

Short communication

Ferrous-Ascorbate Complexes as Carriers of Nitric Oxide

Z V KUROPTEVA AND M E KUDRYAVTSEV

*Institute of Biochemical Physics, Russian Academy of Sciences
Kosygin Str 4, 117977 Moscow Russia*

Abstract. Ferrous-ascorbate is known to form with nitric oxide paramagnetic nitrosyl ferrous-ascorbate complexes, Fe-AA-NO. These complexes yield an EPR signal with g-factor close to 2.02 and an optical absorption spectrum with maxima at 340, 460, and 600 nm. Fe-AA-NO complexes are unstable in the presence of oxygen. Ferrous-ascorbate complexes promote NaNO_2 decay resulting in the formation of NO. Nitric oxide is taken up by Fe-AA complexes to form paramagnetic ferrous-ascorbate nitrosyl complexes, Fe-AA-NO. It is suggested that ferrous-ascorbate complexes can play the role of carriers of NO and, perhaps, O_2 in the blood plasma. Nitrosyl ferrous-ascorbate complexes can also be the NO-containing factor involved in the blood vessel relaxation (endothelium-derived relaxing factor, EDRF).

Key words: Nitrosyl ferrous-ascorbate complexes — Nitric oxide — Carriers — EDRF

It has been established earlier that many nitrocompounds are reduced in the animal organisms as well as in tissue homogenates to form nitric oxide (Shubin and Kuropteva 1983, Kuropteva and Pastushenko 1985, Zhumabaeva et al 1987, Kuropteva et al 1991). Probably, NO production is an important factor of the nitrocompounds activity at the organism level. A number of works have appeared in recent years concerning the role of nitric oxide as a factor involved in basic processes in man and animals (Palmer et al 1987, Moncada et al 1989, Moncada and Higgs 1990). Also, L-arginine has been shown to be the endogenous NO source (Palmer et al 1988, Moncada et al 1989, Moncada and Higgs 1990). NO is known to be the principal constituent of a factor regulating blood vessel relaxation (endothelium-derived relaxing factor, EDRF) and can act as a central nervous system messenger, cytotoxic action mediator in immunologically activated cells, etc (Moncada et al

Correspondence to Z V Kuropteva, Institute of Biochemical Physics of Russian Academy of Sciences, Kosygin Str , 4, 117977 Moscow, Russia
E-mail: chembio@glas.apc.org

1988, Stuehr and Nathan 1989, De Vente et al 1990, O'Connor et al 1990, Beckman 1991, Drapier et al 1991) How these active molecules can be transferred to the sites of their action remains, however, unclear NO carriers (and EDRF, accordingly) have been shown to contain thiols (Palmer et al 1987, Ignarro 1990), and it was proposed that NO-carriers are Fe-S complexes, and Fe-S-NO complexes were suggested as the hypothetical structure of EDRF (Lancaster and Hibbs 1990, Vanin 1991) Actually, these complexes are easily formed and are sufficiently stable and easily registered by EPR technique In our opinion, however, Fe-S-NO complexes are too stable to play the carrier role They actually are formed in the organism but may play the role of NO scavengers These complexes may serve the elimination of excessive NO molecules from the organism or may represent NO storage

Herein, we present data concerning another type of complexes which can play the role as NO and, possibly, O₂ carriers ferrous-ascorbate complexes which form with NO nitosyl ferrous-ascorbate complexes, Fe-AA-NO

Materials and Methods

The following chemicals were used FeCl₃ and NaNO₂ from REACHIM Company (Russia), L-ascorbic acid from Sigma (St Louis, MO, USA) The FeCl₃ AA NaNO₂ ratio in water solutions was 1 5 8 All solutions were prepared in argon atmosphere (argon blowing during 2–3 min) The experiments were performed at pH 6.5–8.0 NaNO₂ solution was added to the stock Fe-AA complexes solution and optical absorption spectra were immediately recorded Similar samples were frozen to liquid nitrogen temperature in the form of columns, 3.5 mm in diameter and 30 mm in length, to measure EPR spectra

EPR spectra were recorded with a Bruker ER-300 spectrometer at 77 K Optical absorption measurements were performed with a "Specord UV-VIS" spectrophotometer

Results and Discussion

Fig 1 (curve 1) shows the EPR spectrum of nitosyl ferrous-ascorbate complexes (Fe-AA-NO) in argon atmosphere at 77 K Fe-AA-NO complexes have asymmetric EPR signal with g-factor close to 2.02 During an blowing through mixed solution or during incubation under an atmosphere, the EPR signal disappears Fig 1 (curve 2) shows for comparison the known EPR spectrum of Fe-S-NO nitosyl complexes obtained by mixing FeCl₃ and reduced glutation solution with sodium nitrite

The optical absorption spectra of the same solutions are shown in Fig 2A Spectrum 2A 2 was recorded immediately after solution mixing (within 2–3 min), and Fig 2 A 3 shows the spectrum of the same sample recorded after 5–7 min Optical absorption spectrum 2 A 2 has three expressed absorption maxima 340, 460,

