

Short Communication

Identification of All α_1 -Adrenoceptor Subtypes in Rat Lung

M. NOVÁKOVÁ AND J. MYSLIVEČEK

*Institute of Physiology, 1st Faculty of Medicine,
Charles University, Prague, Czech Republic. E-mail: noma@1fl.cuni.cz*

Abstract. The function of lung tissue is regulated *via* a release of neurotransmitters from autonomic nerves. The neurotransmitters of sympathetic nervous system, adrenaline and noradrenaline, activate both α -adrenoceptors and β -adrenoceptors. Although the function and expression of β -adrenoceptors can be considered major, some doubts exist about the function and expression of α_1 -adrenoceptor subtypes in the lung tissue. Therefore a set of competition binding experiments was employed in order to discriminate between the α_1 -adrenoceptor binding site subtypes in the rat lung. We identified three subpopulations of α_1 -adrenoceptor binding sites in the rat lung (α_{1A} , α_{1B} and α_{1D}).

Key words: α_1 -adrenoceptor subtypes — Lung — Rat

α -adrenoceptors belong to the most numerous family of membrane receptors – the G-protein coupled receptor family. They can be divided into three subtypes: α_{1A} , α_{1B} and α_{1D} (Zhong and Minneman 1999). All these subtypes activate $G_{q/11}$ family of G-proteins and consequently phospholipase C β (PLC) that cleaves phosphatidylinositol to inositoltrisphosphate and diacylglycerol. Both molecules are able to act as second messengers. Tissue distribution of these receptor subtypes is now generally considered to be a mixture of all three subtypes (Zhong and Minneman 1999). Although the role of α_1 -adrenoceptors in the lung function is assumed to be minor, some findings have revealed the role of low-affinity prazosine binding sites (previously assumed as α_{1L} receptors) in allergic bronchoconstriction (Nobata et al. 2002). This finding is also important as the ligand used (JTH-601) in this study has high affinity for α_{1A} - and α_{1B} -adrenoceptors but low affinity for α_{1D} -adrenoceptors (Takahashi et al. 2000). Surprisingly, some papers have described only two subtypes in the lung tissue (Yang et al. 1998). On the other hand, in the middle of 90's, Hiramatsu et al. (1994) have suggested that the α_1 -adrenoceptors of rat lung are composed of three distinct subtypes. Similarly, all three mRNA subtypes have been detected in the lung tissue (Faure et al. 1994; Alonso-Llamazares

Correspondence to: Jaromír Mysliveček, Institute of Physiology, 1st Faculty of Medicine, Charles University, Albertov 5, 128 00 Prague, Czech Republic.
E-mail: jmys@1fl.cuni.cz

et al. 1995). Nevertheless, the relationship between mRNA and protein expression is not always obvious (Zhong and Minneman 1999). Therefore we have employed the set of competition binding experiments with selective antagonists for α_{1A} (RS 17053), α_{1B} (L-765,314) and α_{1D} (BMY 7378) aiming to determine the binding site subtype proportion in the lung tissue.

