

Minireview

The Role of Angiotensin II and its Receptors in Regulation of Adipose Tissue Metabolism and Cellularity

S. ZORAD¹, M. FICKOVA¹, B. ZELEZNA², L. MACHO¹ and J. G. KRAL³

1 Institute of Experimental Endocrinology, Slovak Academy of Sciences, Vlárská 3, 833 06 Bratislava, Slovakia

2 Institute of Molecular Genetics, Academy of Sciences of the Czech Republic, Flemingovo nám. 2, 16637 Prague, Czech Republic

3 State University of New York, Health Science Center at Brooklyn, 450 Clarkson Avenue, Brooklyn, New York 11203, USA

Abstract. Angiotensin II exerts its action via at least two distinct receptor subtypes designated AT₁ and AT₂. AT₁ receptors seem to be responsible for most of the known angiotensin II effects while the role of AT₂ receptors is not yet clear. Adipocytes of adult rats express exclusively the AT₁ subtype. Angiotensin II stimulates prostacyclin release in adult rat adipocytes and in mouse preadipocytes. In the latter prostacycline release is completely blocked by an AT₂ receptor antagonist. Adipocyte angiotensin II receptors seem to be regulated by age and fat mass. Blockade of these receptors by an AT₁ antagonist seems to prevent adipose tissue hypertrophy. Moreover, adipose tissue contains all the main components of the renin-angiotensin system such as angiotensinogen, angiotensin converting enzyme, angiotensin II and angiotensin II receptors. Angiotensinogen expression in adipocytes is stimulated by a high fat diet concurrent with enlargement of fat mass, associated with insulin resistance. Angiotensin converting enzyme inhibitors improve insulin sensitivity. Taken together, there is evidence of interaction between insulin and angiotensin II in regulation of adipose tissue metabolism and cellularity. Clarification of these interactions could lead to significant progress in pharmacological treatment of obesity and its comorbidity.

Key words: Angiotensin receptor subtypes — Renin-angiotensin system — Adipocytes — Obesity — Insulin resistance

Introduction

Numerous dietary, genetic and hormonal factors are known to regulate the amount of adipose tissue in mammals (Hollenberg 1990; Kissebah and Krakower 1994). Excess of adipose tissue mass, obesity, is often associated with noninsulin dependent diabetes mellitus and hypertension (Hollenberg 1990). Recent evidence has implicated angiotensin II (A II) in the regulation of adipose tissue cellularity (Crandall et al. 1994b; Darimont et al. 1994). This idea is supported by the presence of the main components of the renin – angiotensin system (RAS) in adipose tissue. In the rat, adipose tissue seems to be the largest source of angiotensinogen (A O G E N), a precursor of angiotensin I (A I) (Frederich et al. 1992). The A I is converted to A II by angiotensin converting enzyme (ACE), a dipeptidyl carboxypeptidase, which is expressed in the stroma – vascular tissue of fat pads as well (Crandall et al. 1992). Moreover, A II binding sites have recently been characterized in membranes of adipocytes (Crandall et al. 1993).

Generally, A II binds to two structurally and pharmacologically different receptor subtypes AT₁ and AT₂ (Bottari et al. 1993). So far the AT₁ subtype has been detected in rat adipocytes (Crandall et al. 1993), while AT₂ was found in mouse preadipocytes (Darimont et al. 1994). A II, a well-known vasoconstrictor, seems in addition to regulate growth and development (Timmermans et al. 1993). Adipose tissue is one of the few tissues capable of expanding through most of adult life (Klyde and Hirsh 1979). The presence of many components of the RAS in adipose tissue and the possible function of A II in growth and development led to investigate the role of components of the RAS, especially A II and A II receptors, in the regulation of adipocyte metabolism and adipose tissue cellularity.

