

## Effects of Strontium Ions on Contraction and Action Potential in Rabbit Papillary Muscles. A Comparison with Effects of Tetraethylammonium Ions

M ŠTENGL AND P PUČELÍK

*Department of Physiology, Faculty of Medicine, Charles University,  
301 66 Pilsen, Czech Republic*

**Abstract.** The effects of  $\text{Sr}^{2+}$  on contraction and action potential were studied in rabbit papillary muscles and compared with effects of tetraethylammonium ( $\text{TEA}^+$ ). The membrane potential was measured with KCl-filled microelectrodes and the contraction was simultaneously recorded using a mechanoelectrical transducer. A partial (90 %) substitution of extracellular  $\text{Ca}^{2+}$  ( $\text{Ca}_e^{2+}$ ) by  $\text{Sr}^{2+}$  produced stimulation frequency-dependent prolongation of the action potential (AP) with a dominant phase “plateau” as well as prolongation of the contraction. At low frequencies where the AP prolongation was well pronounced, the contraction became biphasic. The effect of  $\text{Sr}^{2+}$  on both AP and contraction was blocked by nifedipine (10  $\mu\text{mol/l}$ ) or by increasing  $\text{Ca}_e^{2+}$ . Ryanodine suppressed the early contraction component only. AP was prolonged to a similar extent and in the same frequency-dependent manner by  $\text{TEA}^+$  (20 mmol/l). Despite similar AP configuration, no biphasic contraction developed in the presence of  $\text{TEA}^+$ . High  $\text{Ca}_e^{2+}$  (10 mmol/l) or low  $\text{Na}_e^+$  (70 mmol/l) suppressed the  $\text{TEA}^+$  effect on AP. The data indicate that the two components of the biphasic contraction are of different origin, the early one is activated by activator cation released from the sarcoplasmic reticulum while the late one results from the  $\text{Sr}^{2+}$  entry across the sarcolemma via L-type  $\text{Ca}^{2+}$  channels.

**Key words:** Heart — Strontium — Biphasic contraction — Tetraethylammonium

### Introduction

$\text{Sr}^{2+}$  can substitute for  $\text{Ca}^{2+}$  in some electrophysiological processes as well as in generating tension at myofibrillar level. It was reported both to permeate calcium channel as a charge carrier of the L-type calcium current ( $I_{\text{CaL}}$ , Kohlhardt et

---

Correspondence to Dr Milan Štengl, Department of Physiology, Faculty of Medicine, Charles University, Lidická 1, 301 66 Pilsen, Czech Republic. E-mail: stengl@1fp.cuni.cz

al 1973, Hess et al 1986) and to activate the contractile apparatus (Kerrick et al 1980) However, there is a difference observed in cardiac muscle  $\text{Sr}^{2+}$  prolongs, usually quite markedly, the durations of AP and of contraction (Kondo 1987) These effects, however, were studied at low stimulation frequencies (Braveny and Šumbera 1972, King and Bose 1983, Kondo 1987) and their frequency-dependence was not described At low stimulation frequencies, partial (90%) substitution of  $\text{Ca}^{2+}$  with  $\text{Sr}^{2+}$  in dog trabeculae (King and Bose 1983) or in different species (Bravený and Šumbera 1972) was reported to induce development of biphasic contractions where the early component was attributed to intracellular release of activator cation and the late one to  $\text{Sr}^{2+}$  entry across sarcolemma The way of  $\text{Sr}^{2+}$  entry, however remained obscure The L-type  $\text{Ca}^{2+}$  channels were suggested but  $\text{Sr}^{2+}$  could enter the cell also via the  $\text{Na}^+/\text{Ca}^{2+}$  exchanger working in the reverse mode

In this study, the frequency-dependence of  $\text{Sr}^{2+}$  effects on both AP and contraction will be described Furthermore, the  $\text{Sr}^{2+}$  effects will be compared with the effects of a classical potassium conductance blocker,  $\text{TEA}^+$  (for review, see Stanfield 1983), another intervention which prolongs the duration of the action potential (APD) Application of  $\text{TEA}^+$  prolongs APD in neurons (Tasaki and Hagiwara 1957), cardiac Purkinje fibers (Ito and Surawicz 1981), and working ventricular myocardium (Ochi and Nishiye 1974) The effect was attributed to a block of outward potassium current (Armstrong 1971) It was also shown, in canine Purkinje fibers that the effect on APD was strongly frequency-dependent showing a reverse use dependence (Ito and Surawicz 1981) The comparison of  $\text{Sr}^{2+}$  and  $\text{TEA}^+$  effects allows (under certain assumptions) to discuss the importance of  $I_{\text{CaL}}$  and/or  $\text{Na}^+/\text{Ca}^{2+}$  exchange for development of the late component In addition, it will be shown that the  $\text{TEA}^+$  effect is antagonized by interventions increasing intracellular  $\text{Ca}^{2+}$  which could play a role in the reverse use dependence of the  $\text{TEA}^+$  effect

## Materials and Methods

Adult rabbits (1.5–3 kg) of either sex were anaesthetized with pentobarbital (60 mg/kg body weight and 1000 IU/kg heparin, i.v.) Hearts were quickly excised and placed in Tyrode solution (see the composition below) Thin papillary muscles (diameter 0.4–0.8 mm, length  $\sim$  4 mm) were dissected from the right ventricle and placed in the experimental chamber, where they were attached to an isometric force transducer (RCA 5734, for more detail, see Pučelik et al 1983) After an initial stabilization period, the resting tension was set so that the developed twitch tension reached 90–95% of maximum at 1 Hz stimulation frequency The bath was perfused with Tyrode solution at 36°C at a constant flow rate (6–10 ml/min)

Muscle preparations were stimulated by punctate electrodes (square-wave voltage pulses 0.1 ms duration, amplitude  $\geq$  50% above threshold) at a chosen stimulation frequency (3.33, 2, 1.43, 1, 0.2, 0.1, or 0.03 Hz) Membrane potential was

measured with glass microelectrodes (filled with 3 mol/l KCl, resistance  $\sim 20 \text{ M}\Omega$ ), simultaneously with the isometric tension viewed on an oscilloscope and recorded using an analogue tape recorder (Racal Thermionic, GB) and/or a pen recorder (RFT NEK polygraph, FRG).

