

**Title: Effect of maternal renin-angiotensin-aldosterone system activation on social coping strategies and gene expression of oxytocin and vasopressin in the brain of rat offspring in adulthood**

Running title: Maternal RAAS activation and programming of social coping

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**Abstract**

The intrauterine condition in which the mammalian foetus develops has an important role in prenatal programming. The aim of this study was to determine the extent to which activation of the maternal renin-angiotensin-aldosterone system (RAAS) could influence social behaviour strategies in offspring via changes in social neurotransmitters in the brain. Pregnant female Wistar rats were implanted with osmotic minipumps which continually released angiotensin II for 14 days at concentration of 2 µg/kg/h. The adult offspring (angiotensin and control groups) underwent a social interaction test. The mRNA expression of vasopressin, oxytocin and the oxytocin receptor in selected brain areas was measured by in situ hybridisation. Prenatal exposure to higher levels of angiotensin II resulted in a strong trend toward decreased total social interaction time and significantly decreased time spent in close proximity and frequency of mutual sniffing. The angiotensin group showed no changes in oxytocin mRNA expression in the hypothalamic paraventricular or supraoptic nuclei, but this group had reduced vasopressin mRNA expression in the same areas. We concluded that maternal activation of RAAS (via higher levels of angiotensin II) caused inhibition of some socio-cohesive indicators and decreased vasopressinergic activity of offspring. Taken together, these results suggest a reactive rather than proactive social coping strategy.

Keywords: renin-angiotensin-aldosterone system; angiotensin II; social coping; prenatal programming

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1 **Full title:**

2 **Effect of maternal renin-angiotensin-aldosterone system activation on social coping**  
3 **strategies and gene expression of oxytocin and vasopressin in the brain of rat offspring**  
4 **in adulthood**

5

6 Running head: Maternal RAAS activation and programming of social coping

7

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21

22 **Abstract**

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24 role in prenatal programming. The aim of this study was to determine the extent to which  
25 activation of the maternal renin-angiotensin-aldosterone system (RAAS) could influence  
26 social behaviour strategies in offspring via changes in social neurotransmitters in the brain.  
27 Pregnant female **Wistar** rats were implanted with osmotic minipumps which continually  
28 released angiotensin II **for 14 days at concentration of 2 µg/kg/h**. The adult offspring  
29 (angiotensin and control groups) underwent a social interaction test. The mRNA expression of  
30 vasopressin, oxytocin and the oxytocin receptor in selected brain areas was measured by *in*  
31 *situ* hybridisation. Prenatal exposure to higher levels of angiotensin II resulted in a strong  
32 trend toward decreased total social interaction time, ~~prolonged latency until first contact~~ and  
33 significantly decreased time spent in close proximity and frequency of mutual sniffing. The  
34 angiotensin group showed no changes in oxytocin mRNA expression in the hypothalamic  
35 paraventricular or supraoptic nuclei, but this group had reduced vasopressin mRNA  
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37 levels of angiotensin II) caused inhibition of some socio-cohesive indicators and decreased  
38 vasopressinergic activity of offspring. Taken together, these results suggest a reactive rather  
39 than proactive social coping strategy.

40

41 **Keywords:** renin-angiotensin-aldosterone system, angiotensin II, social coping, prenatal  
42 programming

43

44 **Introduction**

45 Prenatal programming is a very complex process. During a critical developmental  
46 period, a stimulus or insult can alter foetal or neonatal physiological processes, which can  
47 have a long-lasting or permanent effect on the brain, behaviour and disease risk (Barker,  
48 1998; Bale et al., 2010). The intrauterine conditions in which the mammalian foetus develops  
49 has an important role in this process (Barker, 1998).

50 One of the systems, which can be prenatally programmed, is the renin–angiotensin–  
51 aldosterone system (RAAS) (Nuyt, 2008). The RAAS is an important regulator of the blood  
52 pressure and the fluid/electrolyte homeostasis (Zhuo and Li, 2011). However, its role in  
53 modulating different physiological/pathological functions has also been described. The  
54 components of the RAAS are present in the central nervous system **modulating** sensory  
55 information, emotional and behavioural responses, learning, memory, pain, anxiety and stress  
56 (Llorens-cortes and Mendelsohn, 2002; Von Bohlen Und Halbach and Albrecht, 2006; Bali et  
57 al., 2014). RAAS is necessary for proper development of the foetus and all the components of  
58 this system are expressed in early human development (Schutz et al., 1996).