Mechanism of A II action

A II, an octapeptide, exerts its biological action through two main subtypes of receptors. These receptor subtypes are structurally and biochemically well characterized and they both have been cloned (Murphy et al. 1991; Sasaki et al. 1991; Kambayashi et al. 1993; Mukoyama et al. 1993). The A II receptor subtype, AT₁, is a member of the G-protein-coupled seven-transmembrane receptor class containing 359 amino acids (Murphy et al. 1991; Sasaki et al. 1991; Griendling et al. 1993). In the rat, but not in humans two different AT₁ receptor isoforms termed AT_{1A} and AT_{1B} were found (Chiu et al. 1994). The amino acid sequences of these two receptor isoforms have 96% identity. The only functional consequence of the small difference in amino acid sequence is differential regulation of isoreceptor gene expression (Chiu et al. 1994). The AT₁ receptor subtype seems to mediate most of the known A II actions such as smooth muscle contraction, aldosterone synthesis and secretion, neuropeptide secretion, increase of thirst and salt appetite and

renal sodium reabsorption (Bottari et al. 1993). This A II receptor subtype is believed to be coupled to at least two different G proteins, Gi and Gq (Bottari et al. 1993). Activation of Gi by A II inhibits adenylate cyclase in some tissues (Pobiner et al. 1985). Activation of Gq or more specifically, activation of some so far unknown member of the Gq family by A II leads to stimulation of phospholipase C (phosphoinositidase C) which cleaves phosphatidylinositol 4,5 bisphosphate (PIP₂), originated in phosphatidylinositol, into inositol-1,4,5,-trisphosphate (IP₃) and 1,2-diacylglycerol (DAG) (Berridge and Irvine 1989). IP₃ causes an increase of intracellular calcium concentration whereas DAG activates protein kinase C (PKC). In addition, AT₁ mediated stimulation of adenylate cyclase (Missale et al. 1989), phospholipase A₂ (Pfeilschifter 1989), and phospholipase D has been described in some tissues (Pfeilschifter et al. 1992).

The other A II receptor subtype, AT₂, seems to belong to the unique family of seven-transmembrane receptors not coupled to G-proteins (Mukoyama et al. 1993). It consists of 363 aminoacids with 32% homology to the AT₁ receptor. The function of AT₂ is not yet known. AT₂ binding sites are more abundant in embryonic and neonatal than adult tissues (Bottari et al. 1993). This may suggest that A II via AT₂ receptors is involved in regulation of differentiation and growth. This conclusion is strengthened by recent findings of Stoll et al. (1995) that AT₂ receptors mediate inhibition of proliferation of coronary endothelial cells. Only sparse data exist concerning AT₂ signal transduction. These suggest AT₂ coupling to particulate guanylate cyclase or phosphotyrosine phosphatase (Bottari et al. 1992; Griendling et al. 1993; Kambayashi et al. 1993).

Besides different structures of AT₁ and AT₂ receptors the pharmacology of ligand and binding is the most apparent difference between these receptors. Currently a number of highly selective AT₁ and AT₂ nonpeptide antagonists/ligands have been developed (Timmermans et al. 1993; Chiu et al. 1994). Losartan {2-*n*-butyl-4-chloro-5-hydroxymethyl-1-[2'-(1H-tetrazol-5-yl)biphenyl-4-yl-methyl]imidazole} and other losartan-like compounds are highly selective for the AT₁ receptor displaying about 10,000-fold greater affinity than for AT₂ (Chiu et al. 1994). Losartan is able to block virtually all of the physiological effects of A II. In addition, the agonist binding to AT₁ receptors is sensitive to dithiothreitol (DTT) (Whitebread et al. 1989; Tsutsumi et al. 1992) and the nonhydrolysable analogs of GTP (Dudley et al. 1990). In turn, AT₂ receptor binding is stimulated by DTT, and generally the AT₂ receptors are insensitive to GTP analogs (Dudley et al. 1991; Zorad et al. 1994). A II binding to AT₂ receptors is insensitive to losartan, but is blocked by AT₂-selective ligands such as peptidic CGP 42112A [nicotinic acid-Tyr-(*N*^E-benzyloxycarbonyl-Arg)Lys-His-Pro-Ile-OH] or nonpeptidic PD 123177 {1-(4-amino-3-methylphenyl)methyl-5-diphenylacetyl-4,5,6,7-tetrahydro-1H-imidazo[4,5-c]pyridine-6-carboxylic acid-2HCl} (Timmermans et al. 1993; Heemskerk et al. 1993; Zorad et al. 1994). The above agents are about 1000–3500-fold more selec-

tive for AT₂ than for AT₁ (Chiu et al. 1994). The use of AT₁ and AT₂ selective ligands is the simplest approach to discriminating between these receptor subtypes, especially since the AT₂ receptor response is not yet known.