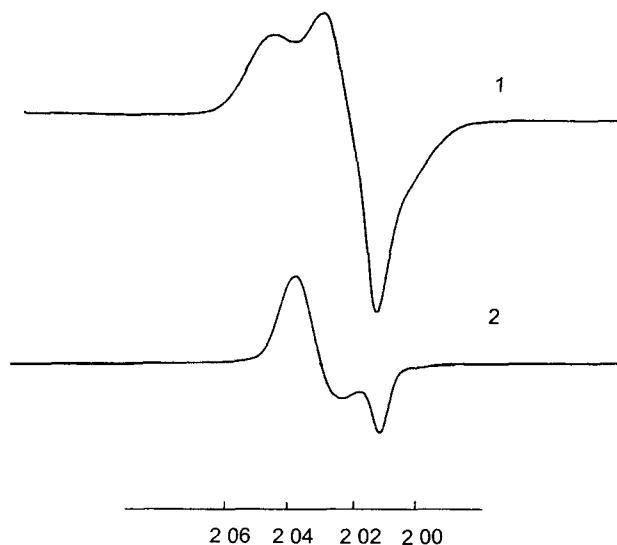


Figure 1. ESR spectra of nitrosyl complexes 1 – ferrous-ascorbate Fe-AA-NO, 2 – ferrous-thiol Fe-S-NO. Settings: microwave power 20 mW, magnetic field modulation 4 G, temperature 77 K. Magnetic field values are given in the g-factor units.

and 600 nm. It can be assumed that this spectrum is due to Fe-AA-NO complexes recorded by EPR method (Fig. 1 curve 1) in the same conditions as the optical spectrum in Fig. 2A.2. Spectrum 2A.3 is the sum of absorptions of two complexes: that of Fe-AA-NO and of a new formed, the spectrum of which was obtained as the difference between absorption spectra 2A.3 and 2A.2. The difference spectrum is shown in Fig. 2B, it has an absorption maximum in the visible region at 400 nm. This complex does not yield any EPR signal. It should be noted that ascorbic acid has no absorption in the wavelength range studied, and FeCl₃ and AA mixture absorption spectra are represented in Fig. 2A.1. Spectra 2A.2 and 2A.3 were recorded with the reference cell containing NaNO₂ solution.

As mentioned above, the paramagnetic Fe-AA-NO complexes are unstable in the presence of molecular oxygen, and then EPR signal disappeared after air blowing. There can be two explanations for this effect: either O₂ oxidizes AA and the complex disintegrates, or NO is substituted by molecular oxygen with the formation of diamagnetic Fe-AA-O₂ complex. We obtained the data supporting the latter suggestion (unpublished data).

Thus, we could show that ferrous-ascorbate complexes Fe-AA can be formed at pH values close to physiological ones. These complexes promote NaNO₂ decay giving rise to NO. Nitric oxide is taken up by Fe-AA complexes, resulting in the formation of paramagnetic nitrosyl ferrous-ascorbate complexes Fe-AA-NO, with

