doi: 10.4149/gpb_2021038

Peripheral microRNA alteration and pathway signaling after mild traumatic brain injury

Katarina Matyasova^{1,*}, Nikoleta Csicsatkova^{1,*}, Peter Filipcik^{1,2}, Igor Jurisica^{1,3,4,5} and Martin Cente^{1,2}

Martin Cente ^{1,2}
¹ Institute of Neuroimmunology, Slovak Academy of Sciences, Bratislava, Slovakia
² Axon Neuroscience R&D Services SE, Bratislava, Slovakia
³ Osteoarthritis Research Program, Division of Orthopedic Surgery, Schroeder Arthritis Institute, University Health Network,
Toronto, Ontario, Canada
⁴ Data Science Discovery Centre for Chronic Diseases, Krembil Research Institute, University Health Network, Toronto,
Ontario, Canada
⁵ Departments of Medical Biophysics and Computer Science, University of Toronto, Toronto, Ontario, Canada
Abstract. Discovering novel diagnostic biomarkers and signatures for traumatic brain injury (TBI)
represents a major challenge in the brain trauma research. Detailed analysis of post-concussive
molecular pathways based on experimental data could provide a new insight into the pathophysi-
ological sequelae and mapping of recovery mechanisms involved in TBI. MicroRNAs (miRNAs)
detectable in peripheral body fluids after TBI are promising carriers of this missing knowledge. In
order to define the signature of peripheral miRNAs signaling associated with mild TBI (mTBI), we
performed a comprehensive meta-analysis of miRNA profiles in mTBI patients using multiple cu-
rated pathway databases. Using a bioinformatic pipeline with integrated data analysis we identified
a set of genes that are connected to deregulated circulating miRNAs following the mTBI. Identified
genes belong to specific pathways of MAPK, TGF- β , WNT, TLR2/4, PI3K/AKT, insulin, and growth
factor signaling. Since the enriched pathways markedly overlap among the various biological fluids,
signaling associated with mTBI that is concomitantly reflected in serum, plasma and saliva is robust
and unique. Furthermore, we identified a network of 33 validated interacting proteins and their
regulatory miRNAs that link the post-mTBI signaling in peripheral fluids with neurodegeneration-
associated interaction pathways. Presented data provide a comprehensive insight into molecular
events following mTBI, and the top predicted genes represent a group of novel candidate targets to
be validated in connection with mTBI.
Key words: Mild traumatic brain injury — TBI — miRNA — Signaling pathways — Meta-
analysis
* These authors contributed equally to this work.
Correspondence to: Martin Cente, Institute of Neuroimmunology, Slovak Academy of Sciences, Dubravska cesta 9, 845 10 Bratislava,
Slovakia
E-mail: martin.cente@savba.sk
Igor Jurisica, Data Science Discovery Centre for Chronic Diseases, Krembil Research Institute, University Health
Network, 60 Leonard Avenue, 5KD-407, Toronto, Ontario, Canada M5T 0S8 E-mail: juris@ai.utoronto.ca
© The Authors 2021. This is an open access article under the terms of the Creative Commons Attribution-NonCommercial 4.0 International License
(https://creativecommons.org/licenses/by-nc/4.0/), which permits non-commercial use, distribution, and reproduction in any medium, provided the original work is properly cited.

Introduction

1 2

3 Traumatic brain injury (TBI) is defined by a disruption of 4 normal brain function as a reaction to external physical 5 force (Menon et al. 2010). The estimated incidence of TBI 6 per 100 000 people represents 948 cases in Asia, 1012 cases 7 in Europe and the numbers are even higher (1299 cases) in North America (Dewan et al. 2019). Patients with mild 8 9 TBI (mTBI) represent majority of the hospital-treated adult 10 head injuries (70-90%); however, the overall numbers are 11 likely underestimated since many mild head injuries remain 12 unreported and thus undiagnosed (Cassidy et al. 2004; 13 McCrea et al. 2017). Evaluation by the Glasgow coma scale 14 (GCS) is a widely used diagnostic approach to determine the severity of TBI by observing patient's motor, eye, and 15 verbal responses (Teasdale and Jennett 1976). The score of 16 17 13-15 refers to mTBI, 9-12 points to moderate TBI, and less than 8 to severe TBI (Iankova 2006). In addition to 18 19 GCS, computed tomography (CT) and magnetic resonance 20 imaging (MRI) are the most available and used neuroimag-21 ing techniques in the clinical management of brain injury 22 (Shetty et al. 2016). However, the CT and MRI conclude 23 very often normal findings reporting no abnormalities in 24 mTBI patients (Iverson et al. 2000; Wintermark et al. 2015). 25 Typically, most patients after mTBI or sport-related con-26 cussion fully recover after 7-10 days, although this period 27 can be prolonged in children and adolescents (McCrory 28 et al. 2005).

29 Recent research indicates that TBI may increase the risk 30 for the development of neurodegenerative disorders evolved 31 due to longitudinal sequelae of concussion or repetitive 32 sub-concussive head impacts (Bailes et al. 2013; Manley 33 et al. 2017; Alosco et al. 2018). For instance, single moder-34 ate, single severe TBI or even repeated mild head injuries 35 are associated with the development of chronic traumatic 36 encephalopathy (CTE) as it was reported in professional 37 contact sport athletes (Omalu et al. 2005; McKee et al. 2009; 38 Smith et al. 2013; Omalu and Hammers 2021). Moreover, 39 the risk of Alzheimer's disease has been reported as 1.5 times 40 higher following head trauma (Li et al. 2017). Similarly, the 41 increased risk of Parkinson's disease has been described 42 in military personnel suffering various severities of TBI 43 (Gardner et al. 2018). Additional to the neurodegeneration, head trauma has a significant impact on mental health of 44 45 TBI individuals as well. Patients with TBI frequently suffer from sleep disturbance (Gilbert et al. 2015) and psychiatric 46 47 disorders including depression (Jorge et al. 1993), aggression 48 (Gallant et al. 2018) or posttraumatic stress disorder (Bryant 49 and Harvey 1998).

Although it is not possible to accurately predict the consequences of TBI, peripheral biomarkers could represent
a valuable diagnostic and prognostic asset. In particular, the
group of peripheral microRNAs (miRNAs) has been shown

78

80

81

82

83

84

to be associated with mTBI (Redell et al. 2010; Hicks et al. 54 2020b) and various neurodegenerative disorders (Sheiner-55 man et al. 2017; Siedlecki-Wullich et al. 2019; Uwatoko et al. 56 2019). The diagnostic potential of miRNAs originates from 57 their general abundance, resistance against degradation, 58 59 stability in different biofluids (Keller et al. 2017) and sensitive methods of detection. Furthermore, their regulatory 60 role at post-transcriptional level enables identification of 61 specific disease patterns and molecular signaling pathways 62 associated with pathophysiological processes known to be 63 involved in neurodegenerative diseases. 64

In the present study, we present a meta-analysis of circu-65 lating miRNA profiles in patients experiencing mTBI. Using 66 an integrated bioinformatic pipeline we aimed to identify 67 the molecular signature of candidate target genes, associated 68 signaling pathways and involved protein interactions in the 69 mTBI cascade. Special emphasis was placed on clarifying 70 the link between post-concussive signaling and induction 71 of neurodegeneration-associated interaction pathways. This 72 73 approach helps to improve our understanding of the molecular events present in the brain and eventually mirrored 74 in peripheral fluids during early post-traumatic period and 75 to discover the novel candidate markers/targets associated 76 with mTBI. 77

Methods

Search strategy, selection criteria and data extraction

A systematic search of relevant studies describing the 85 altered miRNA expression levels in human peripheral 86 biofluids of patients diagnosed with mTBI was performed 87 in the PubMed database using the following query: ("trau-88 matic brain injury"[Title/Abstract] OR "traumatic brain 89 injuries" [Title/Abstract] OR "TBI" [Title/Abstract] OR "head 90 trauma"[Title/Abstract] OR "head traumas"[Title/Abstract] 91 OR "concussion" [Title/Abstract] OR "concussions" [Title/ 92 Abstract] OR "head impact" [Title/Abstract] OR "head 93 impacts"[Title/Abstract]) AND ("microRNA"[Title/Ab-94 stract] OR "microRNAs" [Title/Abstract] OR "miRNA" [Title/ 95 Abstract] OR "miRNAs" [Title/Abstract] OR "mirna 96 biomarker" [Title/Abstract] OR "mirna biomarkers" [Title/ 97 Abstract]). In total, 220 articles published before December 98 99 31, 2020, were identified.

The inclusion criteria for the articles were as follows: (1) 100 original research, (2) mTBI patients clinically diagnosed 101 according to GCS \geq 13, (3) miRNA expression study in 102 human peripheral biofluids, (4) acute post-TBI time <15 103 days. First, the titles and abstracts of searched articles were 104 screened, followed by review of full texts to assess the data 105 eligibility. Articles were excluded based on the following 106 criteria: (1) not TBI or miRNA-related articles, (2) reviews, 107

meta-analyses, observational studies, or reports, (3) tissue or animal and cell experiments, (4) studies not published in English. Second, after full-text evaluation, articles were excluded if the severity of TBI was not determined with the GCS at all or did not meet a criterion of GCS score 13-15 that defines mTBI (Iankova 2006). Additionally, the articles with unclear regulation of miRNA (data constraints) or mixed mild to severe TBI cohorts, as well as those describing the longer post injury time (>15 days after the injury) or studies without a non-TBI cohort as a control group, were excluded (Fig. 1). The search strategy resulted in 8 articles included in the meta-analysis (Redell et al. 2010; Yang et al. 2016; Di Pietro et al. 2017; Qin et al. 2018; Sun et al. 2018; Yan et al. 2019; Hicks et al. 2020b; Tas et al. 2020).

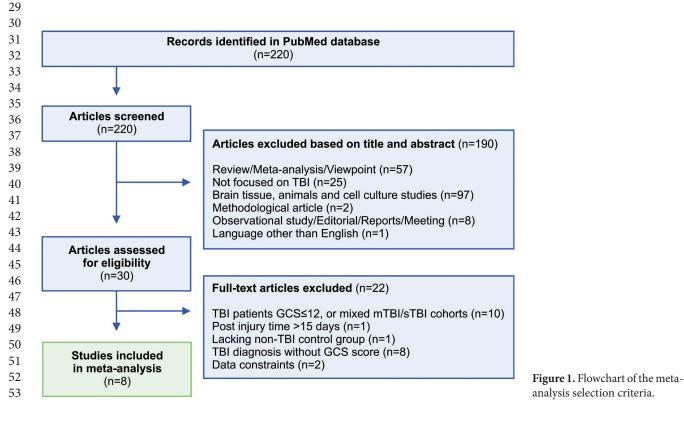
From the final set of selected studies, the following data of interest were extracted: study details (authors, publica-tion year), list of deregulated miRNAs, type of biofluid as a sample source, methodology used to detect miRNAs levels including the normalisation of the data, informa-tion about patients and control cohorts, GCS score of the patients, time of sample collection after the injury, p-value, and regulation status of miRNAs (upregulated or downregulated). Following the statistical method in the articles, a *p*-value <0.05 was considered significant while also taking to account the predictive models for miRNAs as mTBI diagnostic candidates. Curated source miRNA data including cohort details are summarized in Supplementary Material (Supplement 1).

miRNA:target identification

Prediction of gene targets for differentially expressed miR-NAs was performed using the mirDIP: microRNA Data Integration Portal (http://ophid.utoronto.ca/mirDIP) ver-sion 4.1.11.1 (Database version 4.1.0.3.), which integrates 30 different algorithms (Tokar et al. 2018). Identification of miRNA putative targets involved all 30 databases at very high confidence level (39 miRNAs), except for the 4 miR-NAs where a high score class filter was employed to query the predictions. The miR-12136 was the only miRNA that is not included in the mirDIP database and thus did not yield putative targets in any of the used resources. All raw data and search results are included in Supplement 2.

