appropriate membrane lipid composition and fluidity are important for the proper functioning  
234 of integral membrane proteins, membrane bound enzymes, receptors and ion channels  
235 (Reddy et al. 2009, Swapna et al. 2006).

236 The complicated effects of smoking are also evident in the fingerprint region. The  
237 erythrocytes membrane pattern shown in figure 4 and analyzed in table 4, reflect the fact that  
238 erythrocytes hydrocarbon chains ( ${}_{\text{sciss}}\text{CH}_2$  and  ${}_{\text{rock}}\text{CH}_3$ ) and the phospholipids ( ${}_{\text{sym}}\text{PO}_2$ ) can be  
239 found in two different phases, which greatly affected -by all smoking rates involved in this  
240 study- and turned into one phase. Moreover, the  ${}_{\text{sciss}}\text{CH}_2$  vibrations are characteristic of the  
241 nature of the acyl chain packing (Cameron et al. 1980, Swapna et al. 2006) hence; cigarette  
242 smoking has an effective impact on the acyl chain packing within the erythrocytes membrane.  
243 Altogether showed that erythrocytes membrane reacts positively with cigarette smoke  
244 mainstream in a complicated manner without any specific trends where all membrane  
245 constituents are involved, and there were certain vibrational modes that can be used as a  
246 probing marker for cigarette smoking aggravated changes. Regardless the number of cigarette  
247 smoked/day, it affects the NH-OH region of either proteins or phospholipids where each  
248 smoking rate has its own effects. Cigarette smoking resulted in increased erythrocytes  
249 membrane order which affects the membrane fluidity and accordingly its function. The

250 changes in protein secondary structure, acyl chain packing and the detected compositional  
251 changes of the CH region could influence the discoid shape of the erythrocytes.

## 252 **Conflict of interest**

253 There are no conflicts of interest related to any of the participants in this research work.

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329

330 **Figure caption**

331

332 Figure 1. Erythrocytes membrane NH-OH region ( $4000-3000\text{ cm}^{-1}$ ) of the control group and  
333 cigarette smoking ones, showing the underlying bands that detected upon using curve fitting  
334 analysis. The numbers above the peaks to facilitate their assignment.

335

336 Figure 2. Stretching CH region ( $3000-2800\text{ cm}^{-1}$ ) of control and cigarette smoking  
337 erythrocytes membrane.

338

339 Figure 3. Analysis of Amide I band showing the mean peak and the underlying structural  
340 components. The numbers above the peaks is to facilitate their assignment.

341

342 Figure 4. FTIR spectra of fingerprint region of different erythrocytes membranes involved in  
343 this study.

344

345 **Table captions**

346

347 Table 1. Vibrational frequencies, band area and bandwidth of NH-OH structural components  
348 of control and cigarette smoking erythrocytes membrane.

349

350 Table 2. Erythrocytes membrane characteristics of CH stretching region of control and  
351 cigarette smoking groups.

352

353 Table 3. Protein secondary structure components expressed as the area percentage of each  
354 structural component relative to the total band area.