59 During pregnancy, the maternal RAAS can be influenced by several external and  
60 internal factors (O'Regan et al., 2004; Nogueira et al., 2007; Sykes et al., 2014; Xiao et al.,  
61 2014), which can alter the level of the individual components of this system. **In humans,**  
62 **angiotensin converting enzyme (ACE) inhibitors and AT1 receptor (AT1R) antagonist are**  
63 **widely used in therapy of hypertension and it is not unique that pregnant women have this**  
64 **medication (Cooper et al., 2006). However, foetal exposure to angiotensin converting enzyme**  
65 **inhibitors and/or AT1 receptor antagonist has been associated with multiple developmental**  
66 **defects or even intrauterine foetal death (Quan, 2006; Bullo et al., 2012).** On the other hand,  
67 very little data has been provided about the effects of increased angiotensin II (Ang II)

68 concentrations during pregnancy on *in utero* development of the foetus (Svitok et al., in  
69 press).

70 Foetal adaptations to supraoptimal intrauterine conditions (e.g. chronic variable stress)  
71 can lead to changes in the physiological mechanisms associated with coping strategies  
72 (Mueller and Bale, 2007).

73 Two stress response patterns have been distinguished based on social stress research in  
74 animals and humans - proactive (active) and reactive (passive) coping (Henry and Grim,  
75 1990; Koolhaas et al., 1999, 2010).

76 Proactive social coping behaviour with higher sympathetic reactivity has been shown  
77 to increase the predisposition for ~~developing~~ hypertension, atherosclerosis and  
78 tachyarrhythmia (Henry and Grim, 1990; Hessing et al., 1994; Sgoifo et al., 2005). On the  
79 other hand, reactive social coping behaviour is associated with higher parasympathetic tone  
80 and reactivity, predisposing the individual to sudden cardiac death due to bradyarrhythmia  
81 (Koolhaas et al., 1999).

82 Social stressors play an important role during the mammalian life course (Koolhaas et  
83 al., 1997; Sgoifo et al., 2005), and prenatal programming influences the adaptation strategy for  
84 this type of stress. The oxytocin (OT) and vasopressin (AVP) neuropeptides are involved in  
85 inter-neuronal communication within various areas of the brain to modulate social-emotional  
86 behavioural and physiological responses, reflective of the coping style (Holmes et al., 2003;  
87 Koolhaas et al., 2010). ~~While AVP is generally known to increase anxiety-like behaviour,  
88 stress and aggressiveness, OT has the opposite effect, facilitating social attachment, care and  
89 relationships (Insel et al., 1998; Heinrichs et al., 2009).~~

90 Both neuropeptides are produced by magnocellular and parvocellular neurosecretory  
91 neurons in the paraventricular (PVN), supraoptical (SPO) and accessory nuclei of the

92 hypothalamus. AVP is also produced by parvocellular cells in the suprachiasmatic nucleus  
93 (SCN) (Carter, 2007).

94 Increased vasopressinergic activity in discrete brain regions linked to regulation of  
95 stress coping behaviour is associated with increased levels of aggressiveness, and perhaps, a  
96 more proactive coping style in general (Bult et al., 1992; Compaan et al., 1992; Aubry et al.,  
97 1995; Everts et al., 1997; Koolhaas et al., 2010). On the other hand, increased oxytocinergic  
98 activity in these very same neural structures associated with facilitating social attachment,  
99 care and relationships (Insel et al., 1998; Heinrichs et al., 2009) and may be implicated in  
100 more reactive (passive) coping behaviours (de Boer et al., 2003; Koolhaas et al., 2010).

101 In the current study, we investigated the extent to which the activation of maternal  
102 RAAS (via prenatally increased Ang II) could influence social behaviour strategies in adult  
103 offspring via changes in the level of social neurotransmitters in the brain.

104

## 105 **Material and Methods**

106 Our research is part of a larger project focused on the effects of prenatal activation of  
107 regulatory mechanisms of blood pressure. The experiment was performed in compliance with  
108 the Principles of Laboratory Animal Care issued by the Ethical Committee of Comenius  
109 University in Bratislava, Slovak Republic. The experimental design was approved by the  
110 State Veterinary and Food Administration of the Slovak Republic.