Comprehensive pathway enrichment

Putative target genes defined by mirDIP were used to iden-tify significantly enriched pathways performing a pathway enrichment analysis via pathDIP: pathway Data Integration Portal http://ophid.utoronto.ca/pathDIP version 4.0.21.2 (Database version 4.0.7.0) (Rahmati et al. 2020). We have used all 22 pathway sources and only known lit-erature curated (core) pathway memberships considering *q*-value <0.05 (false discovery rate: Benjamini-Hochberg method). Significantly enriched pathways were then pro-cessed to identify enriched terms in the pathway titles (word enrichment analysis; Supplement 3). To visualize



2

3

4

5

6

7

8

9

26

54 55

56

57

60

61

62

63

64

65

66

67

68

69

70

71

72

73

74

75

76

77

78

80

both common and unique terms we used NAViGaTOR ver. 3.0.11, highlighted terms of significantly enriched pathways (blue, purple and black refers to low-to-higher q-value) and produced the SVG file as output. The SVG file was post-processed in CorelDRAW X7 version 17.1.0.572 to its final form (Fig. 4B).

Protein-protein interactions

10 Identified gene targets unique for particular biofluid (Supple-11 ment 2) were used to query Integrated Interactions Database 12 (IID) (http://ophid.utoronto.ca/iid) version 2018-11 (Kotlyar 13 et al. 2019) to retrieve direct physical protein interactions 14 among them (Supplement 4). Using DisGeNet (Pinero et 15 al. 2017), edges were annotated with relevance to neurode-16 generative disease, nervous system disease, tauopathy and Alzheimer's disease (data integrated in IID). NAViGaTOR 17 was then used to visualize and analyse the resulting network. 18 19 To identify the most important proteins within the network, we computed centrality and node degree. Closeness central-20 21 ity provides ranking for each node based on the average 22 length of any breadth-first search path. The most central 23 proteins (and those with degree >20) were selected. To reduce 24 network complexity, other nodes and edges were removed, 25 resulting in the final network in Figure 5 (Supplement 4).

Network analysis and visualization

NAViGaTOR version 3.0.11 (Brown et al. 2009) was used to visualize and annotate miRNA:gene network and physical protein interaction network. Final graphs were exported 58 in SVG format, and completed in CorelDRAW X7 version 59 17.1.0.572 to produce the final images with legends.

Results

To identify signaling pathways associated with mTBI, we analysed the differentially expressed circulating miRNAs in mTBI with integrative bioinformatic tools. The analytic pipeline combined the comprehensive miRNA:gene target prediction (mirDIP), pathway enrichment analysis (path-DIP) and final integration with physical protein-protein interactions (IID) (Fig. 2A).

Following the selection criteria, the data for bioinformatic evaluation were extracted from 220 searched articles resulting in 8 studies focusing on altered miRNA levels in peripheral biofluids of subjects with mTBI. For a more detailed analysis, we separately examined the deregulated miRNAs based on their origin using three biofluid sources, i.e., serum, plasma, and saliva. In total, data for 12 different miRNAs were col-

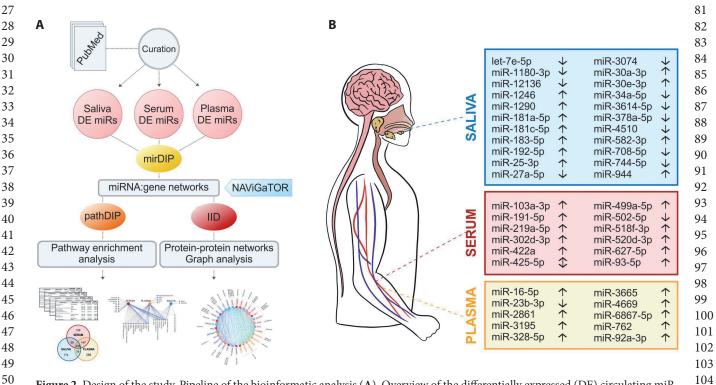


Figure 2. Design of the study. Pipeline of the bioinformatic analysis (A). Overview of the differentially expressed (DE) circulating miR-51 105 NAs following mild TBI evaluated in the study (B). Arrows in the panel B represent direction of deregulation in particular biofluid, as 52 106 reported in the original studies. mirDIP, microRNA Data Integration Portal; pathDIP, pathway Data Integration Portal; IID, Integrated 107 53 Interactions Database.

Table 1. Key deregulated circulating miRNAs involved in the peripheral signaling after mild traumatic brain injury (mTBI)

	.	0 0		
miRNA	Sequence (5'-3')	Regulation	Biofluid	Number of targeted genes
hsa-miR-103a-3p	AGCAGCAUUGUACAGGGCUAUGA	\uparrow		2087
hsa-miR-302d-3p	UAAGUGCUUCCAUGUUUGAGUGU	\uparrow	Serum	1673
hsa-miR-520d-3p	AAAGUGCUUCUCUUUGGUGGGU	\uparrow		1566
hsa-miR-93-5p	CAAAGUGCUGUUCGUGCAGGUAG	\uparrow		2540
hsa-miR-16-5p	UAGCAGCACGUAAAUAUUGGCG	\uparrow		2761
hsa-miR-23b-3p	AUCACAUUGCCAGGGAUUACCAC	\downarrow	Plasma	2284
hsa-miR-92a-3p	UAUUGCACUUGUCCCGGCCUGU	\uparrow		1238
hsa-let-7e-5p	UGAGGUAGGAGGUUGUAUAGUU	\downarrow		1387
hsa-miR-181a-5p	AACAUUCAACGCUGUCGGUGAGU	\uparrow		1597
hsa-miR-181c-5p	AACAUUCAACCUGUCGGUGAGU	\uparrow	Saliva	1798
hsa-miR-25-3p	CAUUGCACUUGUCUCGGUCUGA	\uparrow		1407
hsa-miR-34a-5p	UGGCAGUGUCUUAGCUGGUUGU	\downarrow		1330
hsa-miR-944	AAAUUAUUGUACAUCGGAUGAG	\uparrow		1178

1

19 lected in serum, 10 in plasma, and 22 in saliva (Fig. 2B). 20 These deregulated miRNAs served as peripheral indicators 21 of molecular regulatory mechanisms involved after the mTBI. 22 We identified the putative target genes for deregulated miR-23 NAs in all three biofluids using the mirDIP database portal, 24 and visualized the miRNA:gene networks illustrating the top genes targeted by the most of the deregulated miRNAs 25 26 in serum vs. saliva (Fig. 3A), serum vs. plasma (Fig. 3B) and 27 saliva vs. plasma (Fig. 3C). Combined, we predicted 9,903 28 target genes for deregulated serum miRNAs, 6,544 target 29 genes for plasma miRNAs, and 12,343 for salivary miRNAs.

30 Key involved miRNAs targeting the highest number of 31 genes in particular biofluids are summarized in Table 1, 32 suggesting their higher predictive value for peripheral 33 signaling after mTBI. Comprehensive lists of miRNA: gene targets for all three biofluids are available in Supplement 2. 34 35 Gene targets for each biofluid were separately analysed us-36 ing pathDIP portal to identify enriched pathways. Overall, 37 we identified 771 significantly enriched pathways in serum, 38 757 in plasma, and 710 in saliva after mTBI. Our analysis 39 revealed that majority of identified enriched pathways is 40 shared among the biofluids and only a fraction of enriched 41 pathways is unique to a particular biological fluid (Fig. 4A). Additionally, word enrichment analysis uncovered that most 42 43 of the highly ranked pathway terms (blue and purple edge 44 colour) are common among two or all three biofluids. The 45 top scored pathways associate with MAPK, TGF-β, WNT, TLR2/4, PI3K/AKT, insulin, and growth factor signaling. 46 47 Main mTBI-associated pathophysiological pathways are 48 listed in Table 2. However, we also identified enriched 49 pathways unique for each of the studied biofluids (Fig. 4B). 50 A comprehensive list of identified enriched pathways is 51 provided in Supplement 3.

52 To investigate the link between TBI and neurodegen-53 eration, we mapped identified gene targets of deregulated miRNAs (1,123 targets in serum, 1,007 in plasma and 1,792 in saliva) to proteins. Using IID we retrieved 52,605 direct, human physical protein-protein interactions among unique

Table 2. Representative pathophysiological pathways associated with mild traumatic brain injury (mTBI) identified by pathway enrichment analysis

Pathway name	Pathway source	q-value
PI3K/AKT/mTOR	ACSN2	6.08E-17
TGF-beta signaling	WikiPathways	2.35E-12
Starvation_autophagy	ACSN2	2.59E-11
Canonical WNT	ACSN2	3.10E-11
EGF/EGFR signaling	WikiPathways	4.46E-11
Insulin signaling	WikiPathways	6.34E-11
VEGFA-VEGFR2 signaling	WikiPathways	3.33E-09
BDNF signaling	WikiPathways	5.51E-09
MAPK signaling	KEGG	3.76E-08
FoxO signaling	KEGG	1.40E-07
Endocytosis	KEGG	5.15E-07
Regulation of actin cytoskeleton	KEGG	6.37E-07
Toll-like receptor signalling network	systems-biology.org	8.84E-07
Focal adhesion	WikiPathways	9.27E-07
Immunostimulatory core	ACSN2	3.42E-06
FGF signaling	Panther_Pathway	4.11E-06
Apoptosis	ACSN2	7.16E-06
IGF1R signaling cascade	REACTOME	2.03E-03
Growth factors signaling	ACSN2	6.95E-03
PTEN regulation	REACTOME	7.85E-03
Synaptic vesicle trafficking	Panther_Pathway	1.67E-02
IL-1 pathway	stke	2.26E-02

Literature curated (core) pathways considering *q*-value <0.05 using 106 the false discovery rate (FDR): Benjamini-Hochberg (BH) method. 107

5

54

71

72 73

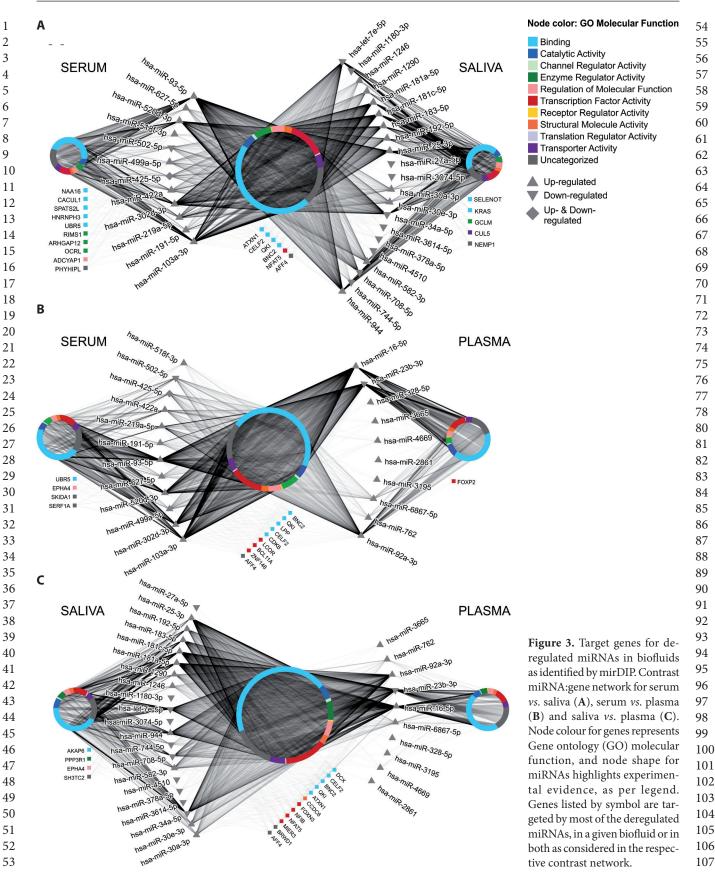
74

75

76

77

78



3

4

5

7

20

21

22

23

24

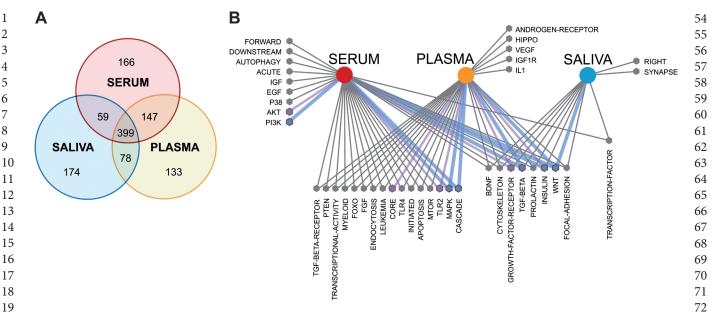


Figure 4. Pathway enrichment analysis using pathDIP. The number of identified enriched pathways and their overlap among the serum, plasma, and saliva (A). The most overrepresented enriched pathways show many shared terms. Edge colour and thickness represents significance of term enrichment (blue, purple, and black refers to low-to-higher q-value) (B).