Experiments were performed on adult Wistar male rats (200–250 g weight, age 46–55 days). They were housed in standard conditions (12/12 light/dark cycle, feeding and drinking water *ad libitum*). Animals were sacrificed by cervical dislocation and decapitation. Lung tissue was isolated, adjacent tissue was carefully discarded, the lung tissue was weighed, cut to small pieces and homogenized with an UltraTurrax homogenizer (Janke and Kunkel, Staufen, Germany) in an ice-cold saline. The homogenates were stored frozen at -20°C until the measurements of radioligand binding. Radioligand binding experiments were performed as described previously (Mysliveček et al. 2003) with minor modifications. The homogenates were re-homogenized just before the addition of tissue to the mixture of radioligand and antagonist. Preliminary saturation binding experiments with ^3H -prazosin (specific activity 2.77 TBq/mmol; Amersham Biosciences, Little Chalfont, Buckinghamshire, UK) revealed the following binding parameters: $B_{\text{max}} = 253$ fmol/mg of protein (255.7 and 250.3) and $K_{\text{D}} = 585$ pmol/l (657.1 and 514.7). The K_{D} value is in good agreement with our previously published data on rat heart α_1 -adrenoceptors (Mysliveček et al. 2003) and B_{max} is similar to the data of other authors on lung ^3H -prazosin binding (Latifpour and Bylund 1983). The competition binding experiments with ^3H -prazosin (560 pmol/l) and specific antagonists were performed in order to ascertain the proportion of α_1 -adrenoceptor subtypes in the lung tissue. The following compounds were used as specific antagonists for α_{1A} , α_{1B} and α_{1D} : RS 17053 (Tocris-Cocoon, Bristol, UK), L-765,314 and BMY 7378 (Sigma-Aldrich, Prague, Czech Republic). The antagonists' ranges were between 10^{-11} and 10^{-5} mol/l (RS 17053), between 10^{-10} and 10^{-4} mol/l (L-765,314), and between 10^{-10} and 10^{-4} mol/l (BMY 7378) using two concentrations *per* grade. The incubations were performed in triplicates in 25°C for 90 min and were terminated by fast washing with ice-cold distilled water – the same washing method as published previously (Mysliveček et al. 2003) – through Whatman GF/B filter that were pre-soaked in distilled water in order to diminish the non-specific binding with the use of a Brandel cell harvester. Generally, the washing of filters using Brandel cell harvester is extremely fast and therefore the differences in the composition of washing fluid are not of great effect on the binding. Moreover, we have compared the binding on filters washed with buffer and ice-cold distilled water and no differences have been found, therefore we have employed this washing method. The filters were dried overnight and the radioactivity retained on them was measured by liquid scintillation spectrometry using Bray's solution. Proteins were determined using modified Lowry's method. Data were analysed using GraphPad Prism 3.0 program (GraphPad Software, San Diego, CA).

All three α_1 -adrenoceptor binding site subtypes were detected in the rat tissue, i.e. all three antagonists competed with ^3H -prazosin in a biphasic manner (see

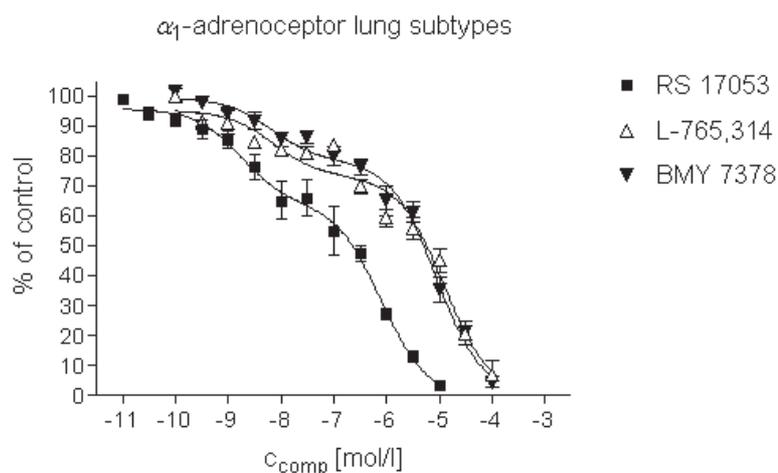


Figure 1. The competition binding of ^3H -prazosin and different antagonists in the lung. Data from three independent experiments are presented as means \pm S.E.M. Abscissa: common logarithm of competitor concentration (c_{comp}) in mol/l. Ordinate: % of total bound. Explanation of symbols is inserted.

Table 1. The affinities and fractions revealed from competition binding experiments. Data from three independent experiments performed in triplicates are expressed as means \pm S.E.M.

Competitor	Fraction 1 (%)	pKB1	pKB2
RS 17053	32.72 \pm 3.49	9.76 \pm 0.29	6.85 \pm 0.16
L-765,314	23.06 \pm 2.83	9.28 \pm 0.29	5.79 \pm 0.25
BMY 7378	21.07 \pm 2.47	9.37 \pm 0.25	5.90 \pm 0.39

Figure 1). The respective values of high affinity fraction, pKB1 and pKB2, are given in Table 1. We can therefore conclude that the lung tissue contains all three binding site subtypes of α_1 -adrenoceptors. As it can be seen from the Table 1, there is another proportion of ^3H -prazosin binding that is undistinguishable by α_1 antagonists.