111

### 112 ***Animals***

113 Wistar rats (VELAZ Praha, Czech Republic) were used in this study. The parental  
114 generation consisted of nine females and four males. Animals were housed in groups of two  
115 or three animals in standard light conditions (12:12 hour light-dark cycle; lights on at 6:00),  
116 with an average temperature of  $21 \pm 2^\circ\text{C}$  and  $55 \pm 5\%$  relative humidity. Water and food

117 (standard laboratory chow) were available *ad libitum*. After an acclimatisation period of 7  
118 days, female rats were divided into two groups: the control group (Ctrl; n = 4), and the  
119 angiotensin II group (AngII; n = 5). The animals were mated overnight. The ovulatory cycle  
120 phase and day 0 of gestation (the presence of spermatozoa in a vaginal smear) were identified  
121 according to Gleich and Frohberg (1977).

122 Osmotic minipumps were implanted on the sixth day of gestation (model 2002, Alzet,  
123 Canada), placed subcutaneously on the back of dams in the AngII group, through which Ang  
124 II (1.36 mg/ml in physiological solution) was continuously released at 2 µg/kg/h for 14 days.  
125 In contrast, an oval object of the same shape and size (sham operation) was implanted into  
126 Ctrl dams. For the surgery, animals were anaesthetised using a solution of ketamine (135  
127 mg/ml; Narketan<sup>TM</sup> 10, Chassot GmbH, Germany) and xylazine (18 mg/ml; Rometar 2%,  
128 Spofa, Czech Republic). The animals were kept under the same conditions as prior to surgery  
129 until birth. Following birth, the litters were culled to eight animals per litter (four males and  
130 four females). Each dam and litter were housed together in individual cages until weaning at  
131 postnatal day 21. After weaning, rats of each gender were housed separately in groups of four  
132 animals per cage.

133 Body weight did not differ between control and AngII dams before, or at the end of  
134 pregnancy. Ang II administration had no effects on litter size or birth weight of the offspring.  
135 Sex ratio of pups was similar between both groups and the body weight did not differ between  
136 groups during the whole course of the experiment. Increased Ang II during pregnancy raised  
137 blood pressure in the offspring. Treatment also increased aldosterone and decreased plasma  
138 renin activity in the offspring (Svitok et al., in press).

139 Animals were anaesthetised with CO<sub>2</sub> and decapitated at postnatal day 79–83. Their  
140 brains were immediately removed and placed into frozen medium (Cryomount, Histolab AB,  
141 Sweden), frozen in dry ice and stored at – 80°C. For *in situ* hybridisation, brains from the

142 litters of 4 Ctrl and 5 AngII females (Ctrl n = 20, males n = 10, females n = 10; AngII n = 20,  
143 males n = 10, females n = 10) were used. The brains of all animals tested in the social  
144 interaction test were also evaluated by *in situ* hybridisation.

145

#### 146 ***Social interaction test***

147 The social interaction test was adapted from File (1980). At 77 days of age, the  
148 offspring (Ctrl: male n = 10; female n = 10; AngII: male n = 10, female n = 10) were tested  
149 for their social interaction behaviours with an unknown test partner of the same weight (no  
150 more than 10 g difference), sex and treatment. Pairs were tested in a random order between  
151 14:00 and 15:00. The dimensions of the test box were 33.5 × 71 cm, with walls 37 cm high.

152 Animals were acclimatised the day before testing by singly placing each animal in the  
153 test box for 20 minutes. Pairs of animals were placed in diagonally opposite corners of the  
154 box. The behaviour of the animals was measured over a period of 5 minutes, which was  
155 recorded using a digital camera (DCR-DVD 92 E; Sony, Japan).

156 The latency until first contact (duration in seconds) and total duration and frequency of  
157 socio-cohesive (following, chasing, mutual sniffing, genital investigation, climbing over,  
158 crawling under, allogrooming contact, time in close proximity) and socio-aversive (escape,  
159 mounting, tail biting) interactions were recorded during the time period from the video.

160

#### 161 ***In situ hybridisation***

162 The mRNA expression of vasopressin, oxytocin (neurotransmitters responsible for  
163 social activity) in the hypothalamic paraventricular (PVN) or supraoptic (SPO) nuclei (the site  
164 of their synthesis) was measured by *in situ* hybridisation.