25 targets across the biofluids. The resulting network was 26 further filtered by selecting only the interactions related to 27 neurodegeneration and focusing on central proteins within 28 the network. Final network comprised 33 key hub proteins 29 connected with interactions relevant in neurodegeneration, nervous system disease, tauopathy and Alzheimer's disease. 30 31 Furthermore, we highlighted in the network the deregulated 32 miRNAs that are linked to these key hub proteins associated 33 with neurodegeneration (Fig. 5). Interestingly, among the 34 interactions we identified several dominant connections that 35 link deregulated miRNAs and identified proteins. Specifi-36 cally, overrepresentation of the miR-93-5p, miR-302d-3p, 37 miR-520d-3p in serum, miR-16-5p, miR-23b-3p in plasma 38 and let-7e-5p, miR-34a-5p, miR-181a-5p, mir-181c-5p, 39 miR-183-5p, miR-744-5p in saliva with neurodegeneration-associated proteins was observed. These deregulated 40 41 miRNAs represent a group of candidate molecules linking the post-mTBI signaling reflected in peripheral fluids with 42 43 neurodegeneration-associated interaction pathways. Com-44 prehensive and final list of interacting proteins is attached 45 in the Supplement 4.

46

47 48

Discussion

49

50 We performed a systematic review of deregulated miRNA 51 profiles in peripheral fluids of individuals suffering mild 52 traumatic brain injury aiming to identify molecular signaling 53 associated with pathophysiology of mTBI. The meta-analysis specifically focused on studies that summarize the altered 78 miRNA level in serum, plasma, and saliva of mTBI individu-80 als. Following selection criteria, the search strategy identified 81 8 studies providing data on 44 deregulated miRNAs that 82 were further analysed by bioinformatic pipeline. Integrated 83 bioinformatic workflow enabled both increased coverage and 84 depth of our analysis by simultaneous querying of multiple 85 databases for more comprehensive prediction of putative 86 gene targets and higher confidence of the outcomes when 87 compared to recent analyses (Di Pietro et al. 2017, 2018; Atif 88 and Hicks 2019). 89

miRNA:gene networks

Highlighted subgroups of identified genes visualized in the 93 94 miRNA:gene networks (Fig. 3) represent a subset of genes targeted by the highest number of deregulated miRNAs 95 across all biofluids after mTBI. Moreover, one of the target 96 genes, CACUL1, was previously described as a target of 97 miR-219a-5p that has been found deregulated after TBI. 98 Experimental validation of CACUL1 expression in the 99 neuronal cell injury model revealed caspase-3-dependent 100 induction of apoptosis regulated by Akt/Foxo3a and p53/ 101 Bcl-2 signaling pathways (Yan et al. 2019). Upregulation of 102 another predicted target gene, GCLM, has been described 103 in the TBI mice model as one of the downstream genes 104 of Nrf2 signaling pathway, playing a neuroprotective role 105 following TBI (Dong et al. 2018). Furthermore, decreased 106 level of CUL5 as measured in rat cerebral cortex and hip-107

73

74

75

76

77

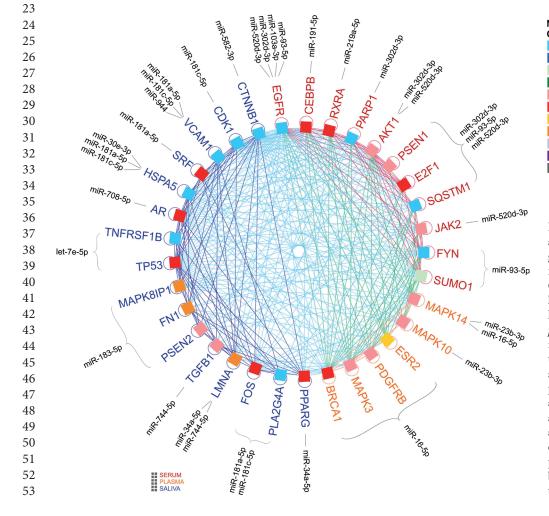
90

91

pocampus 7 days post-TBI was linked to the inhibition of 1 2 ubiquitin-proteasome system (Yao et al. 2006). The neu-3 roprotective effect of ADCYAP1, also known as pituitary 4 adenylate cyclase activating polypeptide (PACAP), has 5 been investigated in the rat model of diffuse axonal injury. 6 Intracerebroventricular administration of PACAP follow-7 ing TBI resulted in a reduction of beta-amyloid precursor 8 protein-immunopositive axons in the corticospinal tract 9 (Farkas et al. 2004; Tamas et al. 2006). Involvement of 10 EPHA4 has been observed in pro-inflammatory response 11 to TBI via regulation of Akt, mTOR, and NF-KB signaling 12 pathways (Kowalski et al. 2019).

13 Deregulation of miRNA levels in brain or peripheral fluids 14 was extensively studied in various animal models of TBI. De-15 spite a certain overlap of miRNA findings, the accumulated evidence indicates major difference between animal models 16 and human TBI studies (Pinchi et al. 2020; Herrold et al. 17 2021). Nevertheless, animal models still carry the translation 18 19 potential of research findings for diagnostic or therapeutic benefits in human medicine. In particular, upregulation of 20 21 2 miRNAs that are highly expressed in the brain, the miR- 9-3p and miR-136-3p was reported in plasma of TBI rats 54 and mTBI patients suggesting their biomarker potential 55 (Das Gupta et al. 2021). Other promising miRNAs, such as 56 miR-142-3p (Liu et al. 2014b), let-7i (Balakathiresan et al. 57 2012), miR-23b (Sun et al. 2018), miR-181a-5p and miR-58 191-5p (Weisz et al. 2020) were identified in both human 59 and animals, indicating shared post-concussive signaling 60 between TBI animal models and clinical studies. 61

Our findings suggest an association of identified genes 62 with development and functioning of central nervous system 63 indicating specific molecular signaling involved in mTBI. 64 Employing the integrative approach, we were able to predict 65 the gene candidates linked to the post-traumatic sequelae of 66 TBI depicting the brain injury pathways together with par-67 ticular healing processes. Our data are partially supported by 68 experimental evidence; however, vast majority of drawn gene 69 predictions is novel and remains to be explored by further 70 functional and clinical studies. Moreover, we hypothesize 71 that identified top ranked genes might represent a group of 72 potential novel biomarkers to be further explored in con-73 nection with TBI conditions. 74



Node color: GO Molecular Function Binding Catalytic Activity Channel Regulator Activity Enzyme Regulator Activity Regulation of Molecular Function Transcription Factor Activity Receptor Regulator Activity

Structural Molecule Activity Translation Regulator Activity

Transporter Activity
Uncategorized

87 88

75

76

77

78

80

81

82

83

84

85

86

89 90

Figure 5. Protein-protein inter-91 actions among predicted target 92 genes of miRNAs deregulated in 93 mTBI. Node colour represents 94 Gene Ontology (GO) molecu-95 lar function, and text colour highlights biofluid source, as 96 per legend. Edges represent 97 neurodegeneration-related in-98 teractions, while edge colour 99 signifies relationships among 100 proteins - dark blue edges are 101 among saliva targets, red edges 102 among serum targets, orange 103 among plasma targets, green 104 edges are between serum and 105 plasma targets, and turquoise 106 between serum/plasma and CSF 107 targets.

2

3 Interestingly, the majority of the signaling pathways as 4 derived from the deregulated peripheral miRNAs after 5 mTBI are shared among the investigated biofluids of serum, 6 plasma and saliva. According to these findings, dysregulated 7 circulating miRNAs target the genes implicated in the same 8 fundamental mechanisms simultaneously manifested in all 9 assessed biofluids. These data support the idea that different 10 biological fluids carry the molecular information on pathophysiological processes related to brain injury and might 11 help to better elucidate the gene signaling or predict targets 12 13 for therapeutical intervention of TBI.

14 Among the significantly enriched pathways the disturbance of hormone signaling represents the most prominent 15 feature identified by bioinformatic analysis. Posttraumatic 16 17 neuroendocrine dysfunction, e.g., hypopituitarism is a welldocumented consequence of TBI (Benvenga et al. 2000; Kelly 18 19 et al. 2000). Besides disturbances of other anterior pituitary 20 hormones, higher prolactin level has been reported in sol-21 diers who sustained blast-induced TBI (Baxter et al. 2013). 22 Likewise, mild hyperprolactinemia was observed in mild 23 to severe TBI patients (Aimaretti et al. 2004). A higher level 24 of prolactin following TBI can be explained by transport repression of prolactin inhibitory factor into the pituitary 25 26 gland (Scranton and Baskin 2015). The pituitary dysfunction 27 has been associated with a decreased serum level of insulin 28 like growth factor 1 (IGF-1), suspected to be also involved 29 in the progression of cognitive dysfunction (Bondanelli et al. 2004; Berg et al. 2010). Inversely, administration of IGF-1 30 31 leads to stimulation of anti-apoptotic pathways mediated by 32 PI3K/Akt and MAPK signals and prevents spatial memory 33 deficits following mTBI in mice (Rubovitch et al. 2010, 2011). 34 Stimulation of these cascades appears to be a promising 35 strategy positively affecting the recovery mechanisms such 36 as angiogenesis, anti-apoptotic survival, synaptic plasticity 37 and neurogenesis (Mangiola et al. 2015).

Findings identified by our analysis are further supported
by a recent study that revealed association of 5 deregulated
miRNAs with insulin, IGF-1R and TGF-β signaling in military personnel with history of mTBI (Devoto et al. 2020).

Pleiotropic cytokine, transforming growth factor-beta 42 43 (TGF- β), plays a role in neurogenesis, removal of amyloid- β protein and modulates the actions of various growth fac-44 tors (Devoto et al. 2020). The general effect of TGF- β in 45 TBI models remains questionable, but its neuroprotective 46 47 function relates to the restriction of chemokines produc-48 tion after brain injury (Dobolyi et al. 2012). Potential 49 pro-apoptotic function of TGF-\u03b3 through activation of 50 the caspase-3 in rats with induced mTBI was also reported 51 (Patel et al. 2017).

52 The enrichment of TGF- β signaling, as reflected by all 53 examined biofluids in our study, supports the idea that this 9

pathway plays an important role in post-concussive signaling54and might represent a viable candidate for management of55post-traumatic sequelae in mTBI patients.56

Another highly over-represented class of signaling path-57 ways identified by our meta-analysis includes the group of 58 59 various growth factors and receptors. One of the identified enriched signaling pathways, the vascular endothelial growth 60 factor (VEGF), was previously reported in connection with 61 angiogenesis, neurogenesis and neuroprotection in TBI 62 models (Nag et al. 1997; Sun et al. 2003; Thau-Zuchman et 63 al. 2010; Lu et al. 2019). A higher concentration of VEGF 64 has been also reported in humans suffering traumatic brain 65 injury with various severities (Mellergard et al. 2010; Helmy 66 et al. 2011; Meabon et al. 2020). Recently, elevated plasma 67 levels of VEGF, together with IL-6 and TNF-a were associ-68 ated with CT lesions in patients following mTBI. It seems 69 70 that a combination of these three molecules could represent 71 a biomarker panel to distinguish mTBI cases with CT findings from mTBI without CT findings (Edwards et al. 2020b). 72

73 TBI was shown to evoke the sensitivity of cells to epidermal growth factor (EGF) cascade via increase of EGFR levels 74 (Addington et al. 2015). Furthermore, neuroprotective effect 75 76 of EGF in post-concussive period is mediated by stimulation of astroglial cells, which leads to improved cognitive func-77 tion and reduced neuronal loss (Sun et al. 2010). Similarly 78 to EGF, the pro-survival function of fibroblast growth factor 80 (FGF) relates to its mitogen activity restricting the apoptosis 81 and necrosis via inhibition of autophagy (Addington et al. 82 2015; Tang et al. 2017). A study on patients also confirmed 83 the role of FGF in neurogenesis and protection of blood 84 brain barrier (Chaban et al. 2020). Brain-derived neuro-85 trophic factor (BDNF) plays a regulatory role in neuronal 86 survival, neurogenesis, and synaptic plasticity (Martinowich 87 and Lu 2008). This effect is mediated via binding to TrkB 88 receptor and activation of three signaling pathways: PI3K, 89 MAPK/ERK, and PLCy (Chao 2003). The elevated plasma 90 level of BDNF was previously observed in children with 91 a concussion suggesting its potential use as a marker of 92 mild head trauma. However, the BDNF levels did not dif-93 ferentiate the severe TBI with loss of consciousness from 94 mTBI without loss of consciousness suggesting that BDNF 95 increase indicates rather functional disturbance related to 96 TBI as such and is not related to the severity of head trauma 97 98 (Tylicka et al. 2020).