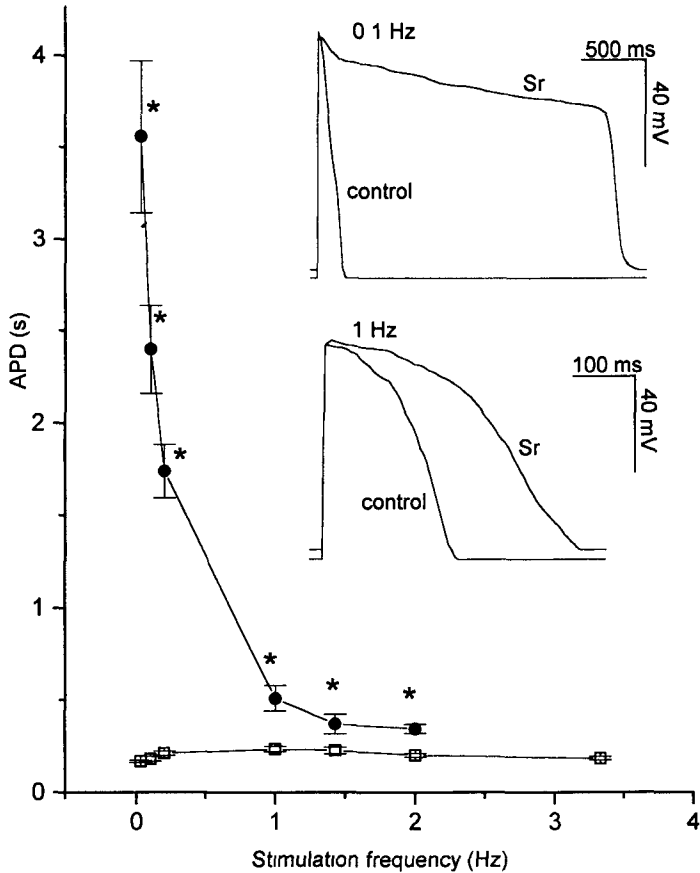
The Tyrode solution had the following composition (in mmol/l): NaCl 137, KCl 4, CaCl<sub>2</sub> 2, MgCl<sub>2</sub> 1, NaH<sub>2</sub>PO<sub>4</sub> 0.5, NaHCO<sub>3</sub> 11, glucose 10. The solution was aerated with 95% O<sub>2</sub> and 5% CO<sub>2</sub> (pH 7.4). Tetraethylammonium bromide (TEA<sup>+</sup>; 20 mmol/l) was added to the normal Tyrode solution to make the TEA<sup>+</sup>-medium. In Sr<sup>2+</sup>-medium, 90% of Ca<sup>2+</sup> was substituted by Sr<sup>2+</sup> (the solution thus contained 1.8 mmol/l Sr<sup>2+</sup> + 0.2 mmol/l Ca<sup>2+</sup>). In some experiments, Sr<sup>2+</sup> and Ca<sup>2+</sup> concentrations were different: 5.8 mmol/l + 0.2 mmol/l or 4 mmol/l + 2 mmol/l, respectively. Solutions containing nifedipine (Sigma, St. Louis, USA) were protected from light. Ryanodine and other chemicals were from Sigma.

Only steady state AP and contractions were used for analysis. Since both Sr<sup>2+</sup> and TEA<sup>+</sup> slowed down the terminal repolarization, and measurement of APD at more positive levels would therefore underestimate their effect, APD was measured at a level corresponding to 90% repolarization. For mechanical data, peak values of active tension are reported and expressed in arbitrary units (*a.u.*). The resting tension level was always taken as zero. When means were calculated the values were standardized to contraction at 1 Hz under control conditions, and are given in relative units (*r.u.*). The contraction duration and the time to peak of contraction were measured from the onset of AP. Data are presented as mean  $\pm$  S.E.M. When appropriate, Student's *t*-test was used for statistical analysis. Differences at  $p \leq 0.05$  were considered to be significant.

## Results

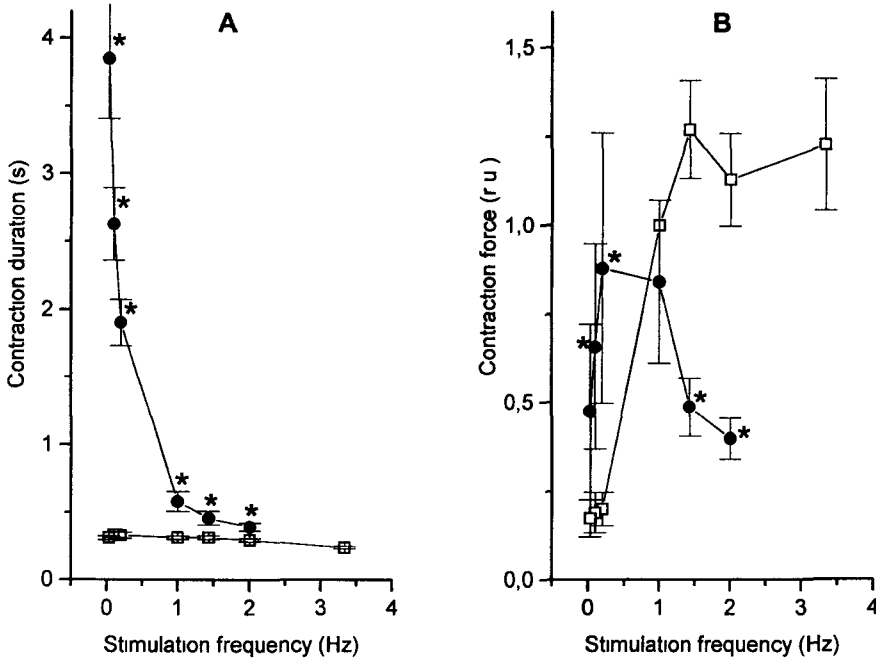
A partial (90%) substitution of extracellular Ca<sup>2+</sup> by Sr<sup>2+</sup> (Sr<sup>2+</sup> 1.8 mmol/l + Ca<sup>2+</sup> 0.2 mmol/l) caused considerable changes of both AP and contraction. AP was prolonged and the effect was stimulation frequency-dependent (Fig. 1). The effect was more pronounced at low stimulation frequencies, e.g. at 1 Hz frequency APD increased from  $235.0 \pm 15.0 \text{ ms}$  ( $n = 6$ ) in control to  $508.8 \pm 69.8 \text{ ms}$  in Sr<sup>2+</sup>-containing medium, and at 0.1 Hz from  $181.7 \pm 12.2 \text{ ms}$  to  $2400.0 \pm 239.3 \text{ ms}$ . The shape of the AP was quite characteristic: dominant phase 2 (plateau) terminated by a relatively fast terminal repolarization (Fig. 1, insets). There was a tendency for mild depolarization of the resting membrane potential (RMP), the changes, however, were not significant (e.g.,  $-80.2 \pm 2.6 \text{ mV}$  ( $n = 6$ ) in control and  $-77.0 \pm 3.0 \text{ mV}$  in Sr<sup>2+</sup>-medium at 0.1 Hz).