165 The *in situ* hybridisation protocol has been previously described elsewhere (Štefánek et  
166 al., 2015). Briefly, consecutive frozen cryostat sections of the rat brain (14 µm) were mounted

167 on adhesion slides (SuperFrost® Plus, MenzelGläser, Thermo Fisher Scientific Inc.,  
168 Germany) for subsequent hybridisation of the chosen mRNAs. A synthetic 41-base  
169 oligonucleotide probe with the sequence 5'-GGG CTC AGC GCT CGG AGA AGG CAG  
170 ACT CAG GGT CGC AGG CG-3' was used, complementary to nucleotides 906–946 of the  
171 rat OT mRNA (GenBank accession number K01701) (Patisaul et al., 2003). For hybridisation  
172 of AVP mRNA, a 48-base probe with the sequence 5'-GCA GAA GGC CCC GGC CGG  
173 CCC GTC CAG CTG CGT GGC GTT GCT CCG GTC-3' was used (directed against the  
174 final 16 amino acids of the glycoprotein that is not shared with oxytocin) (Müller et al., 2000).  
175 All sequences were firstly checked for complementarity with non-specific rat RNA using  
176 BLAST in the EMBL sequence database. None of the probes recognised different non-  
177 specific sequences with sufficient fidelity.

178 Probes were labelled at the 3'-end with [ $\alpha$ -<sup>35</sup>S] dATP (1200 Ci/mmol; Perkin Elmer  
179 Inc., USA) by terminal deoxynucleotidyl transferase (Thermo Fisher Scientific Inc., Canada).  
180 Unincorporated radioactivity was removed with Sephadex G-50 micro-columns (GE  
181 Healthcare, Little Chalfont, UK). The sections were fixed in 4% paraformaldehyde in  
182 phosphate buffered saline (PBS; pH 7.4) for 5 minutes, then rinsed twice in PBS, acetylated in  
183 a solution containing 0.25% acetic anhydride in 0.8% triethanolamine hydrochloride and  
184 0.9% sodium chloride for 10 minutes. Sections were dehydrated through graded ethanol  
185 solutions, delipidated in chloroform for 5 minutes, then dipped in 100 and 95% ethanol prior  
186 to air-drying. Sections were hybridised with the labelled probe in 70  $\mu$ l of hybridisation buffer  
187 per slide (50% formamide, 4 $\times$  saline-sodium citrate (SSC) buffer, 1 $\times$  Denhardt's solution,  
188 10% dextran sulphate, 500  $\mu$ g/ml sheared single-stranded DNA and 250  $\mu$ g/ml yeast tRNA),  
189 which was mixed with 0.1 M dithiothreitol (DTT) and the purified radiolabelled oligoprobe  
190 (270,659 cpm/ $\mu$ l for OT and 186,625 cpm/ $\mu$ l for AVP) before use. The radiolabeled probe  
191 was diluted to a concentration of  $2.7 \times 10^6$  counts/min/ml (OT and AVP). The slides were

192 incubated in humidified boxes for 18 hours at 41°C. Post-hybridisation, the slides were  
193 processed by washing with 1× SSC for 5 minutes at room temperature, then in 1× SSC for  
194 1 hour at 55°C, and finally in 1× SSC for 1 hour at room temperature. Afterwards, the slides  
195 were dipped in distilled water then 70% ethanol, then left to air-dry.

196 Slides with probes for OT and AVP were exposed to Hyperfilm-Beta-Max (Eastman  
197 Kodak Company, Chalon-sur-Saône, France) for 48 hours. The film was developed using a  
198 developer (LQN) solution then fixed in FOMAFIX (FOMA, Hradec Králové, Czech  
199 Republic) solution. After exposure, the slides were stained with cresyl violet to determine the  
200 position and shape of the neuronal structures.

201 Autoradiographs were captured using an optical magnifier and digital camera  
202 (AM423X Dino-Eye, Dino-Lite, Taiwan) and corresponding software (Dino Capture 2.0,  
203 Dino-Lite, Taiwan), then analysed using ImageJ software (version 1.47i; NIH, USA). The  
204 relative density of individual structures was measured for each slide. Optical densities of the  
205 SON and PVN nuclei were determined for OT and AVP. The optical density of signals was  
206 bilateral, but was separately evaluated for each section. As an internal standard, the  
207 background density adjacent to each structure was captured and subtracted from the density of  
208 the region of interest (performed separately for each side). The mean values from the left and  
209 right side were obtained from four to six slides per animal. The rat brain atlas (Paxinos and  
210 Watson, 1998) was used to verify the correct locations.