Growth factor signaling is associated with another im-99 portant cascade of PI3K/Akt/mTOR. When activated, PI3K 100 stimulates Akt serine/threonine kinase 1 (Akt) to trigger 101 mTOR pathway that plays a crucial role in functional re-102 covery process following a traumatic CNS injury (Don et 103 al. 2012). Regulation of mTOR pathway via suppression of 104 PTEN activates PI3K/Akt signaling leading to neuroprotec-105 tion through regulation of apoptosis-related proteins (Ge et 106 al. 2014; Han et al. 2014). 107

1 Neuroinflammation represents an important mechanism 2 in pathophysiology of TBI leading to secondary brain dam-3 age in post-traumatic period (Singh et al. 2016; Sun et al. 4 2019). Our results from bioinformatic analysis agree with 5 previously identified molecules of inflammatory pathways 6 activated in response to TBI, e.g., IL-1, TLR2 and TLR4. 7 Pro-inflammatory cytokine, interleukin 1 (IL-1), is an im-8 portant mediator of neuroinflammatory response after TBI 9 (Basu et al. 2004). Both IL-1 α and IL-1 β have been described 10 previously as rapidly elevated after brain injury (Griffin et 11 al. 1994; Fan et al. 1995). Furthermore, the genetic ablation of IL-1 receptor showed a more positive effect on cognitive 12 function in TBI mice models than ablation of IL-1 α and 13 14 IL-1 β alone (Newell et al. 2018). High level of inflammatory cytokines including IL-1β, IL-6 and CCL2 was acutely 15 elevated in mTBI patients relative to controls. However, the 16 17 elevated CCL2 level was also associated with greater severity of post-concussion symptoms negatively affecting the 18 19 outcome of mTBI patients (Sun et al. 2019). These observa-20 tions highlight the deleterious effect of pro-inflammatory 21 signaling in post-injury period when persisting chronically.

22 Toll-like receptors (TLR) represent another group of 23 highly enriched signaling pathways identified by our analy-24 sis. TLRs have been recognized as essential players of innate 25 immune signaling and regulation of inflammatory responses 26 (Kawai and Akira 2006). Even in the post-TBI conditions 27 activation of TLR 2/4 involves signaling via MAPKs and 28 subsequent stimulation of NF-KB leading to expression of 29 inflammation responsive genes (Downes and Crack 2010; 30 Ved et al. 2021). TLR2/4-mediated secondary brain injury 31 was observed proposing their role in neuroinflammation 32 and cell death (Krieg et al. 2017), while the TLR4/MyD88/ 33 NF-kB signaling pathway might be efficiently targeted by 34 PACAP reducing the secondary inflammatory response and 35 neuronal death after TBI (Mao et al. 2012).

Metabolic disturbance is another invariant feature ofpost-concussive condition in TBI patients.

38 Studies have shown occurrence of hyperglycolysis early 39 after injury, followed by a hypometabolic phase lasting for days (Yoshino et al. 1991; Bergsneider et al. 2001). A high 40 41 glucose level during the first hours after TBI can predict 42 a poor neurologic outcome and mortality rate (Liu-DeRyke 43 et al. 2009; Terzioglu et al. 2015). Additionally, the association between diabetes and increased risk of neurodegeneration 44 developed due head trauma has been reported in veterans 45 with type 2 diabetes and a history of TBI (Zimering et al. 46 47 2019). Possible molecular explanation suggests that elevated 48 pro-inflammatory cytokines inhibit insulin receptor signal-49 ing by targeting the Akt that negatively affects the neuronal 50 energy metabolism. Furthermore, insulin resistance fol-51 lowing TBI deprives this neuroprotective pathway and thus 52 renders the individuals vulnerable to neurodegeneration 53 (Karelina and Weil 2016).

Matyasova et al.

83

84

85

WNT signaling is involved in several critical processes 54 in the brain, including cell proliferation, differentiation, 55 inflammation, neurogenesis, synaptogenesis, axon guid-56 ance, neuron maintenance, and regeneration (Salinas 2012; 57 Marchetti and Pluchino 2013). Various studies implicate the 58 59 WNT pathway as a neuroprotective cascade that upon stimulation might represent a prospective treatment of post-TBI 60 conditions (Zhao et al. 2016; Zhang et al. 2018). Intranasal 61 administration of recombinant Wnt3a in TBI mice model 62 during first 24 hours was shown to inhibit the autophagy and 63 upregulate the expression of growth factors including GDNF 64 and VEGF responsible for neurogenesis and angiogenesis, 65 respectively. In our meta-analysis, WNT pathway has been 66 the top ranked prediction in all examined fluids, suggesting 67 a key role of WNT pathway in post-traumatic signaling and 68 recovery. 69

Previous studies used various bioinformatic approaches to 70 identify signaling pathways linked to deregulated miRNAs 71 after TBI. However, the published pathways were derived 72 73 from a single database, e.g., KEGG or Ingenuity Pathway analysis, respectively, leading to only partial results due to 74 small overlap across pathway databases (Rahmati et al. 2017). 75 Despite this methodological limitation the mTOR, TGF-beta, 76 IGF1R, neurotrophin and inflammatory signaling represent 77 major overlap with findings of our meta-analysis and sup-78 port their involvement in post mTBI conditions by multiple 80 evidence (Qin et al. 2018; Atif and Hicks 2019; Devoto et al. 81 2020; Hicks et al. 2020a; Di Pietro et al. 2021). 82

Link between mTBI and neurodegeneration

The history of mTBI, and especially the repetitive head 86 traumas are associated with increased risk of development 87 of neurodegenerative disorders (McKee and Robinson 2014; 88 Manley et al. 2017; Alosco et al. 2018). To elucidate the 89 relationship between prior TBI with subsequent dementia 90 we tested the hypothesis how molecular signaling identified 91 after mTBI relates to the protein interactions seen in neuro-92 degeneration including Alzheimer's disease and tauopathies. 93 Bioinformatic analysis revealed group of genes targeted by 94 deregulated miRNAs after mTBI with a known interaction 95 in neurodegenerative conditions. Within this subpopulation 96 several central proteins implicated in the neurodegenerative 97 disorders were identified. In particular, the Alzheimer's dis-98 ease-associated proteins, presenilin 1 (PSEN1) and presenilin 99 2 (PSEN2) showed major interactions within the network. 100

PSEN1 viaincrease of cytoplasmic CTNNB1, another101identified key hub protein within the network, negatively102regulates the WNT and Notch signaling (Murayama et al.1031998; Marambaud et al. 2002). Since these pathways play an104important role in neuroregeneration and brain homeostasis,105downregulation of this signaling by TBI might negatively106affect the process of recovery and contribute to the longi-107

tudinal sequelae of TBI, which might eventually progress 1 2 towards neurodegeneration. Furthermore, targeting of 3 gamma-secretase complex including presenilin to reduce 4 amyloid beta production represents a perspective therapeutic 5 strategy for treatment of Alzheimer's disease (De Strooper 6 et al. 2012), and might be potentially employed also for the 7 attenuation of neurotrauma following TBI (Mannix et al. 8 2011; Lin et al. 2017).

9 In addition, among the key hub proteins identified 10 within the protein interaction network, there are several other candidates that have been previously reported in 11 connection with TBI. Up-regulation of AKT1 (Edwards et 12 13 al. 2020a), FOS (Wang et al. 2014), FYN (Liu et al. 2014a), 14 MAPK3 (Ou et al. 2014), CEBPB (Sandhir and Berman 15 2010), PLA2G4A (Sarkar et al. 2020), RXRA (Zhao et al. 16 2019), E2F1 (Liu et al. 2013), HSPA5 (Truettner et al. 2007) 17 and downregulation of AR (Golz et al. 2019), MAPK10 18 (Long et al. 2013) was observed in various animal TBI 19 models or human TBI cases. While the increase of HSPA5 20 is thought to be neuroprotective in neurodegeneration 21 (Casas 2017), the activation of E2F1 is associated with post-22 traumatic neuronal apoptosis (Liu et al. 2013). Activation 23 of JAK2-STAT3 pathway together with upregulation of 24 CEBPB couples the activation of astrocytes with growth 25 factor and neuroinflammatory responses after TBI (Oliva 26 et al. 2012). Along with these findings the TNFRSF1B and 27 SRF-deficient mice exhibit poorer outcome following TBI 28 (Yang et al. 2010; Forstner and Knoll 2020). On the other 29 side experimental administration of EGF (Sun et al. 2010) 30 and FN1 (Tate et al. 2007) in animal TBI models suggests 31 their neuroprotective role. Furthermore, the upregulation 32 of CDK1 by dexmedetomidine (Wu et al. 2018; Yang et al. 33 2021) and inhibition of PARP1 (Stoica et al. 2014) in TBI 34 models also appears to be viable neuroprotective strategy.

Our study also identified a subgroup of proteins, e.g.,
BRCA1, LMNA and ESR2 that have not been associated
with TBI so far, and thus might represent novel candidates
for validation in TBI conditions.

39 Presence of validated targets within the protein interac-40 tion network supports the findings of integrated bioinfor-41 matic analysis with experimental evidence. Interestingly, 42 the interaction network of neurodegeneration-associated 43 proteins highlights the TGF-β, AKT1, MAPK, TP53, TNF 44 and growth factor signaling that were also identified among the pathways enriched in mTBI. This overlap indicates 45 overrepresentation of signaling related to these proteins 46 47 and implicates their central role in molecular regulatory 48 mechanisms shared between mTBI and neurodegeneration.

To our knowledge this is the first integrative bioinformatic analysis systematically combining comprehensive miRNA-target prediction, multiple pathway sources, and rich annotated physical protein interactions aiming to identify peripheral microRNA signaling associated with mild traumatic brain injury. Results of the integrative 54 analysis reveal specific signaling pathways and highlight 55 the key enriched molecular mechanisms reflected across the 56 serum, plasma and saliva of individuals after mTBI. Find-57 ings of the meta-analysis suggest that peripheral miRNAs 58 could represent relevant biomarkers with added value to 59 the currently used diagnostic approaches and prognostics 60 of recovery after TBI. 61

Strengths and limitations

Despite the supportive data for our findings in the scientific 65 literature we assessed the risk of bias, among which the input 66 microRNA data represent reasonable source of variability 67 to be considered. In particular, various methodological ap-68 proaches used for the measurement of miRNA, such as 69 next-generation sequencing, real-time PCR or microarray 70 techniques, provide a variable depth of information and 71 sensitivity of detection for evaluated miRNAs. Furthermore, 72 73 many studies lack the sample quality check or provide no statement about exclusion of low-quality samples prior 74 analysis (e.g., hemolysis for plasma, contamination of sa-75 liva with cellular RNA). Third, use of different endogenous 76 calibrators and normalization procedures for calculation of 77 miRNA fold change has to be considered as it has a direct 78 impact on accuracy of reported miRNA expression values. 80 Fourth, variable statistical methods used in the research 81 articles or lacking validation step of primary data obtained 82 by whole transcriptome analysis might result in distinct 83 pattern of false positives and negatives affecting the inclu-84 sion of miRNAs in the integrated analyses. We summarize 85 these issues to be critically considered by the readers during 86 interpretation of our findings. 87

On the other hand, the process of data curation accounted 88 for multiple variable factors and data constraints providing 89 more consistent data input when compared to previous meta-90 analyses. Evaluation of the data under the strict statistical 91 and predictive parameters supports obtained findings at high 92 confidence level. Moreover, we consider the use of integrative 93 bioinformatic pipeline as a major advantage of the employed 94 meta-analytic approach since it enables the identification of 95 enriched signaling pathways and molecular mechanisms 96 based on cumulative support from multiple database sources 97 and thus to a substantial degree corrects for the variabilities 98 99 of input experimental data.