In the presence of Sr<sup>2+</sup>, the contraction was prolonged correspondingly with the AP prolongation in a frequency-dependent manner (Fig. 2A): e.g., at 1 Hz, the contraction duration increased from  $311.7 \pm 11.9 \text{ ms}$  ( $n = 6$ ) in control to



**Figure 1.** Effect of  $\text{Sr}^{2+}$  on APD at various stimulation frequencies  $\text{Sr}^{2+}$  replacing 90% of extracellular  $\text{Ca}^{2+}$  ( $\text{Sr}^{2+}$  1.8 mmol/l +  $\text{Ca}^{2+}$  0.2 mmol/l)  $n = 6$ , \*, significantly different from control,  $p \leq 0.05$ . Open squares control. Filled circles  $\text{Sr}^{2+}$ . Data point reflecting APD in  $\text{Sr}^{2+}$ -medium at the highest frequency (3.33 Hz) was omitted since APD was longer than the cycle length (300 ms), hence the real frequency was lower. The effect of  $\text{Sr}^{2+}$  increases with the decreasing frequency. Top inset: Effect of  $\text{Sr}^{2+}$  on AP at the stimulation frequency of 0.1 Hz. Bottom inset: Effect of  $\text{Sr}^{2+}$  on AP at the stimulation frequency of 1 Hz.

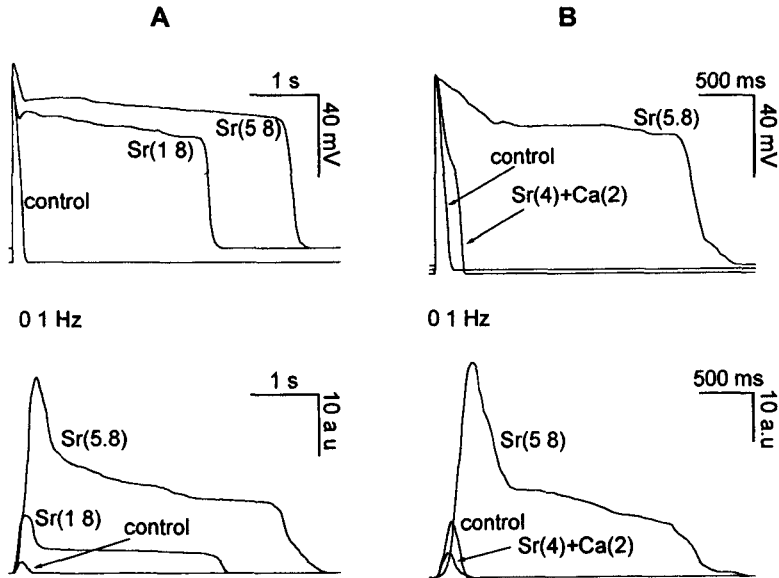
582.5 ± 73.8 ms in  $\text{Sr}^{2+}$ -containing solution, and at 0.1 Hz from 331.7 ± 19.9 ms to 262.8 ± 26.8 ms. Moreover, at low stimulation frequencies (0.2 Hz and less) where the AP prolongation was well pronounced, the contraction became clearly biphasic with the first fast and the second slow components. The relative amplitude of the second component was frequency-dependent, being 31.1 ± 4.8% ( $n = 6$ ) of the



**Figure 2.** Effect of Sr<sup>2+</sup> on contraction. Sr<sup>2+</sup> replacing 90% of extracellular Ca<sup>2+</sup> (Sr<sup>2+</sup> 1.8 mmol/l + Ca<sup>2+</sup> 0.2 mmol/l) *n* = 6, \*, significantly different from control, *p* ≤ 0.05. Open squares: control. Filled circles: Sr<sup>2+</sup>. Data point reflecting contraction duration (A) or contraction force (B) in Sr<sup>2+</sup>-medium at the highest frequency (3.33 Hz) was omitted since APD was longer than the cycle length (300 ms), hence the real frequency was lower. A: Effect of Sr<sup>2+</sup> on contraction duration at various stimulation frequencies (in the case of biphasic contractions, the values represent total duration). B: Effect of Sr<sup>2+</sup> on contraction force at various stimulation frequencies (in the case of biphasic contractions, the values represent peak force of the contraction, i.e., peak force of the first component). Contraction force was standardized to contraction at 1 Hz in control.

first component at 0.2 Hz, 39.9 ± 4.9% at 0.1 Hz and 48.8 ± 6.8% at 0.03 Hz. It is possible that both components were also present at higher frequencies but they were not clearly distinguishable. The time to peak of contraction was prolonged at any frequency tested, e.g. 103.3 ± 7.8 ms (*n* = 6) in control and 165.8 ± 12.9 ms in Sr<sup>2+</sup>-containing solution at 0.1 Hz. The contraction force was reduced at high stimulation frequencies (2 and 1.43 Hz), while an increase was observed at low frequencies (0.2 and 0.1 Hz; Fig. 2B).