Some A II receptors seem to have atypical binding properties with regard to interaction with subtype selective ligands, DTT and analogs of GTP (Timmermans et al. 1993). E.g., they are either not blocked by AT₁/AT₂ blockers or are blocked by AT₂ blockers and are sensitive to nonhydrolyzable analogs of GTP (Tsutsumi and Saavedra 1992). The biological function of "atypical" A II receptors is not known.

Renin-angiotensin system in adipose tissue

Adipose tissue is one of the "nonclassical" RAS tissues expressing high levels of AOPEN (Cassis et al 1988). AOPEN mRNA and secretion in white adipocytes is nutritionally regulated unlike in the liver (Frederich et al. 1992). During fasting the AOPEN mRNA and release from adipocytes is decreased. Refeeding and obesity increase AOPEN secretion from adipocytes (Frederich et al. 1992).

ACE is another RAS component present in adipose tissue, specifically in the stroma-vascular fraction (Crandall et al.1992). In addition A II receptors have recently been identified in rat epididymal adipocytes (Crandall et al. 1993) and mesenteric and retroperitoneal fat depots (Crandall et al. 1994a). Adipocytes from aged rats seem to exclusively express the AT₁ subtype of A II receptors (Fig. 1). However, Darimont et al. (1994) showed AT₂ receptors to be present in differentiated adipocytes derived from an Ob1771 mouse preadipocyte clonal line upon stimulation by A II. The stimulated adipocytes release a prostacyclin (PGI₂) which plays a key role in preadipose cell differentiation *in vitro* (Vassaux et al. 1992). Thus the nutritionally induced increase of local production of AOPEN by mature adipocytes may, via increased local levels of A II and PGI₂, lead to hyperplasia of adipose tissue, once the adipocytes reach a critical size (Kral 1976; Faust et al.1978). The A II stimulated prostacyclin release in cultured mouse adipocytes was totally blocked by the specific AT₂ ligand PD 123 177 (Darimont et al. 1994). On the other hand Crandall et al. (1994b) observed A II stimulated release of PGF_{1α}, a stable metabolite of prostacyclin, in adult rat adipocytes expressing exclusively AT₁ receptors. These conflicting results suggest either species differences or age differences. The latter is supported by the higher abundance of AT₂ subtype in fetal and young tissues (Saavedra 1992; Bottari et al. 1993).

The results of Darimont et al. (1994) suggest a role of A II in adipose tissue hyperplasia, while the results of Crandall et al. (1994b) show the possible role of this hormone in hypertrophy of fat tissue. Losartan, an AT₁ specific antagonist, inhibited the age dependent increase in adipocyte size (Crandall et al. 1994b). In the same experiment the number of AII receptors in adipocyte plasma membranes