Conclusions

Integrated bioinformatic analysis revealed enrichment of
common signaling pathways involved in the post-concussive104
105conditions as reflected by the profile of circulating micro-
RNAs in various body fluids. Since the identified pathways107

62

63

64

100

101

102

substantially overlap among the biofluids we hypothesize that 1 2 the specific signaling associated with mTBI is concomitantly 3 reflected in various biological fluids. Furthermore, our data 4 indicate that biofluid-specific microRNA profiles could 5 serve for detailed characterization of molecular pathways in-6 volved in the pathophysiology and healing processes of mild 7 traumatic brain injury. Top hub proteins identified within 8 the network represent novel putative candidates involved 9 in molecular signaling after mTBI. Our findings implicate 10 a molecular link between mTBI signaling and neurodegener-11 ation-associated interaction pathways. However, to validate the concept of mTBI-induced neurodegeneration involving 12 13 predicted targets further experimental research is necessary.

14 Acknowledgement. Bioinformatic analyses were in part supported 15 by the Ministry of Education of the Slovak Republic, awarded to 16 BrainTest, s.r.o. within the research scheme "Stimuli for research 17 and development" and research grants VEGA 2/0118/19, VEGA 18 2/0154/19 and APVV-17-0668, APVV-19-0568, APVV-20-0615. 19 We are grateful to Zlatica Cerna for her help with graphical design 20 and artistic illustration. Development and maintenance of used 21 tools and databases were supported in part by funding from Natural 22 Sciences Research Council (NSERC #203475), Canada Foundation 23 for Innovation (CFI #225404, #30865), Ontario Research Fund 24 (RDI #34876), IBM and Ian Lawson van Toch Fund. The funders 25 had no role in study design, data collection and analysis, decision to publish, or preparation of the manuscript. 26

Conflict of interest. The authors declare no conflict of interest.

References

27

28

29

30

31

- Addington CP, Roussas A, Dutta D, Stabenfeldt SE (2015): Endogenous repair signaling after brain injury and complementary bioengineering approaches to enhance neural regeneration. Biomark Insights 10, 43-60 https://doi.org/10.4137/BMI.S20062
- Aimaretti G, Ambrosio MR, Di Somma C, Fusco A, Cannavo S, Gasperi M, Scaroni C, De Marinis L, Benvenga S, degli Uberti EC et al. (2004): Traumatic brain injury and subarachnoid haemorrhage are conditions at high risk for hypopituitarism: screening study at 3 months after the brain injury. Clin. Endocrinol. (Oxf). 61, 320-326
- 42 https://doi.org/10.1111/j.1365-2265.2004.02094.x
- Alosco ML, Koerte IK, Tripodis Y, Mariani M, Chua AS, Jarnagin J,
 Rahimpour Y, Puzo C, Healy RC, Martin B et al. (2018): White
 matter signal abnormalities in former National Football League
 players. Alzheimers Dement. (Amst). 10, 56-65
 https://doi.org/10.1016/j.dadm.2017.10.003
- Atif H, Hicks SD (2019): A review of microRNA biomarkers in traumatic brain injury. J. Exp. Neurosci. 13, 1179069519832286
 https://doi.org/10.1177/1179069519832286
- Bailes JE, Petraglia AL, Omalu BI, Nauman E, Talavage T (2013):
 Role of subconcussion in repetitive mild traumatic brain injury.
- 52 J. Neurosurg. **119**, 1235-1245
- 53 https://doi.org/10.3171/2013.7.JNS121822

Balakathiresan N, Bhomia M, Chandran R, Chavko M, McCarron
RM, Maheshwari RK (2012): MicroRNA let-7i is a promising
serum biomarker for blast-induced traumatic brain injury. J.
Neurotrauma 29 , 1379-1387
https://doi.org/10.1089/neu.2011.2146
Basu A, Krady JK, Levison SW (2004): Interleukin-1: a master
regulator of neuroinflammation. J. Neurosci. Res. 78 , 151-156
https://doi.org/10.1002/jnr.20266
Baxter D, Sharp DJ, Feeney C, Papadopoulou D, Ham TE, Jilka
S, Hellyer PJ, Patel MC, Bennett AN, Mistlin A et al. (2013): Pituitary dysfunction after blast traumatic brain injury: The
UK BIOSAP study. Ann. Neurol. 74, 527-536
https://doi.org/10.1002/ana.23958
Benvenga S, Campenni A, Ruggeri RM, Trimarchi F (2000): Clini-
cal review 113: Hypopituitarism secondary to head trauma. J.
Clin. Endocrinol. Metab. 85 , 1353-1361
https://doi.org/10.1210/jcem.85.4.6506
Berg C, Oeffner A, Schumm-Draeger PM, Badorrek F, Brabant G,
Gerbert B, Bornstein S, Zimmermann A, Weber M, Broecker-
Preuss M et al. (2010): Prevalence of anterior pituitary dysfunc-
tion in patients following traumatic brain injury in a German
multi-centre screening program. Exp. Clin. Endocrinol. Dia-
betes 118 , 139-144
https://doi.org/10.1055/s-0029-1225611
Bergsneider M, Hovda DA, McArthur DL, Etchepare M, Huang
SC, Sehati N, Satz P, Phelps ME, Becker DP (2001): Metabolic
recovery following human traumatic brain injury based on
FDG-PET: time course and relationship to neurological dis-
ability. J. Head Trauma Rehabil. 16, 135-148
https://doi.org/10.1097/00001199-200104000-00004
Bondanelli M, De Marinis L, Ambrosio MR, Monesi M, Valle D,
Zatelli MC, Fusco A, Bianchi A, Farneti M, degli Uberti EC
(2004): Occurrence of pituitary dysfunction following trau-
matic brain injury. J. Neurotrauma 21, 685-696
https://doi.org/10.1089/0897715041269713
Brown KR, Otasek D, Ali M, McGuffin MJ, Xie W, Devani B, Toch
IL, Jurisica I (2009): NAViGaTOR: Network Analysis, Visuali-
zation and Graphing Toronto. Bioinformatics 25, 3327-3329
https://doi.org/10.1093/bioinformatics/btp595
Bryant RA, Harvey AG (1998): Relationship between acute stress
disorder and posttraumatic stress disorder following mild
traumatic brain injury. Am. J. Psychiatry 155 , 625-629
https://doi.org/10.1176/ajp.155.5.625
Casas C (2017): GRP78 at the Centre of the Stage in cancer and
neuroprotection. Front Neurosci. 11 , 177
https://doi.org/10.3389/fnins.2017.00177
Cassidy JD, Carroll LJ, Peloso PM, Borg J, von Holst H, Holm L, Kraus J, Coronado VG, et al. (2004): Incidence, risk factors
and prevention of mild traumatic brain injury: results of the WHO Collaborating Centre Task Force on Mild Traumatic
Brain Injury. J. Rehabil. Med. 43 , 28-60
https://doi.org/10.1080/16501960410023732
Chaban V, Clarke GJB, Skandsen T, Islam R, Einarsen CE, Vik A,
Damas JK, Mollnes TE, Haberg AK, Pischke SE (2020): Systemic
inflammation persists the first year after mild traumatic brain
injury: results from the prospective trondheim mild traumatic
brain injury study. J. Neurotrauma 37 , 2120-2130

https://doi.org/10.1089/neu.2019.6963

107

1	Chao MV (2003): Neurotrophins and their receptors: a conver-
2	gence point for many signalling pathways. Nat. Rev. Neurosci.
3	4, 299-309
4	https://doi.org/10.1038/nrn1078
5	Das Gupta S, Ciszek R, Heiskanen M, Lapinlampi N, Kukkonen J,
6	Leinonen V, Puhakka N, Pitkanen A (2021): Plasma miR-9-3p
7	and miR-136-3p as potential novel diagnostic biomarkers for
	experimental and human mild traumatic brain injury. Int. J.
8	Mol. Sci. 22, 1563
9	https://doi.org/10.3390/ijms22041563
10	De Strooper B, Iwatsubo T, Wolfe MS (2012): Presenilins and
11	gamma-secretase: structure, function, and role in Alzheimer
12	disease. Cold Spring Harb. Perspect. Med. 2, a006304
13	https://doi.org/10.1101/cshperspect.a006304
14	Devoto C, Lai C, Qu BX, Guedes VA, Leete J, Wilde E, Walker WC,
15	Diaz-Arrastia R, Kenney K, Gill J (2020): Exosomal microR-
16	NAs in military personnel with mild traumatic brain injury:
	preliminary results from the chronic effects of neurotrauma
17	Consortium Biomarker Discovery project. J. Neurotrauma
18	37, 2482-2492
19	https://doi.org/10.1089/neu.2019.6933
20	Dewan MC, Rattani A, Gupta S, Baticulon RE, Hung YC, Punchak
21	M, Agrawal A, Adeleye AO, Shrime MG, Rubiano AM et al.
22	(2019): Estimating the global incidence of traumatic brain
23	injury. J. Neurosurg. 130, 1080-1097
24	https://doi.org/10.3171/2017.10.JNS17352
25	https://doi.org/10.1136/bjsports-2020-103274
26	Di Pietro V, Ragusa M, Davies D, Su Z, Hazeldine J, Lazzarino G, Hill
27	LJ, Crombie N, Foster M, Purrello M et al. (2017): MicroRNAs
28	as novel biomarkers for the diagnosis and prognosis of mild and
	severe traumatic brain injury. J. Neurotrauma 34, 1948-1956
29	https://doi.org/10.1089/neu.2016.4857
30	Di Pietro V, Yakoub KM, Scarpa U, Di Pietro C, Belli A (2018):
31	MicroRNA signature of traumatic brain injury: from the bio-
32	marker discovery to the point-of-care. Front. Neurol. 9, 429
33	https://doi.org/10.3389/fneur.2018.00429
34	Di Pietro V, O'Halloran P, Watson CN, Begum G, Acharjee A,
35	Yakoub KM, Bentley C, Davies DJ, Iliceto P, Candilera G et
36	al. (2021): Unique diagnostic signatures of concussion in the
37	saliva of male athletes: the Study of Concussion in Rugby Union
38	through MicroRNAs (SCRUM). Br. J. Sports Med. (in press)
39	https://doi.org/10.1136/bjsports-2020-103274
	Dobolyi A, Vincze C, Pal G, Lovas G (2012): The neuroprotective
40	functions of transforming growth factor beta proteins. Int.
41	J. Mol. Sci. 13, 8219-8258
42	https://doi.org/10.3390/ijms13078219
43	Don AS, Tsang CK, Kazdoba TM, D'Arcangelo G, Young W, Zheng
44	XF (2012): Targeting mTOR as a novel therapeutic strategy
45	for traumatic CNS injuries. Drug Discov. Today 17, 861-868
46	https://doi.org/10.1016/j.drudis.2012.04.010
47	Dong W, Yang B, Wang L, Li B, Guo X, Zhang M, Jiang Z, Fu J, Pi
48	J, Guan D et al. (2018): Curcumin plays neuroprotective roles
49	against traumatic brain injury partly via Nrf2 signaling. Toxicol.
50	Appl. Pharmacol. 346 , 28-36
	https://doi.org/10.1016/j.taap.2018.03.020
51 52	Downes CE, Crack PJ (2010): Neural injury following stroke: are
52	Toll-like receptors the link between the immune system and
53	the CNS? Br. J. Pharmacol. 160, 1872-1888