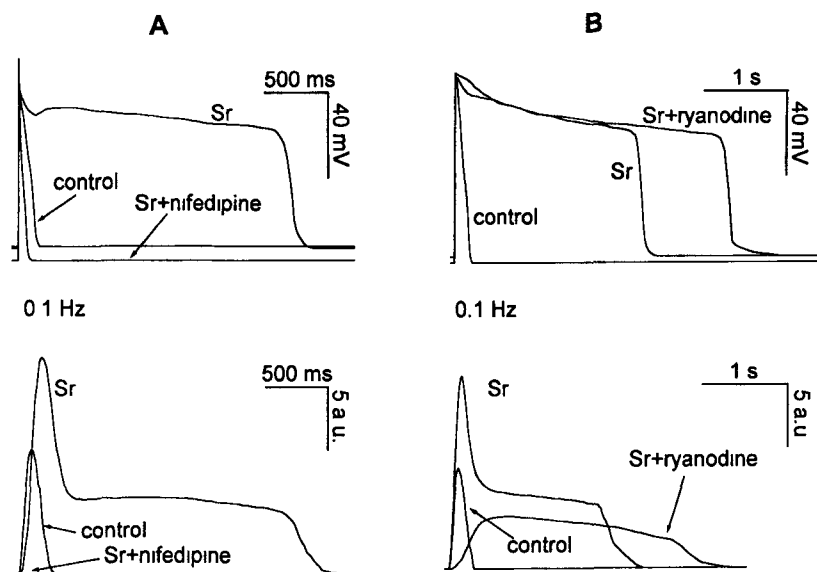
To investigate the mechanisms underlying the effect of Sr<sup>2+</sup>, the following experiments were performed. When the Sr<sup>2+</sup> concentration in the experimental solution was increased from 1.8 to 5.8 mmol/l (0.2 mmol/l Ca<sup>2+</sup> present in both



**Figure 3.** Modulation of the Sr<sup>2+</sup> effect by Ca<sup>2+</sup> and by Sr<sup>2+</sup>. Top panels: APs. Bottom panels: Contractions. Stimulation frequency 0.1 Hz. *A*: Increasing the Sr<sup>2+</sup> concentration (from 1.8 to 5.8 mmol/l with 0.2 mmol/l Ca<sup>2+</sup> present all the time) further enhanced the effect. *B*: Increasing the Ca<sup>2+</sup> concentration from 0.2 to 2 mmol/l (simultaneously decreasing the Sr<sup>2+</sup> concentration from 5.8 to 4 mmol/l to keep the total concentration constant) suppressed the effect.

solutions) the above described changes of both AP and contraction became more obvious (Fig. 3A). Interestingly, these effects were blocked by the application of a solution containing 2 mmol/l Ca<sup>2+</sup> and 4 mmol/l Sr<sup>2+</sup> (Fig. 3B). The application of solution with Ca<sup>2+</sup> totally omitted and substituted by Sr<sup>2+</sup> (2 or 6 mmol/l) led to permanent depolarization and refractery. The application of nifedipine (10  $\mu$ mol/l) blocked the Sr<sup>2+</sup>-induced prolongation of AP and totally suppressed the contraction (Fig. 4A). Ryanodine (1  $\mu$ mol/l), a selective blocker of sarcoplasmic reticulum release channels, eliminated the first contraction component and induced a further prolongation of AP and of the second contraction component (Fig. 4B).

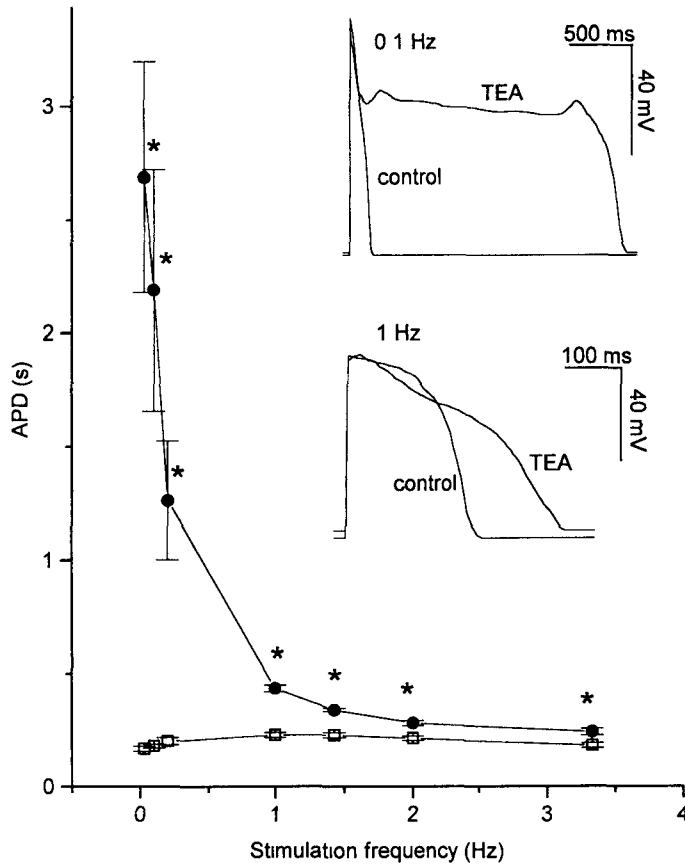
TEA<sup>+</sup> (20 mmol/l) induced a prolongation of AP as expected. The effect was stimulation frequency-dependent (Fig. 5), small at high frequencies (1 Hz and higher) and very pronounced at low ones (0.2 Hz and less). While APD at the stimulation frequency of 1 Hz increased from  $228.0 \pm 11.6$  ms ( $n = 6$ ) to only  $435.0 \pm 14.8$  ms, at 0.1 Hz APD changed from  $180.0 \pm 8.9$  ms to  $2190.0 \pm 533.0$  ms. Also, the shape of AP in the presence of TEA<sup>+</sup> was strikingly similar to that in



**Figure 4.** Pharmacological modulation of the Sr<sup>2+</sup> effect. Top panels: APs. Bottom panels: Contractions. Stimulation frequency: 0.1 Hz. **A**: Nifedipine (10  $\mu$ mol/l) eliminated Sr<sup>2+</sup>-dependent prolongation of AP and totally suppressed the contraction. **B**: Ryanodine (1  $\mu$ mol/l) eliminated the first contraction component only, while AP and the second contraction component became more prolonged.

the presence of Sr<sup>2+</sup>; the dominant phase 2 (plateau) terminated by a relatively fast terminal repolarization (Fig. 5, insets). RMP did not change significantly (e.g.,  $-73.0 \pm 4.1$  mV in the absence and  $-76.8 \pm 2.5$  mV in the presence of TEA<sup>+</sup> at 0.1 Hz;  $n = 6$ ).