211

## 212 *Statistical analysis*

213 All data were analysed using Kolmogorov-Smirnov tests to determine whether they fit  
214 a normal distribution. The results were analysed using Statistica version 7.0 software  
215 (StatSoft, Inc., Tulsa, OK, USA). Data were analysed by ~~general-linear-model~~ two-way  
216 analysis of variance (ANOVA) with the treatment and ~~sex-gender~~ as co-factors. If the

217 interaction was significant ( $p < 0.05$ ), differences in behavioural parameters between control  
218 and AngII rats were estimated by Fischer LSD *post hoc* test. ~~If data did not fit a normal~~  
219 ~~distribution, the Kruskal-Wallis test was used.~~

220

## 221 **Results**

### 222 *Social interaction test*

223 For the latency until first contact, the two-way ANOVA did not reveal any significant  
224 effect of treatment ( $F_{(1,36)} = 0.001$ ,  $p = 0.97$ ), sex ( $F_{(1,36)} = 0.167$ ,  $p = 0.685$ ) and interaction  
225 treatment vs. sex ( $F_{(1,36)} = 2.124$ ,  $p = 0.154$ ).

226 For the total time spent participating in socio-cohesive interactions, the two-way  
227 ANOVA did not reveal any significant effect of treatment ( $F_{(1, 36)} = 2.26$ ,  $p = 0.14$ ), sex ( $F_{(1, 36)}$   
228  $= 1.62$ ,  $p = 0.21$ ) and interaction treatment vs. sex ( $F_{(1,36)} = 1.717$ ;  $p = 0.2$ ). We recognised  
229 only a trend towards a lower number of socio-cohesive interactions in AngII females in  
230 comparison with Ctrl females ( $p = 0.054$ ; Fig. 1a).

231 For the time spent in close proximity, the two-way ANOVA revealed significant effect  
232 of treatment ( $F_{(1,36)} = 7.487$ ;  $p < 0.05$ ), sex ( $F_{(1,36)} = 6.343$ ;  $p < 0.05$ ), and interaction treatment  
233 vs. sex ( $F_{(1,36)} = 10.456$ ;  $p < 0.01$ ). In total, AngII animals spent less time in close proximity  
234 compared to Ctrl animals ( $p < 0.05$ ; Fig. 1b). We observed differences between males and  
235 females, such that AngII females spent less time in close proximity to the other animal during  
236 this test compared to Ctrl females ( $p < 0.001$ ). Ctrl males also spent less time in close  
237 proximity than Ctrl females ( $p < 0.001$ ; Fig. 1b). The two-way ANOVA revealed significant  
238 effect of treatment for frequency of sniffing ( $F_{(1,36)} = 6.1425$ ;  $p < 0.05$ ). Animals in the AngII  
239 group had a lower frequency of sniffing in comparison with Ctrl males ( $p < 0.05$ ; Fig. 1c).

240 Socio-aversive interactions were not observed for any animal in either the AngII or  
241 Ctrl groups.

242

243 < Please insert Fig. 1 >

244

### 245 *In situ hybridisation*

246 The two-way ANOVA revealed significant effect of treatment ( $F_{(1,36)} = 4.4973$ ;  $p <$   
247  $0.05$ ) for AVP mRNA expression in PVN. AngII animals had lower AVP mRNA expression  
248 compared with Ctrl group (Fig. 2).

249 Statistical analysis revealed also significant effect of interaction treatment vs. sex  
250 ( $F_{(1,36)} = 5.637$ ;  $p = 0.023$ ) for AVP mRNA expression in the SPO. AngII males had lower  
251 AVP mRNA expression compared with Ctrl males (Fig. 2).

252

253 < Please insert Fig. 2 >

254

255 There were no significant effect of treatment and/or sex in OT mRNA expression in  
256 either the hypothalamic PVN or SPO between.

257

### 258 **Discussion**

259 In the current study we applied a new model for evaluating developmental  
260 programming of cardiovascular disease. This model involved the activation of maternal  
261 RAAS via implantation of osmotic minipumps that continually released Ang II during most of  
262 the gestational period. The research presented here is part of a larger project focused on the  
263 effects of prenatal activation of regulatory mechanisms of blood pressure, part of which has  
264 already been published (Svitok et al., in press).