Edwards KA, Motamedi V, Osier ND, Kim HS, Yun S, Cho YE,	55
Lai C, Dell KC, Carr W, Walker P et al. (2020a): A moderate	56
blast exposure results in dysregulated gene network activity	57
related to cell death, survival, structure, and metabolism. Front.	58
Neurol. 11, 91	59
https://doi.org/10.3389/fneur.2020.00091	60
Edwards KA, Pattinson CL, Guedes VA, Peyer J, Moore C, Davis	61
T, Devoto C, Turtzo LC, Latour L, Gill JM (2020b): Inflamma-	62
tory cytokines associate with neuroimaging after acute mild traumatic brain injury. Front. Neurol. 11 , 348	63
https://doi.org/10.3389/fneur.2020.00348	64
Fan L, Young PR, Barone FC, Feuerstein GZ, Smith DH, McIntosh	
TK (1995): Experimental brain injury induces expression of	65
interleukin-1 beta mRNA in the rat brain. Brain Res. Mol.	66
Brain Res. 30 , 125-130	67
https://doi.org/10.1016/0169-328X(94)00287-O	68
Farkas O, Tamas A, Zsombok A, Reglodi D, Pal J, Buki A, Lengvari	69
I, Povlishock JT, Doczi T (2004): Effects of pituitary adenylate	70
cyclase activating polypeptide in a rat model of traumatic brain	71
injury. Regul. Pept. 123 , 69-75	72
https://doi.org/10.1016/j.regpep.2004.05.014	73
Forstner P, Knoll B (2020): Interference of neuronal activity-	74
mediated gene expression through serum response factor	75
deletion enhances mortality and hyperactivity after traumatic	76
brain injury. FASEB J. 34, 3855-3873	77
https://doi.org/10.1096/fj.201902257RR	78
Gallant C, Barry N, Good D (2018): Physiological underarousal as	80
a mechanism of aggressive behavior in university athletes with	
a history of concussion. Brain Behav. 8, e01038	81
https://doi.org/10.1002/brb3.1038	82
Gardner RC, Byers AL, Barnes DE, Li Y, Boscardin J, Yaffe K (2018):	83
Mild TBI and risk of Parkinson disease: A chronic effects of	84
neurotrauma consortium study. Neurology 90, e1771-e1779	85
https://doi.org/10.1212/WNL.00000000005522	86
Ge XT, Lei P, Wang HC, Zhang AL, Han ZL, Chen X, Li SH, Jiang	87
RC, Kang CS, Zhang JN (2014): miR-21 improves the neurologi-	88
cal outcome after traumatic brain injury in rats. Sci. Rep. 4, 6718	89
https://doi.org/10.1038/srep06718	90
Gilbert KS, Kark SM, Gehrman P, Bogdanova Y (2015): Sleep	91
disturbances, TBI and PTSD: Implications for treatment and	92
recovery. Clin. Psychol. Rev. 40, 195-212	93
https://doi.org/10.1016/j.cpr.2015.05.008 Golz C, Kirchhoff FP, Westerhorstmann J, Schmidt M, Hirnet T,	94
Rune GM, Bender RA, Schafer MKE (2019): Sex hormones	95
modulate pathogenic processes in experimental traumatic brain	96
injury. J. Neurochem. 150, 173-187	
https://doi.org/10.1111/jnc.14678	97
Griffin WS, Sheng JG, Gentleman SM, Graham DI, Mrak RE,	98
Roberts GW (1994): Microglial interleukin-1 alpha expres-	99
sion in human head injury: correlations with neuronal and	100
neuritic beta-amyloid precursor protein expression. Neurosci.	101
Lett. 176, 133-136	102
https://doi.org/10.1016/0304-3940(94)90066-3	103
Han Z, Chen F, Ge X, Tan J, Lei P, Zhang J (2014): miR-21 alleviated	104
apoptosis of cortical neurons through promoting PTEN-Akt	105
signaling pathway in vitro after experimental traumatic brain	106
injury. Brain Res. 1582, 12-20	107

https://doi.org/10.1111/j.1476-5381.2010.00864.x

55

56

57 58

59

60

61

62

63

64

65

66

67 68 69

70

71

72 73

74

75

76

77 78 80

81

82

83

84

85

86 87

88

89

90

91 92

93

94

95

96 97

98

99

100 101

102

103

104

105

106

107

1	https://doi.org/10.1016/j.brainres.2014.07.045	https://doi.org/10.1186/s12974-019-1605-2
2	Helmy A, Carpenter KL, Menon DK, Pickard JD, Hutchinson PJ	Krieg SM, Voigt F, Knuefermann P, Kirschning CJ, Plesnila N, Rin-
3	(2011): The cytokine response to human traumatic brain in-	gel F (2017): Decreased secondary lesion growth and attenuated
	jury: temporal profiles and evidence for cerebral parenchymal	immune response after traumatic brain injury in Tlr2/4(-/-)
4	production. J. Cereb. Blood Flow Metab. 31, 658-670	mice. Front. Neurol. 8, 455
5	https://doi.org/10.1038/jcbfm.2010.142	https://doi.org/10.3389/fneur.2017.00455
6	Herrold AA, Kletzel SL, Foecking EM, Saban KL, Przybycien-Szy-	Li Y, Li X, Zhang S, Zhao J, Zhu X, Tian G (2017): Head injury
7	manska MM, Zilliox M, Bhaumik D, Lange D, Radke JR, Salinas	as a risk factor for dementia and Alzheimer's disease: A sys-
8		
9	I et al. (2021): miRNAs as potential biomarkers for traumatic	tematic review and meta-analysis of 32 observational studies.
	brain injury: pathway from diagnosis to neurorehabilitation.	PLoS One 12 , e0169650
10	J. Head Trauma Rehabil. 36, E155-E169	https://doi.org/10.1371/journal.pone.0169650
11	https://doi.org/10.1097/HTR.000000000000632	Lin HJ, Hsu CC, Chio CC, Tian YF, Lin MT, Lin TW, Chang CH,
12	Hicks SD, Olympia RP, Onks C, Kim RY, Zhen KJ, Fedorchak G,	Chang CP (2017): Gamma-secretase inhibitors attenuate neu-
13	DeVita S, Rangnekar A, Heller M, Zwibel H et al. (2020a):	rotrauma and neurogenic acute lung injury in rats by rescuing
	Saliva microRNA biomarkers of cumulative concussion. Int.	the accumulation of hypertrophic microglia. Cell Physiol.
14	J. Mol. Sci. 21, 7758	Biochem. 44, 1726-1740
15	https://doi.org/10.3390/ijms21207758	https://doi.org/10.1159/000485778
16	Hicks SD, Onks C, Kim RY, Zhen KJ, Loeffert J, Loeffert AC,	Liu-DeRyke X, Collingridge DS, Orme J, Roller D, Zurasky J,
17		
18	Olympia RP, Fedorchak G, DeVita S, Rangnekar A et al.	Rhoney DH (2009): Clinical impact of early hyperglycemia
	(2020b): Diagnosing mild traumatic brain injury using saliva	during acute phase of traumatic brain injury. Neurocrit. Care
19	RNA compared to cognitive and balance testing. Clin. Transl.	11, 151-157
20	Med. 10 , e197	https://doi.org/10.1007/s12028-009-9228-6
21	https://doi.org/10.1002/ctm2.197	Liu DZ, Sharp FR, Van KC, Ander BP, Ghiasvand R, Zhan X,
22	Iankova A (2006): The Glasgow Coma Scale: clinical application in	Stamova B, Jickling GC, Lyeth BG (2014a): Inhibition of SRC
23	emergency departments. Emerg. Nurse 14, 30-35	family kinases protects hippocampal neurons and improves
	https://doi.org/10.7748/en2006.12.14.8.30.c4221	cognitive function after traumatic brain injury. J. Neurotrauma
24	Iverson GL, Lovell MR, Smith S, Franzen MD (2000): Prevalence	31, 1268-1276
25	of abnormal CT-scans following mild head injury. Brain Inj.	https://doi.org/10.1089/neu.2013.3250
26	14, 1057-1061	Liu L, Sun T, Liu Z, Chen X, Zhao L, Qu G, Li Q (2014b): Traumatic
27		
28	https://doi.org/10.1080/02699050050203559	brain injury dysregulates microRNAs to modulate cell signaling
29	Jorge RE, Robinson RG, Arndt SV, Starkstein SE, Forrester AW,	in rat hippocampus. PLoS One 9 , e103948
	Geisler F (1993): Depression following traumatic brain injury:	https://doi.org/10.1371/journal.pone.0103948
30	a 1 year longitudinal study. J. Affect Disord. 27, 233-243	Liu W, Liu X, Yang H, Zhu X, Yi H, Zhu X, Zhang J (2013):
31	https://doi.org/10.1016/0165-0327(93)90047-N	Phosphorylated retinoblastoma protein (p-Rb) is involved in
32	Karelina K, Weil ZM (2016): Neuroenergetics of traumatic brain	neuronal apoptosis after traumatic brain injury in adult rats.
33	injury. Concussion 1, CNC9	J. Mol. Histol. 44, 147-158
34	https://doi.org/10.2217/cnc.15.9	https://doi.org/10.1007/s10735-013-9481-z
	Kawai T, Akira S (2006): TLR signaling. Cell Death Differ. 13,	Long J, Cai L, Li J, Zhang L, Yang H, Wang T (2013): JNK3
35	816-825	involvement in nerve cell apoptosis and neurofunctional
36	https://doi.org/10.1038/sj.cdd.4401850	recovery after traumatic brain injury. Neural. Regen. Res.
37		8 , 1491-1499
38	Keller A, Rounge T, Backes C, Ludwig N, Gislefoss R, Leidinger P,	
39	Langseth H, Meese E (2017): Sources to variability in circulating	http://dx.doi.org/10.3969/j.issn.1673-5374.2013.16.006
	human miRNA signatures. RNA Biol. 14, 1791-1798	Lu J, Guan F, Cui F, Sun X, Zhao L, Wang Y, Wang X (2019):
40	https://doi.org/10.1080/15476286.2017.1367888	Enhanced angiogenesis by the hyaluronic acid hydrogels im-
41	Kelly DF, Gonzalo IT, Cohan P, Berman N, Swerdloff R, Wang C	mobilized with a VEGF mimetic peptide in a traumatic brain
42	(2000): Hypopituitarism following traumatic brain injury and	injury model in rats. Regen. Biomater. 6, 325-334
43	aneurysmal subarachnoid hemorrhage: a preliminary report.	https://doi.org/10.1093/rb/rbz027
44	J. Neurosurg. 93, 743-752	Mangiola A, Vigo V, Anile C, De Bonis P, Marziali G, Lofrese G
45	https://doi.org/10.3171/jns.2000.93.5.0743	(2015): Role and Importance of IGF-1 in Traumatic Brain
	Kotlyar M, Pastrello C, Malik Z, Jurisica I (2019): IID 2018 up-	Injuries. Biomed. Res. Int. 2015, 736104
46	date: context-specific physical protein-protein interactions in	https://doi.org/10.1155/2015/736104
47	human, model organisms and domesticated species. Nucleic	Manley G, Gardner AJ, Schneider KJ, Guskiewicz KM, Bailes J,
48		
49	Acids Res. 47, D581-D589	Cantu RC, Castellani RJ, Turner M, Jordan BD, Randolph C et
	https://doi.org/10.1093/nar/gky1037	al. (2017): A systematic review of potential long-term effects
50	Kowalski EA, Chen J, Hazy A, Fritsch LE, Gudenschwager-Basso	of sport-related concussion. Br. J. Sports Med. 51, 969-977
51	EK, Chen M, Wang X, Qian Y, Zhou M, Byerly M et al. (2019):	https://doi.org/10.1136/bjsports-2017-097791
52	Peripheral loss of EphA4 ameliorates TBI-induced neuroin-	Mannix RC, Zhang J, Park J, Lee C, Whalen MJ (2011): Detrimental
53	flammation and tissue damage. J. Neuroinflammation 16, 210	effect of genetic inhibition of B-site APP-cleaving enzyme 1 on