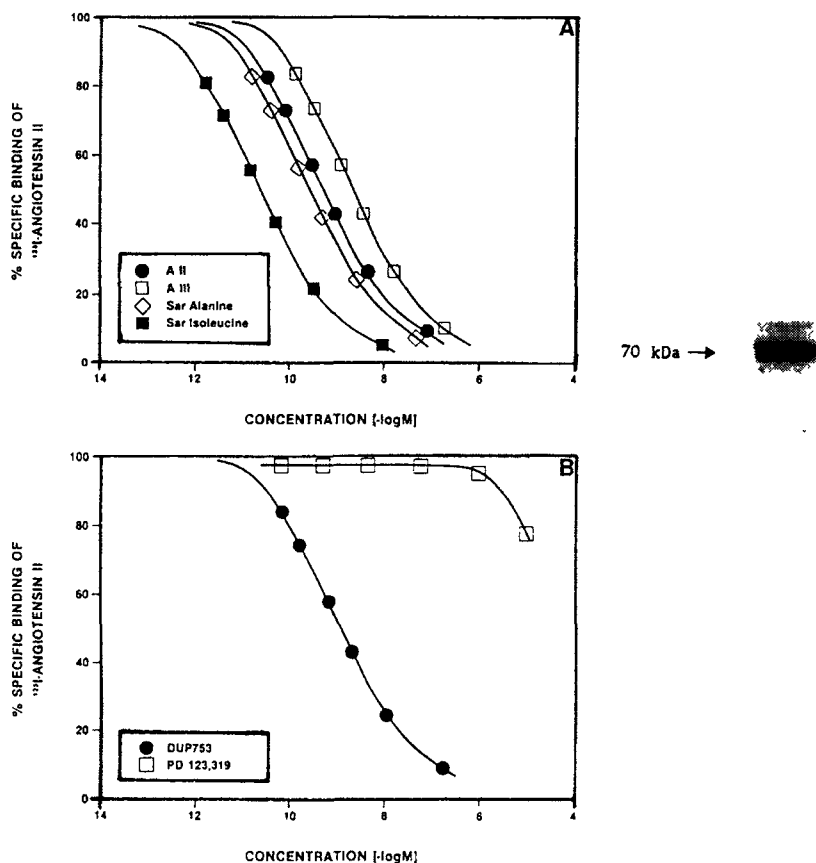


Figure 1. Left side: Displacement of ^{125}I -A II bound to fat cell membranes by A II, agonists and antagonists. (A) Compounds that are nonselective for receptor subtypes; (B) Selective antagonists for AT_1 (DuP 753 = losartan) and AT_2 (PD 123319), respectively [from Crandall et al. (1993); with permission]. Right side: Western-Blot analysis of membranes from rat epididymal fat. The AT_1 specific antibody was prepared as described in Zelezna et al. (1992).

was significantly lower in adult, obese than in young, lean rats. Hypothetically the increased adipose tissue mass in adult, obese rats produces more A II locally which may downregulate the A II receptors in adipocytes.

Taken together the recent experimental data underline the possible importance of AII and AII receptors in the regulation of adipocyte metabolism and in adipose tissue growth and development.

Therapeutic implications related to adipose tissue RAS

Since excessive enlargement of adipose tissue is an important risk factor for the development of prevalent disorders such as insulin resistance, diabetes and hypertension (Kissebah and Krakower 1994), elucidation of factors regulating adipose tissue growth is of major interest. Recent experimental data suggest a role for A II in this process (Crandall et al. 1994b; Darimont et al. 1994). Confirmation of the regulatory role of A II in development of adipose tissue and understanding of the function of RAS in this tissue may provide an important tool for treating obesity. Indeed, it was already shown that the losartan treatment of rats results in significant decrease of adipocytes cell volume (Crandall et al. 1994b). Moreover, clinical studies suggest that ACE inhibitors used as antihypertensive agents increase insulin sensitivity in patients with diabetes and hypertension (Connell and McLellan 1991; Lewis et al. 1993; Torlone et al. 1993; Okša et al. 1994). Interestingly, in healthy individuals A II infusion also enhanced insulin sensitivity (Fliser et al. 1993; Raymond and DiPette 1993). These data and the fact that fat tissue is one of the main targets for insulin suggest an interaction between the RAS and insulin receptors and the glucose transporter system in fat tissue. In addition, with increased use of losartan clinically, data on the effect of AT₁ receptor blockade on glucose metabolism, seem especially important in hypertensive diabetics.

Acknowledgements. This paper has been supported by grant of Slovak Academy of Sciences No. GA-SAV 2-541/95, and partially by grant of the Grant Agency of the Czech Republic No. GACR 306/94/1480. The authors wish to thank Dr. David L. Crandall (Wyeth-Ayerst Research, Princeton) for reviewing the manuscript and for valuable suggestions.