In contrast to the considerable effect of TEA<sup>+</sup> on APD, the contraction time course was not influenced markedly. Although a mild prolongation (15–20%) was observed at stimulation frequencies of 1 Hz and 0.5 Hz, it was by far not comparable with the Sr<sup>2+</sup> effect (Figs. 6A, 2A). The late contraction component did not appear at any stimulation frequency. The time to peak of contraction remained also unchanged in the presence of TEA<sup>+</sup> at any frequency tested (e.g.,  $102.5 \pm 7.0$  ms and  $104.2 \pm 7.2$  ms at 0.1 Hz;  $n = 6$ ). The contrast between the effects of TEA<sup>+</sup> on APD and contraction duration is well documented in Fig. 7. It shows that the contraction duration is independent on the TEA<sup>+</sup>-induced prolongation of AP. On the other hand, the Sr<sup>2+</sup>-dependent prolongation of AP correlates well with the contraction prolongation. At low frequencies (1 Hz and less), the application of TEA<sup>+</sup> elicited a pronounced positive inotropic effect (Fig. 6B) e.g., at 0.1 Hz

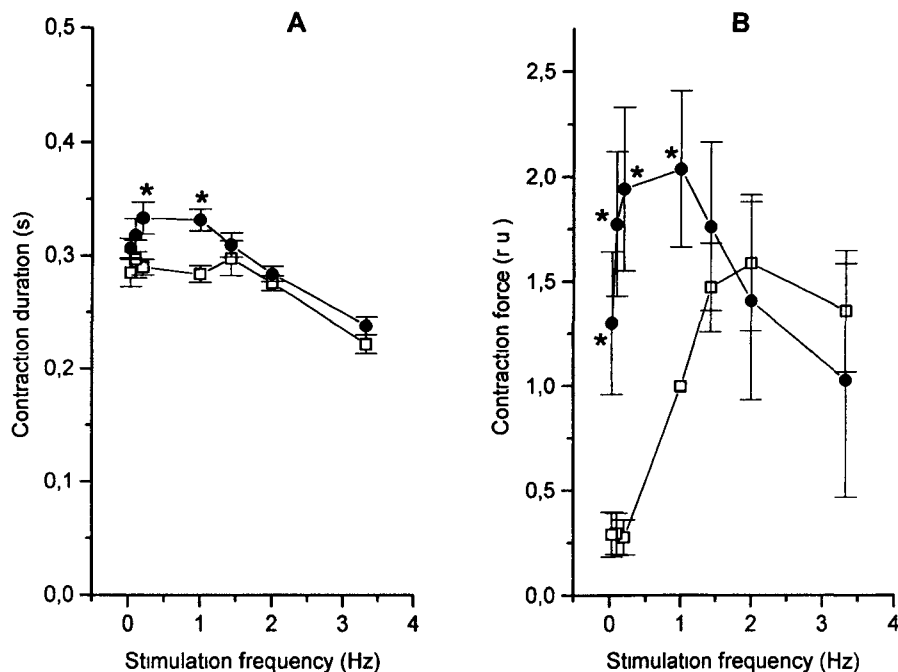


**Figure 5.** Effect of  $\text{TEA}^+$  on APD at various stimulation frequencies  $n = 6$ , \*, significantly different from control,  $p \leq 0.05$ . Open squares control. Filled circles  $\text{TEA}^+$ . The effect of  $\text{TEA}^+$  increases with the decreasing frequency. Note the similarity with the effect of  $\text{Sr}^{2+}$ . Top inset: Effect of  $\text{TEA}^+$  on AP at the stimulation frequency of 0.1 Hz. Bottom inset: Effect of  $\text{TEA}^+$  on AP at the stimulation frequency of 1 Hz.

$\text{TEA}^+$  increased the contraction force from  $0.3 \pm 0.1$  r.u. ( $n = 6$ ; standardized to the contraction force at 1 Hz in control) to  $1.8 \pm 0.3$  r.u. On the other hand, at high frequencies (3.3 and 2 Hz), there was a tendency to reduced contraction force.

To obtain some additional information about the possible role of the reverse mode  $\text{Na}^+/\text{Ca}^{2+}$  exchange in the development of the late contraction component, we performed experiments with the reverse mode enhanced by increasing  $\text{Ca}_e^{2+}$  (up to 10 mmol/l) or decreasing  $\text{Na}_e^+$  (70 mmol/l, completed by sucrose). Interestingly, these interventions completely suppressed the  $\text{TEA}^+$ -dependent AP prolongation



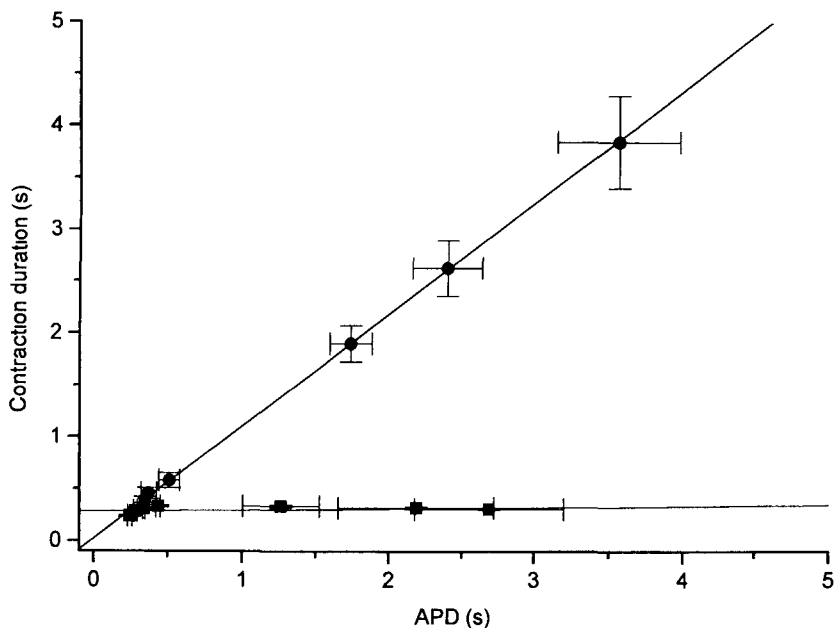


**Figure 6.** Effect of TEA<sup>+</sup> on contraction  $n = 6$ , \*, significantly different from control,  $p \leq 0.05$  Open squares control Filled circles TEA<sup>+</sup> A Effect of TEA<sup>+</sup> on contraction duration at various stimulation frequencies B Effect of TEA<sup>+</sup> on contraction force at various stimulation frequencies Contraction force was standardized to contraction at 1 Hz in control

(Fig. 8A, B). Both interventions would be expected to increase Ca<sub>i</sub><sup>2+</sup> and subsequently the contraction force. Regarding this issue, the results were somewhat controversial. We observed reduction, increase as well as no change of the contraction force. It may be related to the fact that tension was not measured continuously in our experiments, and the resting tension level was reset to zero before each record. An increase of the resting tension could then be responsible for the apparent reduction or no change of active contraction.