14

55

56

57 58

59

60

61

62

63

64

65

66

67

68

69

70 71

72

73

74 75

76

77

78

80

81

82

83

84

85

86

87

88

89

90

91

92

93 94

95

96

97 98

99

100

101

102

103

104

105

106

 functional outcome after controlled cortical impact in young adult mice. J. Neurotrauma 28, 1855-1861 https://doi.org/10.1089/neu.2011.1759 Mao SS, Hua R, Zhao XP, Qin X, Sun ZQ, Zhang Y, Wu YQ, Jia MX, Cao JL, Zhang YM (2012): Exogenous administration of PACAP alleviates traumatic brain injury in rats through a mechanism involving the TLR4/MyD88/NF-kappaB pathway. J. Neurotrauma 29, 1941-1959 https://doi.org/10.1089/neu.2011.2244 	 https://doi.org/10.1016/j.apmr.2010.05.017 Murayama M, Tanaka S, Palacino J, Murayama O, Honda T, Sun X, Yasutake K, Nihonmatsu N, Wolozin B, Takashima A (1998): Direct association of presenilin-1 with beta-catenin. FEBS Lett. 433, 73-77 https://doi.org/10.1016/S0014-5793(98)00886-2 Nag S, Takahashi JL, Kilty DW (1997): Role of vascular endothelial growth factor in blood-brain barrier breakdown and angiogenesis in brain trauma. J. Neuropathol. Exp. Neurol.
V, Baki L, Wen P, Efthimiopoulos S, Shao Z et al. (2002): A pre- senilin-1/gamma-secretase cleavage releases the E-cadherin intracellular domain and regulates disassembly of adherens junctions. EMBO J. 21 , 1948-1956	 56, 912-921 https://doi.org/10.1097/00005072-199708000-00009 Newell EA, Todd BP, Mahoney J, Pieper AA, Ferguson PJ, Bassuk AG (2018): Combined blockade of interleukin-1alpha and -1beta signaling protects mice from cognitive dysfunction after
Marchetti B, Pluchino S (2013): Wnt your brain be inflamed? Yes, it Wnt! Trends Mol. Med. 19 , 144-156	traumatic brain injury. eNeuro 5 , 1-15 https://doi.org/10.1523/ENEURO.0385-17.2018 Oliva AA, Jr., Kang Y, Sanchez-Molano J, Furones C, Atkins CM
Martinowich K, Lu B (2008): Interaction between BDNF and serotonin: role in mood disorders. Neuropsychopharmacol-	(2012): STAT3 signaling after traumatic brain injury. J. Neurochem. 120 , 710-720 https://doi.org/10.1111/j.1471-4159.2011.07610.x
https://doi.org/10.1038/sj.npp.1301571 McCrea MA, Nelson LD, Guskiewicz K (2017): Diagnosis and	Omalu B, Hammers J (2021): Letter: Recommendation to create new neuropathologic guidelines for the post-mortem diag- nosis of chronic traumatic encephalopathy. Neurosurgery 89 , E97-E98
N Am. 28 , 271-286 https://doi.org/10.1016/j.pmr.2016.12.005	https://doi.org/10.1093/neuros/nyab138 Omalu BI, DeKosky ST, Minster RL, Kamboh MI, Hamilton RL, Wecht CH (2005): Chronic traumatic encephalopathy in
J, Graf-Baumann T, Kelly J, Lovell M, Schamasch P (2005): Summary and agreement statement of the 2nd International	a National Football League player. Neurosurgery 57 , 128-134 https://doi.org/10.1227/01.NEU.0000163407.92769.ED Ou S, Liu GD, Zhou LS, Xia X, Bai SR, Li J, Cui J, Cheng JM, Li YM,
Med. 39 , 196-204 https://doi.org/10.1097/01.jsm.0000159931.77191.29 McKee AC, Cantu RC, Nowinski CJ, Hedley-Whyte ET, Gavett BE,	Zhang XY et al. (2014): Bioinformatics analysis of gene expres- sion profiles in the rat cerebral cortex following traumatic brain injury. Eur. Rev. Med. Pharmacol. Sci. 18 , 101-107
Budson AE, Santini VE, Lee HS, Kubilus CA, Stern RA (2009): Chronic traumatic encephalopathy in athletes: progressive tauopathy after repetitive head injury. J. Neuropathol. Exp. Neurol. 68 , 709-735	Patel RK, Prasad N, Kuwar R, Haldar D, Abdul-Muneer PM (2017): Transforming growth factor-beta 1 signaling regulates neuro- inflammation and apoptosis in mild traumatic brain injury. Brain Behav. Immun. 64 , 244-258
McKee AC, Robinson ME (2014): Military-related traumatic brain injury and neurodegeneration. Alzheimers Dement. 10 , S242-253	https://doi.org/10.1016/j.bbi.2017.04.012 Pinero J, Bravo A, Queralt-Rosinach N, Gutierrez-Sacristan A, Deu- Pons J, Centeno E, Garcia-Garcia J, Sanz F, Furlong LI (2017): DisGeNET: a comprehensive platform integrating information on human disease-associated genes and variants. Nucleic Acids
Meabon JS, Cook DG, Yagi M, Terry GE, Cross DJ, Muzi M, Pagulayan KF, Logsdon AF, Schindler AG, Ghai V et al. (2020):	Res. 45, D833-D839 https://doi.org/10.1093/nar/gkw943
A (VEGF-A) is associated with a history of blast exposure. J. Neurol. Sci. 417 , 117049	Pinchi E, Frati P, Arcangeli M, Volonnino G, Tomassi R, Santoro P, Cipolloni L (2020): MicroRNAs: The new challenge for traumatic brain injury diagnosis. Curr. Neuropharmacol. 18, 319-331
Mellergard P, Sjogren F, Hillman J (2010): Release of VEGF and FGF in the extracellular space following severe subarachnoidal haemorrhage or traumatic head injury in humans. Br. J. Neurosurg. 24 , 261-267	https://doi.org/10.2174/1570159X17666191113100808 Qin X, Li L, Lv Q, Shu Q, Zhang Y, Wang Y (2018): Expression profile of plasma microRNAs and their roles in diagnosis of mild to severe traumatic brain injury. PLoS One 13 , e0204051 https://doi.org/10.1371/journal.pone.0204051
Menon DK, Schwab K, Wright DW, Maas AI, Demographics, Clini- cal Assessment Working Group of the I, Interagency Initiative toward Common Data Elements for Research on Traumatic Brain I, Psychological H (2010): Position statement: definition of traumatic brain injury. Arch. Phys. Med. Rehabil. 91 , 1637-1640	Rahmati S, Abovsky M, Pastrello C, Jurisica I (2017): pathDIP: an annotated resource for known and predicted human gene- pathway associations and pathway enrichment analysis. Nucleic Acids Res. 45 , D419-D426 https://doi.org/10.1093/nar/gkw1082
	 adult mice, J. Neurotrauma 28, 1855-1861 https://doi.org/10.1089/neu.2011.1759 Mao SS, Hua R, Zhao XP, Qin X, Sun ZQ, Zhang Y, Wu YQ, Jia MK, Cao JL, Zhang YM (2012): Exogenous administration of PACAP alleviates traumatic brain injury in rats through a mechanism involving the TLR4/MyD88/NF-kappaB pathway. J. Neurotrauma 29, 1941-1959 https://doi.org/10.1089/neu.2011.2244 Marambaud P, Shioi J, Serban G, Georgakopoulos A, Sarner S, Nagy V, Baki L, Wen P, Efthimiopoulos S, Shao Z et al. (2002): A presenilin-1/gamma-secretase cleavage releases the E-cadherin intracellular domain and regulates disassembly of adherens junctions. EMBO J. 21, 1948-1956 https://doi.org/10.1093/emboj/21.8.1948 Marchetti B, Pluchino S (2013): Wnt your brain be inflamed? Yes, it Wnt! Trends Mol. Med. 19, 144-156 https://doi.org/10.1016/j.molmed.2012.12.001 Martinowich K, Lu B (2008): Interaction between BDNF and serotonin: role in mood disorders. Neuropsychopharmacology 33, 73-83 https://doi.org/10.1088/sj.npp.1301571 McCrea MA, Nelson LD, Guskiewicz K (2017): Diagnosis and management of acute concussion. Phys. Med. Rehabil. Clin. N Am. 28, 271-286 https://doi.org/10.1016/j.pmr.2016.12.005 McCrory P, Johnston K, Meeuwisse W, Aubry M, Cantu R, Dvorak J, Graf-Baumann T, Kelly J, Lovell M, Schamasch P (2005): Summary and agreement statement of the 2nd International Conference on Concussion in Sport, Prague 2004. Br. J. Sports Med. 39, 196-204 https://doi.org/10.1097/NEN.0b013e3181a9d503 McKee AC, Cantu RC, Nowinski CJ, Hedley-Whyte ET, Gavett BE, Budson AE, Santini VE, Lee HS, Kubilus CA, Stern RA (2009): Chronic traumatic encephalopathy in athletes: progressive tauopathy after repetitive head injury. J. Neuropathol. Exp. Neurol. 68, 709-735 https://doi.org/10.1097/NEN.0b013e3181a9d503 McKee AC, Robinson ME (2014): Military-related traumatic brain injury and neurodegeneration. Alzheimers Dement. 10, S242-253 https://doi.org/10.

1	Rahmati S, Abovsky M, Pastrello C, Kotlyar M, Lu R, Cumbaa CA,	Smith DH, Johnson VE, Stewart W (2013): Chronic neuropatholo-
2	Rahman P, Chandran V, Jurisica I (2020): pathDIP 4: an ex-	gies of single and repetitive TBI: substrates of dementia? Nat.
3	tended pathway annotations and enrichment analysis resource	Rev. Neurol. 9, 211-221
4	for human, model organisms and domesticated species. Nucleic	https://doi.org/10.1038/nrneurol.2013.29
5	Acids Res. 48, D479-D488	Stoica BA, Loane DJ, Zhao Z, Kabadi SV, Hanscom M, Byrnes KR,
6	https://doi.org/10.1093/nar/gkz989	Faden AI (2014): PARP-1 inhibition attenuates neuronal loss,
7	Redell JB, Moore AN, Ward NH, 3rd, Hergenroeder GW, Dash PK	microglia activation and neurological deficits after traumatic
	(2010): Human traumatic brain injury alters plasma microRNA	brain injury. J. Neurotrauma 31, 758-772
8	levels. J. Neurotrauma 27, 2147-2156	https://doi.org/10.1089/neu.2013.3194
9	https://doi.org/10.1089/neu.2010.1481	Sun D, Bullock MR, Altememi N, Zhou Z, Hagood S, Rolfe A,
10	Rubovitch V, Edut S, Sarfstein R, Werner H, Pick CG (2010): The	McGinn MJ, Hamm R, Colello RJ (2010): The effect of epi-
11	intricate involvement of the Insulin-like growth factor receptor	dermal growth factor in the injured brain after trauma in rats.
12	signaling in mild traumatic brain injury in mice. Neurobiol.	J. Neurotrauma 27 , 923-938
13	Dis. 38 , 299-303	https://doi.org/10.1089/neu.2009.1209
14	https://doi.org/10.1016/j.nbd.2010.01.021	Sun L, Liu A, Zhang J, Ji W, Li Y, Yang X, Wu Z, Guo J (2018): miR-
15	Rubovitch V, Shachar A, Werner H, Pick CG (2011): Does IGF-1	23b improves cognitive impairments in traumatic brain injury
16	administration after a mild traumatic brain injury in mice acti-	by targeting ATG12-mediated neuronal autophagy. Behav.
17	vate the adaptive arm of ER stress? Neurochem. Int. 58, 443-446	Brain Res. 340, 126-136
	https://doi.org/10.1016/j.neuint.2011.01.009	https://doi.org/10.1016/j.bbr.2016.09.020
18	Salinas PC (2012): Wnt signaling in the vertebrate central nervous	Sun Y, Bai L, Niu X, Wang Z, Yin B, Bai G, Zhang D, Gan S, Sun C,
19	system: from axon guidance to synaptic function. Cold Spring	Wang S et al. (2019): Elevated serum levels of inflammation-
20	Harb. Perspect. Biol. 4, a008003	related cytokines in mild traumatic brain injury are associated
21	https://doi.org/10.1101/cshperspect.a008003	with cognitive performance. Front. Neurol. 10, 1120
22	Sandhir R, Berman NE (2010): Age-dependent response of	https://doi.org/10.3389/fneur.2019.01120
23	CCAAT/enhancer binding proteins following traumatic brain	Sun Y, Jin K, Xie L, Childs J, Mao XO, Logvinova A, Greenberg
24	injury in mice. Neurochem. Int. 56 , 188-193	DA (2003): VEGF-induced neuroprotection, neurogenesis,
25	https://doi.org/10.1016/j.neuint.2009.10.002	and angiogenesis after focal cerebral ischemia. J. Clin. Invest.
26	Sarkar C, Jones JW, Hegdekar N, Thayer JA, Kumar A, Faden AI,	111, 1843-1851
27	Kane MA, Lipinski MM (2020): PLA2G4A/cPLA2-mediated	https://doi.org/10.1172/JCI200317977
28	lysosomal membrane damage leads to inhibition of autophagy	Tamas A, Zsombok A, Farkas O, Reglodi D, Pal J, Buki A, Lengvari
29	and neurodegeneration after brain trauma. Autophagy 16,	I, Povlishock JT, Doczi T (2006): Postinjury administration of
30	466-485	pituitary adenylate cyclase activating polypeptide (PACAP)
	https://doi.org/10.1080/15548627.2019.1628538	attenuates traumatically induced axonal injury in rats. J. Neu-
31	Scranton RA, Baskin DS (2015): Impaired pituitary axes following	rotrauma 23 , 686-695
32	traumatic brain injury. J. Clin. Med. 4 , 1463-1479	https://doi.org/10.1089/neu.2006.23.686
33	https://doi.org/10.3390/jcm4071463	Tang C, Shan Y, Hu Y, Fang Z, Tong Y, Chen M, Wei X, Fu X, Xu
34	Sheinerman KS, Toledo JB, Tsivinsky VG, Irwin D, Grossman M,	X (2017): FGF2 attenuates neural cell death via suppressing
35	Weintraub D, Hurtig HI, Chen-Plotkin A, Wolk DA, McCluskey	autophagy after rat mild traumatic brain injury. Stem Cells Int. 2017 , 2923182
36	LF et al. (2017): Circulating brain-enriched microRNAs as novel biomarkers for detection and differentiation of neurodegenera-	https://doi.org/10.1155/2017/2923182
37	tive diseases. Alzheimers Res. Ther. 9, 89	Tas D, Kaplan O, Sogut O (2020): Validity of serum miRNA 93
38	https://doi.org/10.1186/s13195-017-0316-0	and miRNA 191 to reduce unnecessary computed tomog-
39	Shetty VS, Reis MN, Aulino JM, Berger KL, Broder J, Choudhri AF,	raphy in patients with mild head trauma. J. Clin. Med. Res.
40	Kendi AT, Kessler MM, Kirsch CF, Luttrull MD et al. (2016):	12, 579-589
41	ACR appropriateness criteria head trauma. J. Am. Coll. Radiol.	https://doi.org/10.14740/jocmr4265
42	13, 668-679	Tate CC, Garcia AJ, LaPlaca MC (2007): Plasma fibronectin is
43	https://doi.org/10.1016/j.jacr.2016.02.023	neuroprotective following traumatic brain injury. Exp. Neurol.
	Siedlecki-Wullich D, Catala-Solsona J, Fabregas C, Hernandez I,	207 , 13-22
44	Clarimon J, Lleo A, Boada M, Saura CA, Rodriguez-Alvarez J,	https://doi.org/10.1016/j.expneurol.2007.05.008
45	Minano-Molina AJ (2019): Altered microRNAs related to syn-	Teasdale G, Jennett B (1976): Assessment and prognosis of coma
46	aptic function as potential plasma biomarkers for Alzheimer's	after head injury. Acta Neurochir. (Wien). 34, 45-55
47	disease. Alzheimers Res. Ther. 11, 46	https://doi.org/10.1007/BF01405862
48	https://doi.org/10.1186/s13195-019-0501-4	Terzioglu B, Ekinci O, Berkman Z (2015): Hyperglycemia is a pre-
49	Singh K, Trivedi R, Devi MM, Tripathi RP, Khushu S (2016): Lon-	dictor of prognosis in traumatic brain injury: Tertiary intensive
50	gitudinal changes in the DTI measures, anti-GFAP expression	care unit study. J. Res. Med. Sci. 20 , 1166-1171
51	and levels of serum inflammatory cytokines following mild	https://doi.org/10.4103/1735-1995.172984
52	traumatic brain injury. Exp. Neurol. 275, 427-435	Thau-Zuchman O, Shohami E, Alexandrovich AG, Leker RR (2010):