265 We hypothesised that the activation of maternal RAAS (via higher levels of Ang II)  
266 would be associated with changes in the social coping behaviour of the offspring. There are

267 no studies in the literature that have described the prenatal effects of Ang II administration on  
268 social behaviour, therefore, we have compared our results with animal models with  
269 upregulation (TGR(mRen2)27) and downregulation (TGR(ASrAOGEN6)680) of RAAS.

270 Studies that have investigated models of downregulated RAAS have reported that rats  
271 spent more time in relatively close spatial proximity of the rats without further motor  
272 interactions compared with controls in social interaction tests (Voigt et al., 2005; Mayorov,  
273 2011). Rats with upregulated RAAS could have a similar effect to the total social interaction  
274 observed in rats with downregulated RAAS (Voigt et al., 2005).

275 Our results showed a trend toward differences in total socio-cohesive interaction.  
276 When we focused on individual parameters (~~latency until first contact~~, time spent in close  
277 proximity and frequency of sniffing), we observed similar results to that reported by Voigt *et*  
278 *al.* (2005), who observed reduced sniffing in rats with downregulated RAAS. The time spent  
279 in close proximity was also found to be decreased in AngII **treated** animals. **Rats with**  
280 **downregulated RAAS were not different in their latency until first contact behaviour (Voigt et**  
281 **al., 2005), similarly, in our study, prenatal exposure to Ang II did not change latency period**  
282 **until first contact.**

283 The present study was based on a prenatal intervention to investigate how blood  
284 pressure regulation mechanisms influence programming of social coping strategies. ~~We~~  
285 ~~recognised two coping strategies, proactive and reactive (Koolhaas et al., 1999).~~ In regard to  
286 behaviour, the proactive strategy is not only characterised by higher levels of aggression,  
287 dominance and territoriality, but also by a tendency to develop routine-like, rigid, intrinsically  
288 driven behaviours (Sgoifo et al., 2005). In contrast, the reactive strategy is characterised by  
289 less aggressive, less territorial and subordinated behaviours, in addition to being more flexible  
290 and driven by changes in environmental stimuli (Koolhaas et al., 1999; Sgoifo et al., 2005).  
291 These two styles of adaptation (coping strategies) also differ in terms of

292 autonomic/neuroendocrine (re)activity. Proactive animals have higher sympathetic activation  
293 and lower (re)activity of the HPA axis, whereas reactive animals are parasympathetically  
294 driven and have higher HPA axis (re)activity (Koolhaas et al., 1999; Sgoifo et al., 2005).

295 From our results, maternal RAAS activation (via higher levels of Ang II) caused inhibition  
296 some aspects of social behaviour in offspring, which we suggest is associated with reactive  
297 rather than proactive social coping strategies. Our suggestion is supported by higher  
298 activation of the HPA axis, that leads to increase of aldosterone levels (Kubzansky and Adler,  
299 2010). Aldosterone may induce a wide spectrum of behavioural manifestations of  
300 psychological stress (e.g., anxiety, depression) (Hlavacova and Jezova, 2008; Hlavacova et  
301 al., 2012). This mineralocorticoid plays a key role in modulation of mood through effects on  
302 relevant mood-regulating brain areas including nucleus tractus solitarii (Murck et al., 2014).  
303 According to Ngarmukos and Grekin (2001), behavioural effect of aldosterone may be  
304 mediated by non-genomic mechanism on ionic exchangers or membrane receptors.  
305 Aldosterone plays a central role in the regulation of plasma sodium through mineralocorticoid  
306 receptors (Bailey et al., 2009). Plasma sodium imbalance can lead to mood disorders and  
307 decreased reactivity to psychogenic stressors (Krause et al., 2011; Leshem, 2011).

308 In our study, AngII animals have been reported to have increased plasma aldosterone  
309 levels compared to controls (Svitok et al., in press), also observed in transgenic rats  
310 (TGR(mRen2)27) with upregulated RAAS. Aldosterone levels were found to be 10-times  
311 higher in rats with upregulated RAAS than in control rats, which supports upregulation of the  
312 renin-angiotensin system, as it is well known that angiotensin II stimulates aldosterone  
313 synthesis (Ferrario, 1983; Zeman et al., 2007).