References

- Berridge M. J., Irvine R. F. (1989): Inositol phosphates and cell signaling. *Nature* **341**, 197—205
- Bottari S. P., King I. N., Reichlin S., Dahlstroem I., Lydon N., de Gasparo M. (1992): The angiotensin AT₂ receptor stimulates protein tyrosine phosphatase activity and mediates inhibition of particulate guanylate cyclase. *Biochem. Biophys. Res. Commun.* **182**, 1094—1099
- Bottari S. P., de Gasparo M., Steckelings U. M., Levens N. R. (1993): Angiotensin II receptor subtypes: characterization, signalling mechanism, and possible physiological implications. *Front. Neuroendocrinol.* **14**, 123—171
- Cassis L. A., Saye J., Peach M. J. (1988): Location and regulation of rat angiotensinogen messenger RNA. *Hypertension* **11**, 591—596
- Chiu A. T., Smith R. D., Timmermans P. B. M. W. M. (1994): Defining angiotensin receptor subtypes. In: *Angiotensin Receptors* (Eds. J. M. Saavedra and P. B. M. W. M. Timmermans), pp. 49—65, Plenum Press, New York
- Connell J. M. C., McLellan A. R. (1991): Hypertension, insulin and atherogenesis. *J. Cardiovasc. Pharmacol.* **18** (Suppl. 2), S45—S50

- Crandall D. L., Gordon G., Herzlinger H. E., Saunders B. D., Zolotor R. C., Cervoni P., Kral J. G. (1992): Transforming growth factor alpha and atrial natriuretic peptide in white adipose tissue depots in rats. *Eur. J. Clin. Invest.* **22**, 676—680
- Crandall D. L., Herzlinger H. E., Saunders B. D., Zolotor R. C., Feliciano L., Cervoni P. (1993): Identification and characterization of angiotensin II receptors in rat epididymal adipocyte membranes. *Metabolism* **42**, 511—515
- Crandall D. L., Herzlinger H. E., Saunders B. D., Armellino D. C., Kral J. G. (1994a): Distribution of angiotensin II receptors in rat and human adipocytes. *J. Lipid Res.* **35**, 1378—1385
- Crandall D. L., Herzlinger H. E., Saunders B. D., Kral J. G. (1994b): Developmental aspects of the adipose tissue renin-angiotensin system: therapeutic implications. *Drug Develop. Res.* **32**, 117—125
- Darimont C., Vassaux G., Ailhaud G., Negrel R. (1994): Differentiation of preadipose cells: paracrine role of prostacyclin upon stimulation of adipose cells by angiotensin-II. *Endocrinology* **135**, 2030—2036
- Dudley D. T., Panek R. L., Major T. C., Lu G. H., Bruns R. F., Klinkefus B. A., Hodges J. C., Weishaar R. E. (1990): Subclasses of angiotensin II binding sites and their functional significance. *Mol. Pharmacol.* **38**, 370—377
- Dudley D. T., Hubbel S. E., Summerfelt M. (1991): Characterization of angiotensin II (AT₂) binding sites in R3T3 cells. *Mol. Pharmacol.* **40**, 360—367
- Faust I. M., Johnson P. R., Stern J. S., Hirsch J. (1978): Diet- induced adipocyte number increase in adult rats: a new model of obesity. *Amer. J. Physiol.* **235**, E279—E286
- Fliser D., Arnold U., Kohl B., Hartung R., Ritz E. (1993): Angiotensin II enhances insulin sensitivity in healthy volunteers under euglycemic conditions. *Hypertension* **11**, 983—988
- Frederich Jr. R. C., Kahn B. B., Peach M. J., Flier J. S. (1992): Tissue-specific nutritional regulation of angiotensinogen in adipose tissue. *Hypertension* **19**, 339—344
- Griendling K. K., Murphy T. J., Alexander R. W. (1993): Molecular biology of renin-angiotensin system. *Circulation* **87**, 1816—1828
- Heemskerk F. M. J., Zorad S., Seltzer A., Saavedra J. M. (1993): Characterization of brain angiotensin II AT₂ receptor subtypes using [¹²⁵I]CGP 42112A. *NeuroReport* **4**, 103—105
- Hollenberg C. H. (1990): Perspectives in adipose tissue physiology. *Int. J. Obesity* **14** (suppl. 3), 135—152
- Kambayashi Y., Bardhan S., Takahashi K., Tsuzuki S., Inui H., Hamakubo T., Inagami T. (1993): Molecular cloning of a novel angiotensin receptor isoform involved in phosphotyrosine phosphatase inhibition. *J. Biol. Chem.* **268**, 24543—24546
- Kissebah A. H., Krakower G. R. (1994): Regional adiposity and morbidity. *Physiol. Rev.* **74**, 761—811
- Klyde B. J., Hirsch J. (1979): Increased cellular proliferation in adipose tissue of adult rats fed a high-fat diet. *J. Lipid Res.* **20**, 705—715
- Kral J. G. (1976): Surgical reduction of adipose tissue in the male Sprague-Dawley rat. *Amer. J. Physiol.* **231**, 1090—1096
- Lewis E. J., Hunsicker L. G., Bain R. P., Rohde R. D. (1993): The effect of angiotensin-converting-enzyme inhibition on diabetic nephropathy. *N. Engl. J. Med.* **329**, 1456—1462
- Missale C., Memo M., Sigala S., Carruba M. O., Spano P. F. (1989): Angiotensin II differentially effects cyclic AMP formation in intact adrenal glomerulosa cells and in