https://doi.org/10.1016/j.expneurol.2015.07.016

den AI (2014): PARP-1 inhibition attenuates neuronal loss, icroglia activation and neurological deficits after traumatic ain injury. J. Neurotrauma 31, 758-772 tps://doi.org/10.1089/neu.2013.3194 , Bullock MR, Altememi N, Zhou Z, Hagood S, Rolfe A, cGinn MJ, Hamm R, Colello RJ (2010): The effect of epi-rmal growth factor in the injured brain after trauma in rats. Neurotrauma 27, 923-938 ps://doi.org/10.1089/neu.2009.1209 Liu A, Zhang J, Ji W, Li Y, Yang X, Wu Z, Guo J (2018): miR-b improves cognitive impairments in traumatic brain injury targeting ATG12-mediated neuronal autophagy. Behav. ain Res. 340, 126-136 ps://doi.org/10.1016/j.bbr.2016.09.020 Bai L, Niu X, Wang Z, Yin B, Bai G, Zhang D, Gan S, Sun C, ang S et al. (2019): Elevated serum levels of inflammation-ated cytokines in mild traumatic brain injury are associated th cognitive performance. Front. Neurol. 10, 1120 tps://doi.org/10.3389/fneur.2019.01120 Jin K, Xie L, Childs J, Mao XO, Logvinova A, Greenberg A (2003): VEGF-induced neuroprotection, neurogenesis, d angiogenesis after focal cerebral ischemia. J. Clin. Invest. 1, 1843-1851 tps://doi.org/10.1172/JCI200317977 A, Zsombok A, Farkas O, Reglodi D, Pal J, Buki A, Lengvari Povlishock JT, Doczi T (2006): Postinjury administration of uitary adenylate cyclase activating polypeptide (PACAP) enuates traumatically induced axonal injury in rats. J. Neu-rauma 23, 686-695 tps://doi.org/10.1089/neu.2006.23.686 C, Shan Y, Hu Y, Fang Z, Tong Y, Chen M, Wei X, Fu X, Xu (2017): FGF2 attenuates neural cell death via suppressing tophagy after rat mild traumatic brain injury. Stem Cells . 2017, 2923182 tps://doi.org/10.1155/2017/2923182 Kaplan O, Sogut O (2020): Validity of serum miRNA 93 d miRNA 191 to reduce unnecessary computed tomog-phy in patients with mild head trauma. J. Clin. Med. Res. , 579-589 tps://doi.org/10.14740/jocmr4265 C, Garcia AJ, LaPlaca MC (2007): Plasma fibronectin is uroprotective following traumatic brain injury. Exp. Neurol. 7,13-22 ps://doi.org/10.1016/j.expneurol.2007.05.008 lle G, Jennett B (1976): Assessment and prognosis of coma er head injury. Acta Neurochir. (Wien). 34, 45-55 ps://doi.org/10.1007/BF01405862 glu B, Ekinci O, Berkman Z (2015): Hyperglycemia is a pre-ctor of prognosis in traumatic brain injury: Tertiary intensive e unit study. J. Res. Med. Sci. 20, 1166-1171 tps://doi.org/10.4103/1735-1995.172984 Zuchman O, Shohami E, Alexandrovich AG, Leker RR (2010): Vascular endothelial growth factor increases neurogenesis

 after traumatic brain injury. J. Cereb. Blood Flow Metab. 30, 1008-1016 https://doi.org/10.1038/jcbfm.2009.271 Tokar T, Pastrello C, Rossos AEM, Abovsky M, Hauschild AC, Tsay M, Lu R, Jurisica I (2018): mirDIP 4.1-integrative database of human microRNA target predictions. Nucleic Acids Res. 46, D360-D370 https://doi.org/10.1093/nar/gkx1144 Truettner JS, Hu B, Alonso OF, Bramlett HM, Kokame K, Dietrich WD (2007): Subcellular stress response after traumatic brain injury. J. Neurotrauma 24, 599-612 https://doi.org/10.1089/neu.2006.0186 Tylicka M, Matuszczak E, Hermanowicz A, Debek W, Karpinska M, Kaminska J, Koper-Lenkiewicz OM (2020): BDNF and IL-8, but not UCHL-1 and IL-11, are markers of brain injury in children caused by mild head trauma. Brain Sci. 10, 665 Uwatoko H, Hama Y, Iwata IT, Shirai S, Matsushima M, Yabe I. Utsumi J, Sasaki H (2019): Identification of plasma microRNA expression changes in multiple system atrophy and Parkinson's disease. Mol. Brain 12, 49 https://doi.org/10.1186/s13041-019-0471-2 Ved R, Sharouf F, Harari B, Muzaffar M, Manivannan S, Ormonde C, Gray WP, Zaben M (2021): Disulfide HMGB1 acts via TLR2/4 receptors to reduce the numbers of oligodendrocyte progenitor cells after traumatic injury in vitro. Sci. Rep. 11, 6181 https://doi.org/10.1038/s41598-021-84932-0 Wang Y, Hameed MQ, Rakhade SN, Iglesias AH, Muller PA, Mou DL, Rotenberg A (2014): Hippocampal immediate early gene transcription in the rat fluid percussion traumatic brain injury model. Neuroreport 25, 954-959 https://doi.org/10.1038/s41598-020-60133-z Wintermark M, Sanelli PC, Anzai Y, Tsiouris AJ, Whitlow CT, Institute ACRNI Jost M, Sequencing of rat hippocampus and human biofluids identifies acute, chronic, focal and diffuse traumatic brain injury: sci. Rep. 8, 4935 https://doi.org/10.1038/s41598-018-23003-3 Yan J, Bu X, Li Z, Wu J, Wang C, Li D, Song J, Wang J (2019): Screening the expressio	 Neurochem. 150, 202-217 https://doi.org/10.1111/jnc.14717 Yang J, You Z, Kim HH, Hwang SK, Khuman J, Guo S, Lo EH, Whalen MJ (2010): Genetic analysis of the role of tumor necrosis factor receptors in functional outcome after traumatic brain injury in mice. J. Neurotrauma 27, 1037-1046 https://doi.org/10.1089/neu.2009.1229 Yang L, Wu H, Yang F, Li P, Huang Y, Zhang X, Qian C (2021): Identification of candidate genes and pathways in dexmedetomidine-induced neuroprotection in rats using RNA sequencing and bioinformatics analysis. Ann. Palliat. Med. 10, 372-384 https://doi.org/10.21037/apm-20-2346 Yang T, Song J, Bu X, Wang C, Wu J, Cai J, Wan S, Fan C, Zhang C, Wang J (2016): Elevated serum miR-93, miR-191, and miR-499 are noninvasive biomarkers for the presence and progression of traumatic brain injury. J. Neurochem. 137, 122-129 https://doi.org/10.1111/jnc.13534 Yao XL, Liu J, Lee E, Ling GS, McCabe JT (2006): Cullin 5 gene expression in the rat cerebral cortex and hippocampus following traumatic brain injury (TBI). Neurosci. Lett. 409, 65-69 https://doi.org/10.1016/j.neulet.2006.09.015 Yoshino A, Hovda DA, Kawamata T, Katayama Y, Becker DP (1991): Dynamic changes in local cerebral glucose utilization following cerebral conclusion in rats: evidence of a hyper- and subsequent hypometabolic state. Brain Res. 561, 106-119 https://doi.org/10.1016/0006-8993(91)90755-K Zhang JY, Lee JH, Gu X, Wei ZZ, Harris MJ, Yu SP, Wei L (2018): Intranasally delivered Wnt3a improves functional recovery after traumatic brain injury by modulating autophagic, apoptotic, and regenerative pathways in the mouse brain. J. Neurotrauma 35, 802-813 https://doi.org/10.1089/neu.2016.4871 Zhao J, Xu C, Cao H, Zhang L, Wang X, Chen S (2019): Identification of target genes in neuroinflammation and neurodegeneration of target genes in neuroinflammation and neurodegeneration after traumatic brain injury in rats. PeerJ. 7, e8324 https://doi.org/10

 wang SK, Khuman J, Guo S, Lo EH, Wha-ic analysis of the role of tumor necrosis nctional outcome after traumatic brain rotrauma **27**, 1037-1046 P, Huang Y, Zhang X, Qian C (2021): idate genes and pathways in dexmedetoprotection in rats using RNA sequencing alysis. Ann. Palliat. Med. 10, 372-384 g C, Wu J, Cai J, Wan S, Fan C, Zhang C, ed serum miR-93, miR-191, and miR-499 arkers for the presence and progression of y. J. Neurochem. 137, 122-129 g GS, McCabe JT (2006): Cullin 5 gene erebral cortex and hippocampus follow-jury (TBI). Neurosci. Lett. 409, 65-69 6/j.neulet.2006.09.015 wamata T, Katayama Y, Becker DP (1991): ocal cerebral glucose utilization following rats: evidence of a hyper- and subsequent Brain Res. 561, 106-119 6/0006-8993(91)90755-K Wei ZZ, Harris MJ, Yu SP, Wei L (2018): Wnt3a improves functional recovery after y by modulating autophagic, apoptotic, ways in the mouse brain. J. Neurotrauma ng L, Wang X, Chen S (2019): Identifica-neuroinflammation and neurodegenera-rain injury in rats. PeerJ. 7, e8324 Moore AN, Hylin MJ, Menge T, Xue H, er D, Johnson EM et al. (2016): Wnt3a, a esenchymal stem cells is neuroprotective cognitive recovery following traumatic ahn G (2019): Type 2 diabetes predicts odegenerative complications in veterans ain injury. J. Endocrinol. Diabetes 6, 137 26/2374-6890/6/3/001137