314 Furthermore, reactivity of the HPA axis to adrenocorticotrophic hormone in transgenic  
315 rats is higher compared to other normotensive rat strains (Husková, 2006). Higher activation  
316 of the HPA axis suggests reactive social coping behaviour.

317           The two main brain neurotransmitters responsible for social activity in animals  
318 ~~(individuals)~~ are vasopressin and oxytocin. Generally, increased vasopressinergic activity in  
319 discrete brain regions is associated with proactive coping styles, and increased oxytocinergic  
320 activity in these very same neural structures is associated with reactive (passive) coping  
321 (Koolhaas et al., 2010). In our study, animals in the AngII group did not differ in OT mRNA  
322 expression in the hypothalamic PVN or SPO nuclei, however, this group had lower  
323 expression of AVP mRNA in the PVN and SPO nuclei of the hypothalamus, ~~which is the site~~  
324 ~~of AVP and OT synthesis~~. These changes again suggest a reactive rather than proactive social  
325 coping strategy.

326           In conclusion, prenatal exposure to higher levels of Ang II caused changes in social  
327 behaviour, suggestive of a reactive rather than proactive social coping strategy, ~~in addition~~  
328 supported by altered vasopressinergic activity.

329

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335

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477 **Captions**

478 **Fig. 1** Total time spent participating in socio-cohesive interactions (**a**), total time spent in  
479 close proximity (**b**), frequency of sniffing (**c**) in angiotensin (AngII: males n = 10, females n =  
480 10) and control (Ctrl: males n = 10; females n = 10) groups. Error bars represent standard  
481 error of the mean and the significance of differences is indicated by asterisk \* p < 0.05 \*\* p <  
482 0.01 \*\*\* p < 0.001.

483 **Fig. 2** *In situ* hybridisation for arginine-vasopressin (AVP) Representative *in situ* photos of  
484 AVP mRNA signals in Ctrl (**a**) and AngII (**b**) group. Comparison of optical density signal for  
485 AVP of angiotensin (AngII: males n = 10, females n = 10) and control (Ctrl: males n = 10,  
486 females n = 10) rats in paraventricular (PVN; **c**) and supraoptical nuclei (SPO; **d**). Error bars  
487 represent standard error of the mean, and the significance of differences is indicated by  
488 asterisk \* p < 0.05.

Fig. 1 [Download full resolution image](#)