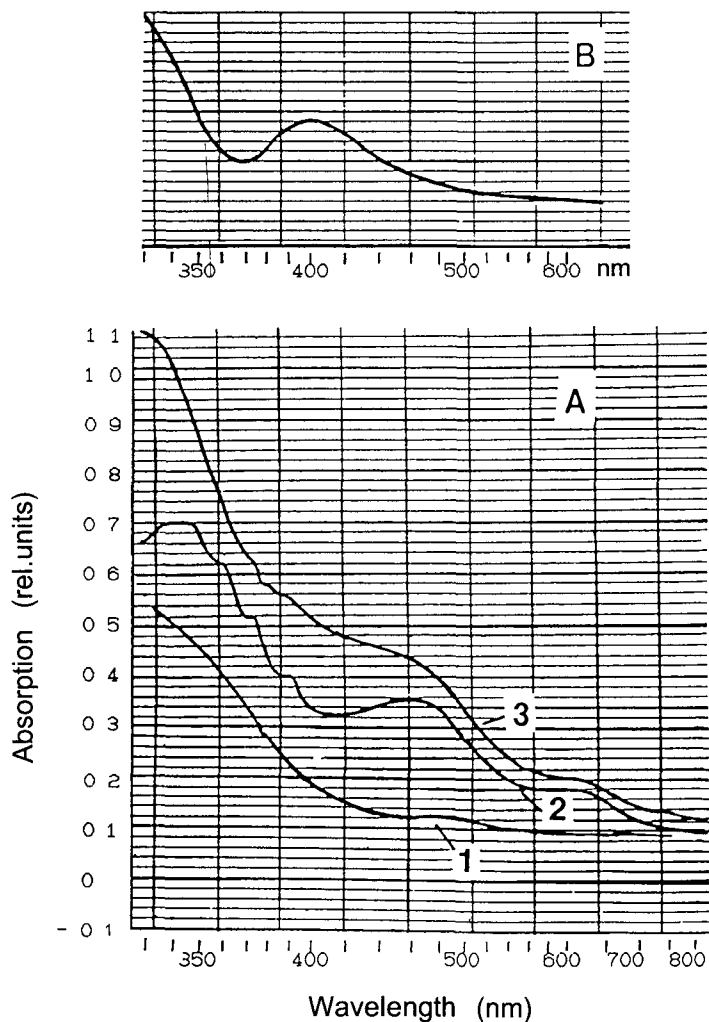


Figure 2. Optical absorption spectra A 1 – FeCl_3 and ascorbic acid solution (Fe-AA complexes), A 2 – after addition of sodium nitrite to A 1, A 3 – 5–7 min after recording of A 2 (in air) B – difference spectrum between A 3 and A 2

g-factor near 2.02. In our opinion, ferrous-ascorbate complexes can play a role as carriers of NO and, perhaps, O_2 in the blood plasma. Ferrous-ascorbate nitrosyl complexes can also be the NO-containing factor that is involved in the blood vessel relaxation (endothelium-derived relaxing factor, EDRF), the structure of which is widely discussed (Palmer et al 1987, Moncada et al 1989, Ignarro 1990, Moncada and Higgs 1990, Vanin 1991).

Acknowledgements. The authors wish to thank prof L A Blumenfeld for useful discussion

References

- Beckman J S (1991) The double-edged role of nitric oxide in brain function and superoxide-mediated injury *J Develop Physiol* **15**, 53–59
- De Vente J, Bol J G M, Berkelmans H S, Schipper J, Steinbusch H M W (1990) Immunocytochemistry of cGMP in the cerebellum of immature, adult and aged rat the involvement of nitric oxide *Eur J Neurosci* **2**, 845–862
- Drapier J-C, Pellat C, Henry Y (1991) Generation of EPR-detectable nitrosyl-iron complexes in tumor target cells cocultured with activated macrophages *J Biol Chem* **266**, 10162–10167
- Ignarro L J (1990) Biosynthesis and metabolism of endothelium-derived nitric oxide *Annu Rev Pharmacol Toxicol* **30**, 535–560
- Kuropteva Z V, Pastushenko O N (1985) Change in paramagnetic blood and liver complexes in animals under the influence of glyceryl trinitrate *Dokl Acad Sci USSR* **281**, 189–191 (in Russian)
- Kuropteva Z V, Yushmanov V E, Jurczyk M U, Khristianovich D S, Kudryavtsev M E, Pisarski T, Sibeldina L A (1991) ESR and ³¹P-NMR study of metabolic changes in mouse tumor and liver after combined misonidazole and irradiation treatment *Appl Magnet Resonance* **2**, 495–510
- Lancaster J R, Hibbs J B (1990) EPR demonstration of iron-nitrosyl complex formation by cytotoxic activated macrophages *Proc Natl Acad Sci USA* **87**, 1223–1227
- Moncada S, Higgs E A (Eds) (1990) Nitric Oxide from L-arginine A Bioregulatory System Elsevier Science Publishers B V, Amsterdam
- Moncada S, Radomski M W, Palmer R M J (1988) Endothelium-derived relaxing factor identification as nitric oxide and role in the control of vascular tone and platelet function *Biochem Pharmacol* **37**, 245–249
- Moncada S, Palmer R M J, Higgs E A (1989) Biosynthesis of nitric oxide from L-arginine A pathway for regulation of cell function and communication *Biochem Pharmacol* **38**, 1709–1715
- O'Connor K J, Knowles K D, Patel K D, Palmer R M J, Moncada S (1990) Nitric oxide has proliferative as well as antiproliferative effects (Abstract) *Arch Pharm Int Ther* **305**, 270
- Palmer R M J, Ferrige A G, Moncada S (1987) Nitric oxide release accounts for the biological activity of endothelium-derived relaxing factor *Nature* **327**, 524–526
- Palmer R M J, Ashton D S, Moncada S (1988) Vascular endothelial cells synthesize nitric oxide from L-arginine *Nature* **333**, 664–666
- Shubin V E, Kuropteva Z V (1983) EPR study of NO generation during reduction of nitrofurans and nitroimidazoles I Hemoglobin solutions *Stud Biophys* **97**, 157–164
- Stuechli D J, Nathan C F (1989) Nitric oxide a macrophage product responsible for cytostasis and respiratory inhibition in tumor target cells *J Exp Med* **169**, 1543–1555
- Vanin A F (1991) Endothelium-derived relaxing factor is a nitrosyl iron complex with thiol ligands (Hypothesis) *FEBS Lett* **289**, 1–3

Zhumabaeva T T, Kudryavtsev M E, Voronina S S, Kuropteva Z V (1987) The influence of metronidazole and local tumor irradiation on paramagnetic metallo-complexes of liver and tumor tissues *Radiobiologiya* **27**, 384—389 (in Russian)

Final version accepted January 28, 1997