## Discussion

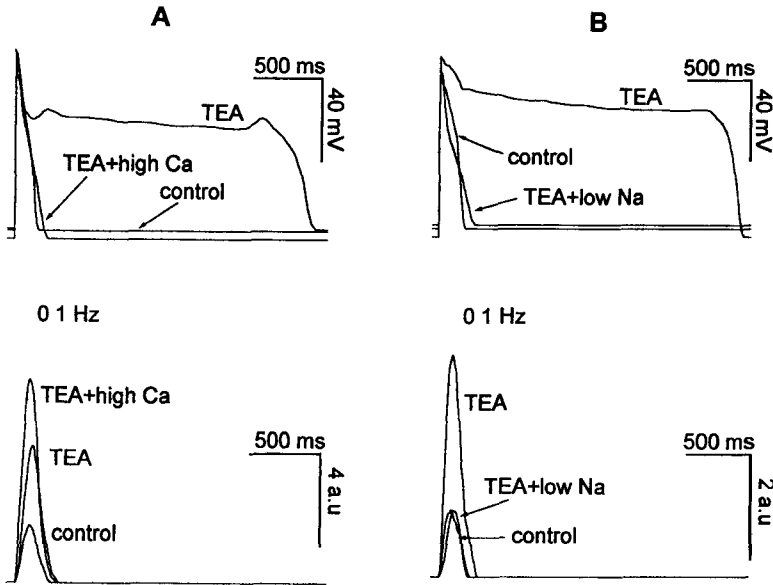
The present results confirm the findings of Bravený and Šumbera (1972) and King and Bose (1983) that at low stimulation frequencies, partial substitution of Ca<sub>e</sub><sup>2+</sup> by Sr<sup>2+</sup> induces an AP prolongation as well as the development of biphasic contraction. Furthermore, we observed a frequency-dependence of these effects in the wide range



**Figure 7.** Dependence of contraction duration on APD in the presence of TEA<sup>+</sup> or Sr<sup>2+</sup> ( $n = 6$ ). Squares: TEA<sup>+</sup>; Circles: Sr<sup>2+</sup>. In the presence of Sr<sup>2+</sup>, APD and contraction duration were prolonged correspondingly and correlated well. On the other hand, TEA<sup>+</sup> prolonged AP but not contraction, hence contraction duration was independent on APD.

of stimulation frequencies (from 0.03 Hz to 3.33 Hz) and suggested that the reverse mode Na<sup>+</sup>/Ca<sup>2+</sup> exchange plays only a negligible role in the development of the late component of the biphasic contraction which is therefore mainly due to Sr<sup>2+</sup> influx through  $I_{CaL}$  channels. Another new finding is that the effect of TEA<sup>+</sup>, a potassium conductance blocker, is antagonized by interventions increasing Ca<sub>i</sub><sup>2+</sup> which could play a role in the reverse use dependence of the TEA<sup>+</sup> effect.

Replacing 90% of Ca<sup>2+</sup> in the extracellular solution with Sr<sup>2+</sup> (1.8 mmol/l Sr<sup>2+</sup> + 0.2 mmol/l Ca<sup>2+</sup>) induced a marked prolongation of AP. The effect was even more pronounced when the Sr<sup>2+</sup> concentration was increased (5.8 mmol/l Sr<sup>2+</sup> + 0.2 mmol/l Ca<sup>2+</sup>). Since the inactivation of  $I_{CaL}$  is not only voltage- but also Ca<sup>2+</sup>-dependent (for review, see Pelzer et al 1990), the rate of inactivation decreases when cations other than Ca<sup>2+</sup> carry the current (Hess et al 1986, McDonald et al 1986). A slower inactivation of  $I_{CaL}$  and subsequently longer influx of Sr<sup>2+</sup> could explain the observed AP prolongation. Another possibility is a block of repolarizing currents. Certainly, the Ca<sup>2+</sup>-activated currents, in the heart first of all the Ca<sup>2+</sup>-dependent transient outward current (Sipido et al 1993), will be sup-



**Figure 8.** Modulation of TEA<sup>+</sup> effect by interventions increasing Ca<sup>2+</sup>. Top panels APs Bottom panels Contractions Stimulation frequency 0.1 Hz *A* The effect of TEA<sup>+</sup> on AP was blocked in the presence of high extracellular Ca<sup>2+</sup> (10 mmol/l) *B* Application of extracellular solution with low Na<sup>+</sup> concentration (70 mmol/l) suppressed the effect of TEA<sup>+</sup> on AP

pressed. Furthermore, in frog skeletal muscle Ba<sup>2+</sup> and Sr<sup>2+</sup> were reported to block the inwardly rectifying K<sup>+</sup> conductance (Standen and Stanfield 1978). Nevertheless, Sr<sup>2+</sup> was around 400 times less effective than Ba<sup>2+</sup>, and should not have any major effect at concentrations used in our experiments. The results are in favour of the first hypothesis: the Sr<sup>2+</sup> effect was blocked by nifedipine, a selective blocker of I<sub>CaL</sub>, as well as by raising the Ca<sup>2+</sup> concentration in external solution from 0.2 to 2 mmol/l (while decreasing the Sr<sup>2+</sup> concentration from 5.8 to 4 mmol/l to keep the total concentrations of both divalent cations constant). Although suppression of the effect by raising Ca<sub>e</sub><sup>2+</sup> does not allow to distinguish between the above mentioned possibilities (increased Ca<sup>2+</sup> would both enhance the Ca<sup>2+</sup>-dependent inactivation of I<sub>CaL</sub> and activate the Ca<sup>2+</sup>-dependent repolarizing currents), experiments with nifedipine strongly support the first hypothesis. Since nifedipine is a selective blocker of I<sub>CaL</sub>, suppression of the Sr<sup>2+</sup> effect is consistent with the first hypothesis. On the other hand, nifedipine was reported to inhibit transient outward K<sup>+</sup> current (Gotoh et al. 1991) and therefore, if the Sr<sup>2+</sup> effect was related to a block of repolarizing currents, nifedipine should promote the effect of Sr<sup>2+</sup> or have

no effect at all which was not the case. Further support is provided by the fact that, in  $\text{Sr}^{2+}$ -treated preparation, APD and contraction duration always correlate very well suggesting a substantial influx of divalent cation, activator of contraction, during all AP.