It has been mentioned previously that the major attention was paid to the function of β -adrenoceptors and the consequences of α_1 -adrenoceptor function are still not clear. The results concerning the role of α_1 -adrenoceptors in asthma are difficult to interpret (for review, see Goldie et al. 1990). Although the role of α_1 -adrenoceptors in the function of respiratory tract is not fully understood yet, some findings suggest the possibility that their role lies in the potentiation of β -adrenoceptor function (Nousiainen et al. 1977; Goldie et al. 1990). The role of α_1 -adrenoceptor subtype has not been studied yet. There are some findings that

lung tissue also contains α_2 -adrenoceptors (for review, see Calzada and de Artinano 2001). Our results show that all three α_1 -adrenoceptor binding site subtypes are present in the lung tissue. All three competitors revealed high-affinity binding sites whose binding affinities (pKB₁, see Table 1) were comparable to that for cloned α_{1A} -, α_{1B} - and α_{1D} -adrenoceptor subtypes. Thus, pKB1 revealed by RS 17053 (α_{1A} -specific antagonist) was almost same (9.76 *vs.* 9.1–9.9) as that indicated by Ford et al. (1996) for cloned α_{1A} -adrenoceptors, pKB1 for L-765,314 (α_{1B} -specific antagonist) was similar (9.28 *vs.* 8.7) to that indicated by Patane et al. (1998) for cloned α_{1B} -adrenoceptors, and pKB1 for BMY 7378 (specific antagonist of α_{1D} -adrenoceptors) was virtually same (9.37 *vs.* 9.0–9.4) as that reviewed by Willems et al. (2003) for cloned α_{1D} -adrenoceptor. On the other hand, the low affinity binding sites (pKB2) have shown the values that implies to the presence of more binding sites with similar affinities. By other words, it shows the presence of one high-affinity binding site and the other (with low affinity) that is mixture of two other α_1 -adrenoceptor binding site subtypes. Moreover, there is another proportion of ³H-prazosin binding that is undistinguishable by α_1 antagonists (see Table 1). As ³H-prazosin also binds to the α_2 -adrenoceptors, it is possible to assign this binding to α_2 -adrenoceptors. These receptors are also present in the lung tissue as have been demonstrated multiple times (for review, see Goldie et al. 1990).

Our results, describing three binding sites, are in good agreement with data from the studies that detected mRNAs for these receptor subtypes (Faure et al. 1994; Alonso-Llamazares et al. 1995). Also, Hiramatsu et al. (1994) have identified three subtypes in rat lung using radioligand binding studies. However, the authors could only concluded, in context with erstwhile knowledge, that in the rat lung α_{1A} , α_{1B} and unknown subtypes are present. On the other hand, Yang et al. (1998) have identified only α_{1A} and α_{1B} binding subtypes in the murine tissue. Similarly, Calzada and de Artinano (2001) have reviewed also the expression of two mRNAs (α_{1A} and α_{1B} only). It is not fully clear why our data vary from those obtained from papers by Yang et al. (1998) and Calzada and de Artinano (2001). Contrary to that, there is another reference (Hiramatsu et al. 1994) showing the binding to all three α_1 -adrenoceptor subtypes. Our finding of three subtypes of α_1 -adrenoceptor binding sites in lung seems to support opinion on the existence of all three (α_{1A} -, α_{1B} - and α_{1D} -adrenoceptor subtypes) in this tissue.

Acknowledgements. This work was supported by grant NATO LST.EAP.CLG.980745. The excellent technical assistance by Miss Eva Šimáková is greatly appreciated.