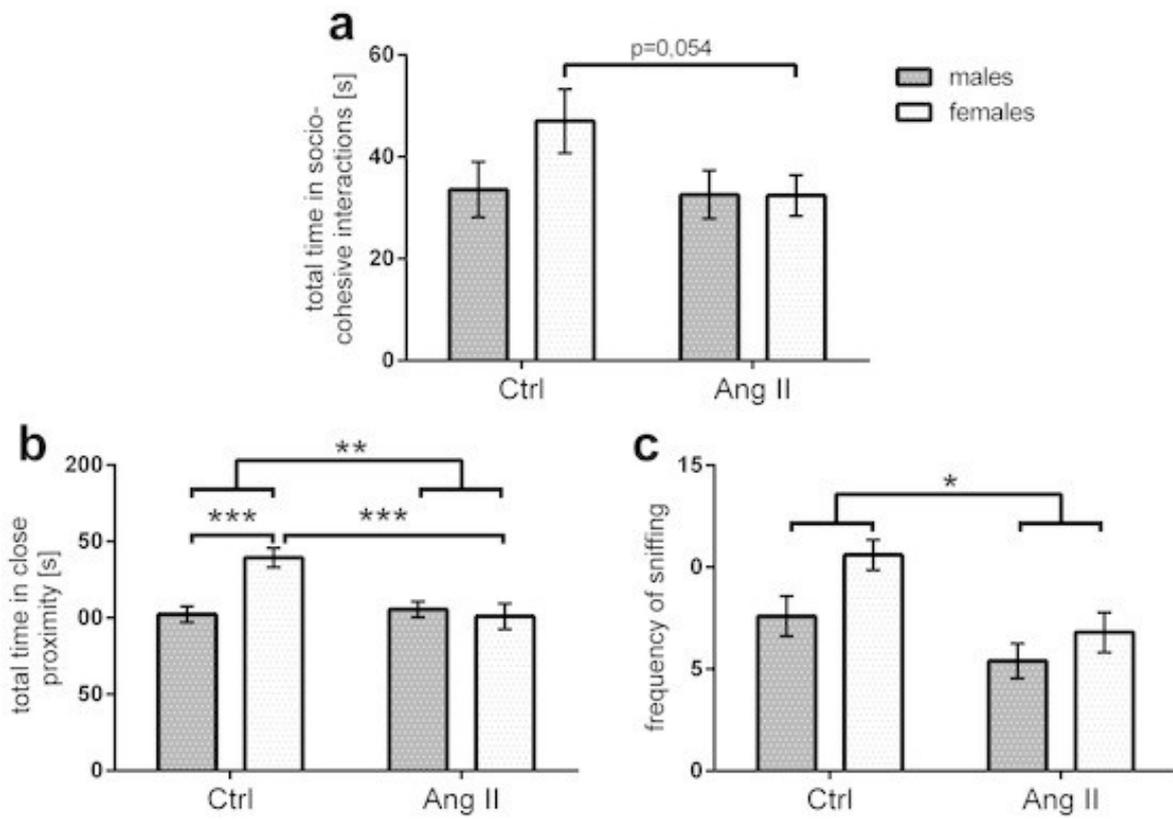


Fig. 2 [Download full resolution image](#)

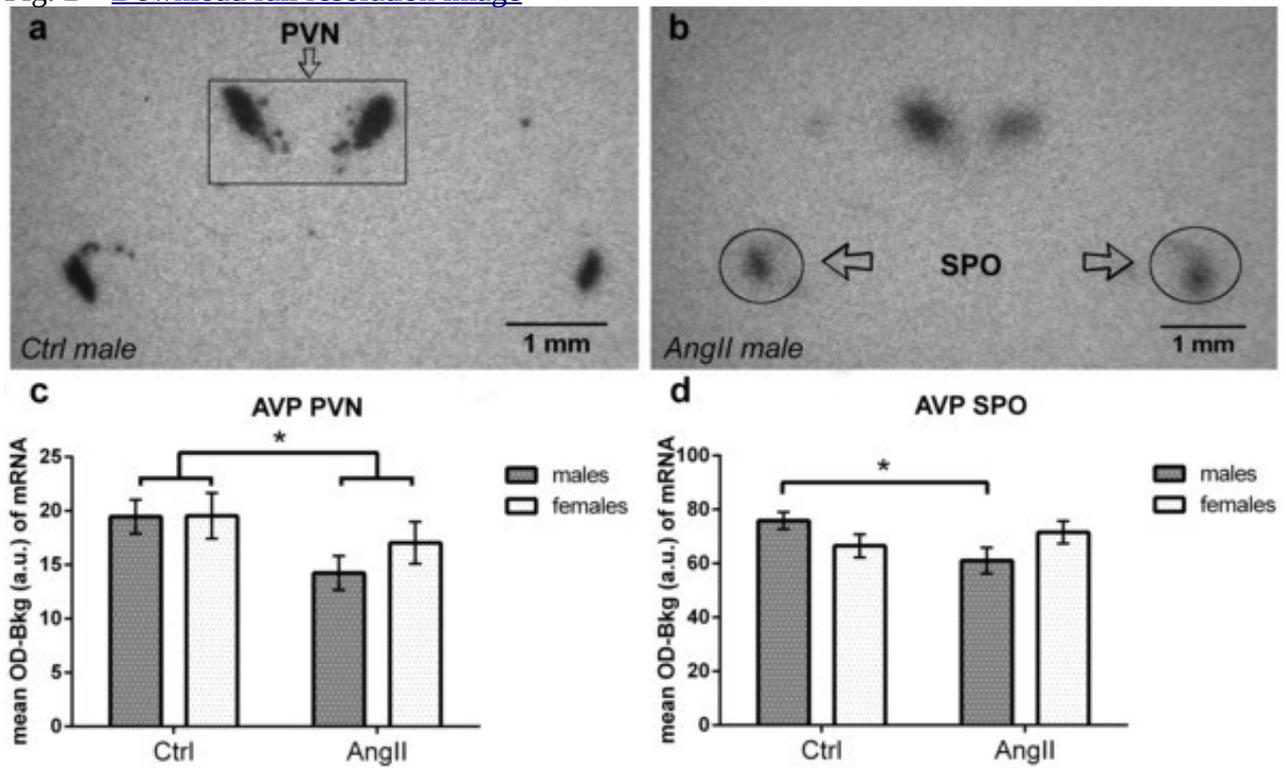


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