The contraction in  $\text{Sr}^{2+}$ -containing medium became biphasic, with the first, fast and the second, slow components. Ryanodine, a blocker of sarcoplasmic reticulum release channels, suppressed the first component but did not influence the second one. The results suggest that the first component is caused by release of activator cation from the sarcoplasmic reticulum, which is in good accordance with the data of King and Bose (1983) obtained in dog trabeculae. The time to peak of contraction in  $\text{Sr}^{2+}$ -treated muscle (i.e., the time to peak of the first component) was always longer than in control suggesting a lower efficiency in activation of sarcoplasmic reticulum release (if we admit that  $\text{Sr}^{2+}$  and  $\text{Ca}^{2+}$  are equally efficient in activating cardiac myofibrils as shown by Kerrick et al. 1980). However, we are not able to distinguish whether the activator of the release is  $\text{Sr}^{2+}$  (then it must be a weaker activator than  $\text{Ca}^{2+}$ ) or  $\text{Ca}^{2+}$  (a lower efficiency due to lower external  $\text{Ca}^{2+}$  concentration). In a recent study Spencer and Berlin (1997) demonstrated that  $\text{Ca}^{2+}$  influx, but not  $\text{Sr}^{2+}$  influx, via sarcolemmal  $I_{\text{CaL}}$  channels can induce  $\text{Sr}^{2+}$  release from the sarcoplasmic reticulum in rat ventricular myocytes; therefore, the second possibility appears to be more probable.

With regard to the late contraction component, its mechanism remained obscure. There was an agreement that external  $\text{Sr}^{2+}$  entering the cell during the prolonged AP is responsible but the pathway remained unclear. King and Bose (1983) concluded that  $I_{\text{CaL}}$  channel is a probable candidate but they admit that at least a part of the late component could result from the  $\text{Na}^+/\text{Ca}^{2+}$  exchange. The exchanger is able to transport  $\text{Sr}^{2+}$  instead of  $\text{Ca}^{2+}$  ( $\text{Na}^+/\text{Sr}^{2+}$  exchange, Kimura et al. 1987). If, during the plateau phase, the membrane potential is more positive than the reversal potential of the exchange, the exchanger would work in the reverse mode, thus transporting  $\text{Sr}^{2+}$  into the cell. Experiments with nifedipine, a selective blocker of  $I_{\text{CaL}}$  channels, seem to confirm the conclusion of King and Bose (1983), that  $I_{\text{CaL}}$  is the main pathway. One has to keep in mind, however, that nifedipine, by blocking the AP prolongation, would also prevent the reverse mode of the  $\text{Na}^+/\text{Ca}^{2+}$  exchange, and therefore, these experiments do not provide sufficient information. To distinguish between  $I_{\text{CaL}}$  and  $\text{Na}^+/\text{Ca}^{2+}$  exchange, it would help to depolarize the membrane as in the presence of  $\text{Sr}^{2+}$ , but without affecting  $I_{\text{CaL}}$ . This is possible either pharmacologically (by blocking the repolarizing currents) or by artificial depolarizing current as in sucrose gap experiments by Bravený and Šumbera (1970). In their experiments, the artificial AP prolongation was accompanied by sustained contraction, but the membrane potential was held at potentials more positive than the plateau potentials in the presence of  $\text{Sr}^{2+}$ . In our experiments, we used the pharmacological approach: block of repolarizing cur-

rents by TEA<sup>+</sup> TEA<sup>+</sup> was described to prolong AP in canine and sheep Purkinje fibers (Ito and Surawicz 1981, Kenyon and Gibbons 1979, respectively) or in guinea pig papillary muscles (Ochi and Nishiye 1974) The underlying mechanism is probably a block of time-dependent outward K<sup>+</sup> currents, transient outward current,  $I_{to}$ , and delayed rectifier,  $I_K$  (Kass et al 1982, Kenyon and Gibbons 1979), while the background K<sup>+</sup> current,  $I_{K1}$ , does not seem to be influenced significantly (Ito and Surawicz 1981) We characterized the TEA<sup>+</sup> effect and found that, concerning AP, it is strikingly similar to the effect of Sr<sup>2+</sup> The late contraction component, however, never developed suggesting that the Ca<sup>2+</sup> influx via the reverse mode Na<sup>+</sup>/Ca<sup>2+</sup> exchange during prolonged AP plateau is not sufficient to activate the contraction If we then assume that concentration gradients for Ca<sup>2+</sup> in TEA<sup>+</sup> experiments and for Sr<sup>2+</sup> in Sr<sup>2+</sup> experiments (subsequently the reversal potentials of the Na<sup>+</sup>/Ca<sup>2+</sup> and Na<sup>+</sup>/Sr<sup>2+</sup> exchangers) are similar, it is possible to conclude that the role of Na<sup>+</sup>/Sr<sup>2+</sup> exchange in the development of the late contraction component is negligible

To make it more clear, experiments were performed with the reverse mode Na<sup>+</sup>/Ca<sup>2+</sup> exchange enhanced by increasing Ca<sub>e</sub><sup>2+</sup> or decreasing Na<sub>e</sub><sup>+</sup> Interestingly, these interventions did not produce the late component but suppressed the TEA<sup>+</sup>-dependent AP prolongation This effect is probably related to an increase of Ca<sub>i</sub><sup>2+</sup> which is known to modulate many membrane currents One possibility is that increased intracellular Ca<sup>2+</sup> would stimulate  $I_{to}$ , its Ca<sup>2+</sup>-sensitive component (Coraboeuf and Carmeliet 1982, Sipido et al 1993) and/or  $I_K$  which was also reported to be Ca<sup>2+</sup>-sensitive (Tohse et al 1987) Other explanation could be the existence of Ca<sup>2+</sup> dependent K<sup>+</sup> current,  $I_{K(Ca)}$  Such a current is well known in arterial smooth muscle (for review, see Nelson and Quayle 1995) but there is no evidence for its existence in the heart ventricle although it was reported in Purkinje fibers (Callewaert et al 1986) and atrial myocytes (Baro and Escande 1989) This phenomenon could also help explain the reverse use dependence of the TEA<sup>+</sup> effect Since Ca<sub>i</sub><sup>2+</sup> is increased at high frequencies, the effect of TEA<sup>+</sup> is antagonized, while at low frequencies when Ca<sub>i</sub><sup>2+</sup> is decreased the TEA<sup>+</sup> effect becomes more pronounced Beside the effect on AP, TEA<sup>+</sup> had a positive inotropic effect at low stimulation frequencies It is probably due to Ca<sup>2+</sup> influx (and subsequently increased sarcoplasmic reticulum Ca<sup>2+</sup> load) via the Na<sup>+</sup>/Ca<sup>2+</sup> exchange working in reverse mode during long lasting AP This influx, however, has to be quite small, since it is not able to induce the late contraction component