References

- Alonso-Llamazares A., Zamanillo D., Tasanova E., Ovalle S., Calvo P., Chinchetru M. A. (1995): Molecular cloning of α_1 -adrenergic receptor and tissue distribution of three α_1 -adrenergic receptor subtypes in mouse. *J. Neurochem.* **65**, 2387–2392
- Calzada B. C., de Artinano A. A. (2001): Alpha-adrenoceptor subtypes. *Pharmacol. Res.* **44**, 195–208

- Faure C., Pimoule C., Arbilla S., Langer S. Z., Graham D. (1994): Expression of α_1 -adrenoceptor subtypes in rat tissues: implications for α_1 -adrenoceptor classification. *Eur. J. Pharmacol.* **268**, 141—149
- Ford A. P., Arredondo N. F., Blue D. R. Jr., Bonhaus D. W., Jasper J., Kava M. S., Lesnick J., Pfister J. R., Shieh I. A., Vimont R. L., Williams T. J., McNeal J. E., Stamey T. A., Clarke D. E. (1996): RS-17053 (N-[2-(2-cyclopropylmethoxyphenoxy)ethyl]-5-chloro- α , α -dimethyl-1H-indole-3-ethanamine hydrochloride), a selective α_1 -adrenoceptor antagonist, displays low affinity for functional α_1 -adrenoceptors in human prostate: implications for adrenoceptor classification. *Mol. Pharmacol.* **49**, 209—215
- Goldie R. G., Paterson J. W., Lulich K. M. (1990): Adrenoceptors in airway smooth muscle. *Pharmacol. Ther.* **48**, 295—322
- Hiramatsu Y., Muraoka R., Kigoshi S., Muramatsu I. (1994): Identification of α_1 -adrenoceptor subtypes in rat lung by binding of [3 H]-prazosin and [3 H]-WB4101. *J. Recept. Res.* **14**, 75—98
- Latifpour J., Bylund D. B. (1983): Characterization of adrenergic receptor binding in rat lung: physiological regulation. *J. Pharmacol. Exp. Ther.* **224**, 186—192
- Myslivičėk J., Řiční J., Kolář F., Tuček S. (2003): The effects of hydrocortisone on rat heart muscarinic and adrenergic α_1 , β_1 and β_2 receptors, propranolol-resistant binding sites and on some consequent steps in intracellular signalling. *Nuanyň Schmiedebergs Arch. Pharmacol.* **368**, 366—376
- Nobata K., Fujimura M., Ishiura Y., Hirose T., Furusyou S., Myou S., Kurashima K., Kasahara K., Nakao S. (2002): α_{1L} -, but not α_{1H} -, adrenoceptor antagonist prevents allergic bronchoconstriction in guinea pigs *in vivo*. *Eur. J. Pharmacol.* **452**, 97—104
- Nousiainen O. T., Arnala I., Airaksinen M. M., Kokkola K. (1977): Potentiation of β -sympathomimetics with α -adrenoceptor blocking drugs in guinea pig trachea and human bronchus preparations. *Pharmacology* **15**, 469—77
- Patane M. A., Scott A. L., Broten T. P., Chang R. S. L., Ransom R. W., DiSalvo J., Forray C., Bock M. G. (1998): 4-Amino-2-[4-[1-(benzyloxycarbonyl)-2(S)-[[[(1,1-dimethylethyl)amino]carbonyl]-piperazinyl]-6,7-dimethoxyquinazoline (L-765,314): a potent and selective α_{1B} -adrenergic receptor antagonist. *J. Med. Chem.* **41**, 1205—1208
- Takahashi M., Taniguchi T., Kanamaru H., Okada K., Muramatsu I. (2000): Pharmacological characterization of [3 H]-JTH-601, a novel α_1 -adrenoceptor antagonist. Binding to recombinant human α_1 -adrenoceptors and human prostates. *Life Sci.* **67**, 2443—2451
- Willems E. W., Valdivia L. F., Villalón C. M., Saxena P. R. (2003): Possible role of α -adrenoceptor subtypes in acute migraine therapy. *Cephalalgia* **23**, 245—257
- Yang M., Reese J., Cotecchia S., Michel M. C. (1998): Murine α_1 -adrenoceptor subtypes. I. Radioligand binding studies. *J. Pharmacol. Exp. Ther.* **286**, 841—847
- Zhong H., Minneman K. P. (1999): α_1 -adrenoceptor subtypes. *Eur. J. Pharmacol.* **375**, 261—276