355

356 Table 4. Analysis of erythrocytes membrane fingerprint region of all subjects included in the  
357 study.

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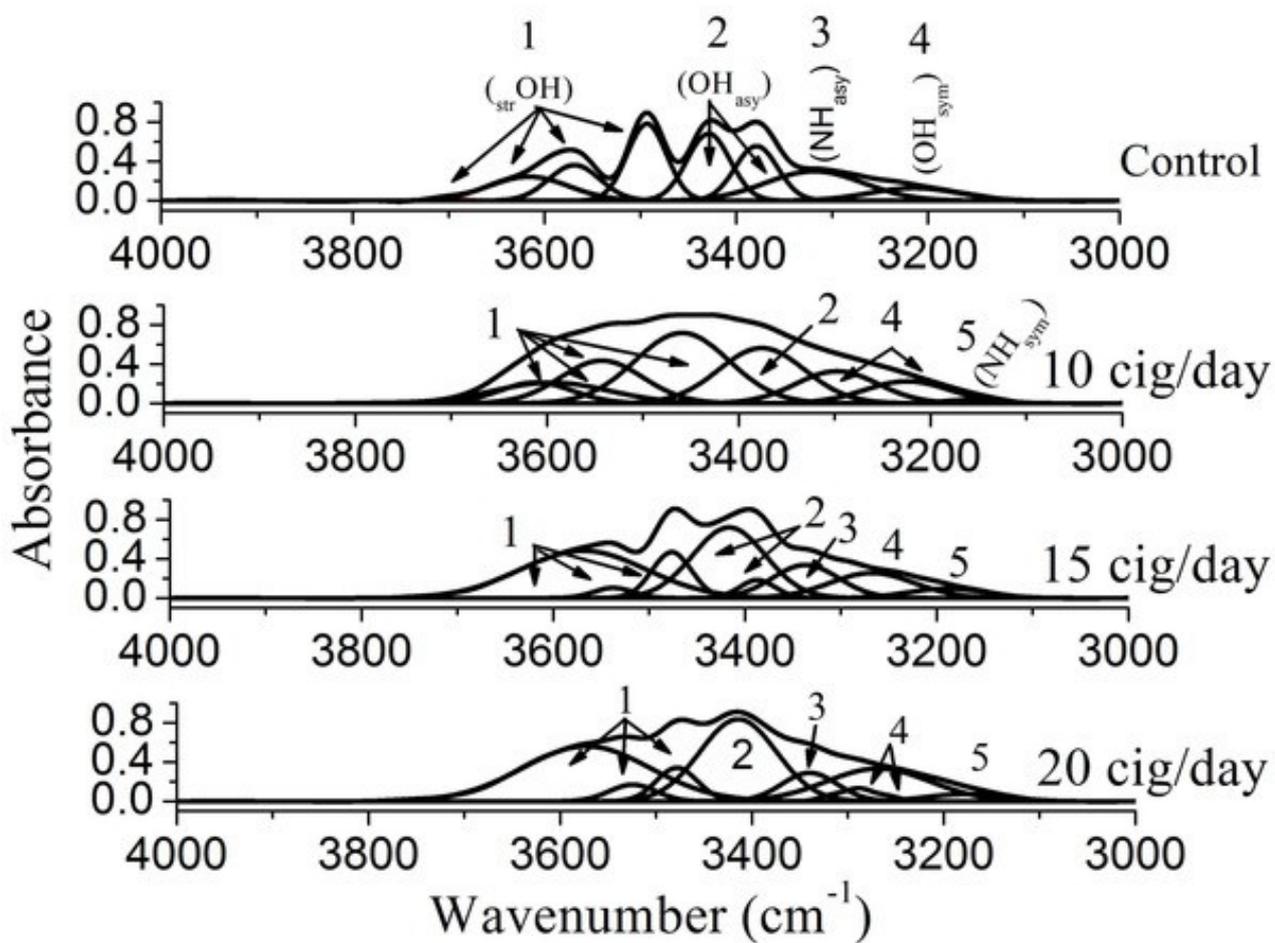


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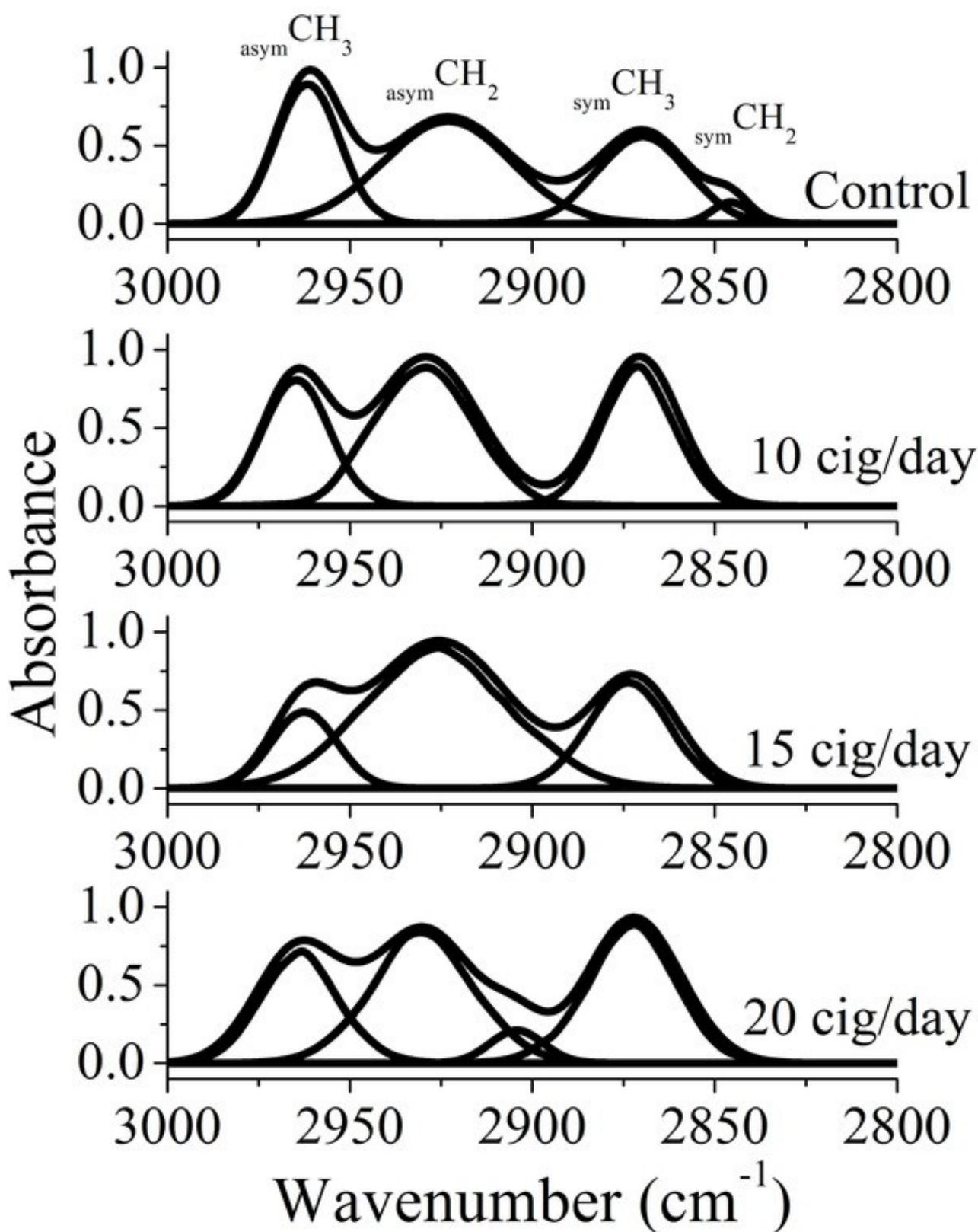