## References

- Armstrong C M (1971) Interaction of tetraethylammonium ion derivatives with potassium channels of giant axons *J Gen Physiol* **58**, 413–437
- Baro I, Escande D (1989) A long lasting Ca<sup>2+</sup>-activated outward current in guinea-pig atrial myocytes *Pflugers Arch* **415**, 63–71

- Bravený P, Šumbera J (1970) Electromechanical correlations in the mammalian heart muscle *Pflugers Arch* **319**, 36—48
- Bravený P, Šumbera J (1972) Biphasic activation of the myocardial contraction *Physiol Bohemoslov* **21**, 73—74
- Callewaert G, Vereecke J, Carmeliet E (1986) Existence of a calcium-dependent potassium channel in the membrane of cow cardiac Purkinje cells *Pflugers Arch* **406**, 424—426
- Coraboeuf E, Carmeliet E (1982) Existence of two transient outward currents in sheep cardiac Purkinje fibers *Pflugers Arch* **392**, 352—359
- Gotoh Y, Imazumi Y, Watanabe M, Shibata E F, Clark R B, Giles W R (1991) Inhibition of transient outward  $K^+$  current by DHP  $Ca^{2+}$  antagonists and agonists in rabbit cardiac myocytes *Amer J Physiol* **260**, H1737—H1742
- Hess P, Lansman J B, Tsien R W (1986) Calcium channel selectivity for divalent and monovalent cations: Voltage and concentration dependence of single channel current in ventricular heart cells *J Gen Physiol* **88**, 293—319
- Ito S, Surawicz B (1981) Effect of tetraethylammonium chloride on action potential in cardiac Purkinje fibers *Amer J Physiol* **241**, H139—H144
- Kass R S, Scheuer T, Malloy K J (1982) Block of outward current in cardiac Purkinje fibers by injection of quaternary ammonium ions *J Gen Physiol* **79**, 1041—1063
- Kenyon J L, Gibbons W R (1979) Influence of chloride, potassium, and tetraethylammonium on the early outward current of sheep cardiac Purkinje fibers *J Gen Physiol* **73**, 117—138
- Kerrick W G L, Malencik D A, Hoar P E, Potter P D, Coby R L, Pociwong S, Fischer E H (1980)  $Ca^{2+}$  and  $Sr^{2+}$  activation: Comparison of cardiac and skeletal muscle contraction models *Pflugers Arch* **386**, 207—213
- Kimura J, Miyamae S, Noma A (1987) Identification of sodium-calcium exchange current in single ventricular cells of guinea-pig *J Physiol (London)* **384**, 199—222
- King B W, Bose D (1983) Mechanism of biphasic contractions in strontium-treated ventricular muscle *Circ Res* **52**, 65—75
- Kohlhardt M, Haastert H P, Krause H (1973) Evidence of non-specificity of the Ca channel in mammalian myocardial fibre membranes: Substitution of Ca by Sr, Ba or Mg as charge carrier *Pflugers Arch* **342**, 125—136
- Kondo N (1987) Can strontium replace calcium as an activator of internal calcium release in cardiac muscles? Study on a dual action of A23187 *J Mol Cell Cardiol* **19**, 391—397
- McDonald T F, Cavalé A, Trautwein W, Pelzer D (1986) Voltage-dependent properties of macroscopic and elementary calcium channel currents in guinea pig ventricular myocytes *Pflugers Arch* **406**, 437—448
- Nelson M T, Quayle J M (1995) Physiological roles and properties of potassium channels in arterial smooth muscle *Amer J Physiol* **268**, C799—C822
- Ochi R, Nishiye H (1974) Effect of intracellular tetraethylammonium on action potential in guinea pig's myocardium *Pflugers Arch* **348**, 305—316
- Pelzer D, Pelzer S, McDonald T F (1990) Properties and regulation of calcium channels in muscle cells *Rev Physiol Biochem Pharmacol* **114**, 107—207
- Pučelík P, Fiala P, Králíček P (1983) Electromechanical relationships of rabbit papillary muscle under interpolated extrasystole conditions and after a pause *Physiol Bohemoslov* **32**, 295—306

- Sipido K R , Callewaert G , Carmeliet E (1993) [Ca<sup>2+</sup>]<sub>i</sub> transients and [Ca<sup>2+</sup>]<sub>i</sub>-dependent chloride currents in single Purkinje cells from rabbit heart *J Physiol (London)* **468**, 641—667
- Spencer C I , Berlin J R (1997) Calcium-induced release of strontium ions from the sarcoplasmic reticulum of rat cardiac ventricular myocytes *J Physiol (London)* **504**, 565—578
- Standen N B , Stanfield P R (1978) A potential- and time-dependent blockade of inward rectification in frog skeletal muscle fibers by barium and strontium ions *J Physiol (London)* **280**, 169—191
- Stanfield P R (1983) Tetraethylammonium ions and the potassium permeability of excitable cells *Rev Physiol Biochem Pharmacol* **97**, 1—67
- Tasaki I , Hagiwara S (1957) Demonstration of two stable potential states in the squid giant axon under tetraethylammonium chloride *J Gen Physiol* **40**, 859—885
- Tohse N , Kameyama M , Irisawa H (1987) Intracellular Ca<sup>2+</sup> and protein kinase C modulate K<sup>+</sup> current in guinea pig heart cells *Amer J Physiol* **253**, H1321—H1324

Final version accepted January 27, 1999