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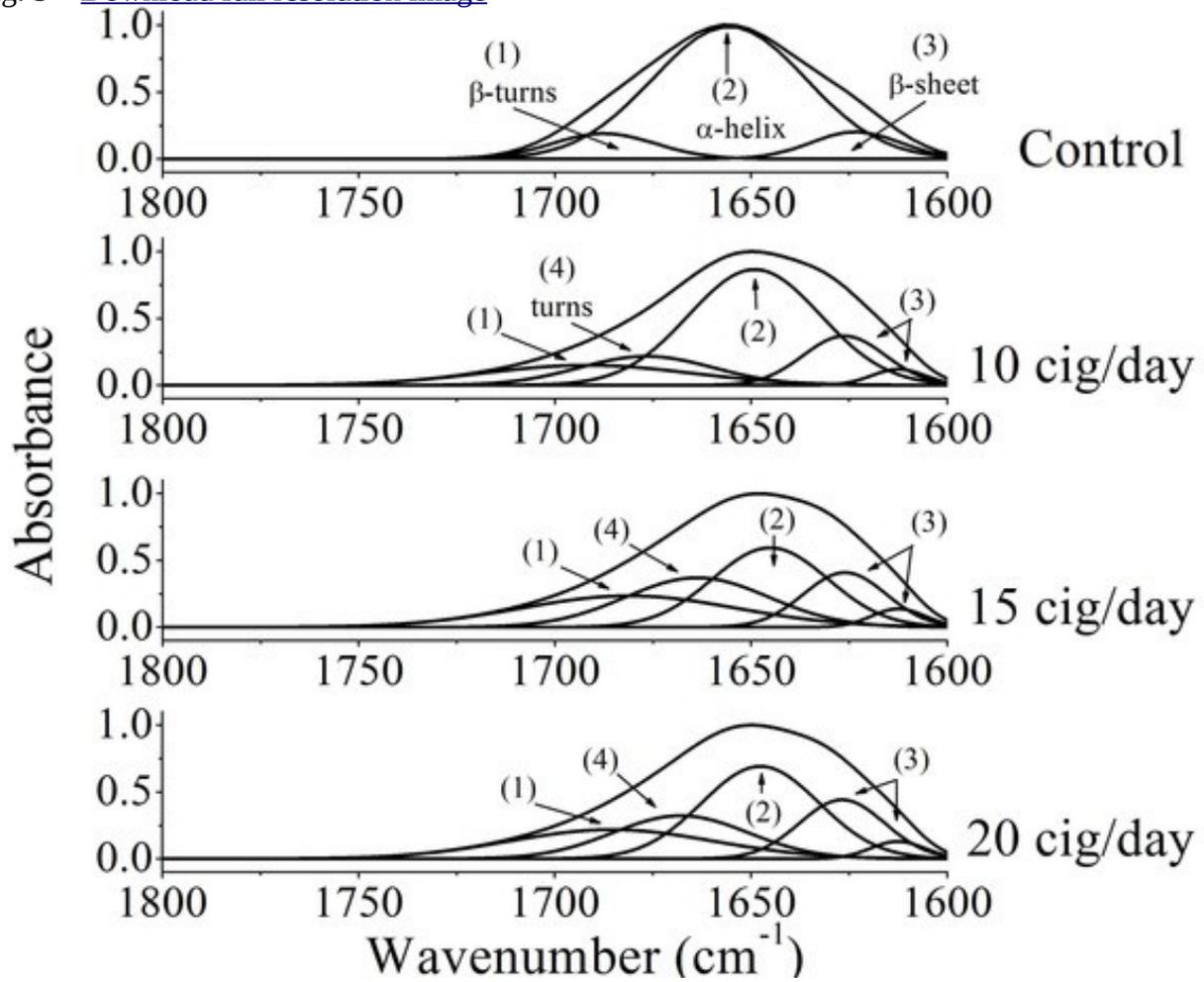


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