- purified membrane preparations. *Regul. Peptides* **24**, 167–178
- Mukoyama M., Nakajima M., Horiuchi M., Sasamura H., Pratt R. E., Dzau V. J. (1993): Expression cloning of type 2 angiotensin I receptor reveals a unique class of seven-transmembrane receptors. *J. Biol. Chem.* **268**, 24539–24542
- Murphy T. J., Alexander R. W., Griendling K. K., Runge M. S., Bernstein K. E. (1991): Isolation of cDNA encoding the vascular type-1 angiotensin receptor. *Nature* **351**, 233–236
- Okša A., Gajdoš M., Fedelešová V., Spustová V., Dzúrik R. (1994): Effects of angiotensin-converting enzyme inhibitors on glucose and lipid metabolism in essential hypertension. *J. Cardiovasc. Pharmacol.* **23**, 79–86
- Pobiner B. F., Hewlett E. L., Garrison J. C. (1985): Role of Ni in coupling angiotensin receptors to inhibition of adenylate cyclase in hepatocytes. *J. Biol. Chem.* **260**, 16200–16209
- Pfeilschifter J. (1989): Cross-talk between transmembrane signalling systems: a prerequisite for the delicate regulation of glomerular haemodynamics by mesangial cells. *Eur. J. Clin. Invest.* **19**, 347–361
- Pfeilschifter J., Huwiler A., Merriweather C., Briner V. A. (1992): Angiotensin II stimulation of phospholipase D in rat renal mesangial cells is mediated by the AT₁ receptor subtype. *Eur. J. Pharmacol.* **225**, 57–62
- Raymond R. T., DiPette D. J. (1993): Pressor doses of angiotensin II increase insulin-mediated glucose uptake in normotensive men. *Amer. J. Physiol. (Endocrinol. Metab. 28)*, E362–E366
- Saavedra J. M. (1992): Brain and pituitary angiotensin. *Endocrine Rev.* **13**, 329–380
- Sasaki K., Yamano Y., Bardhan S., Iwai N., Murray J. J., Hasegawa M., Matsuda Y., Inagami T. (1991) Cloning and expression of a complementary DNA encoding a bovine adrenal angiotensin II type-1 receptor. *Nature* **351**, 230–233
- Stoll M., Steckelings U. M., Paul M., Bottari S. P., Metzger R., Unger T. (1995): The angiotensin AT₂-receptor mediates inhibition of cell proliferation in coronary endothelial cells. *J. Clin. Invest.* **95**, 651–657
- Timmermans P. B. M. W. M., Wong P. C., Chiu A. T., Herblin W. F., Benfield P., Carini D. J., Lee R. J., Wexler R. R., Saye J. A. M., Smith R. D. (1993): Angiotensin II receptors and angiotensin II receptor antagonists. *Pharmacol. Rev.* **45**, 205–251
- Torlone E., Britta M., Ramboitti A. M., Perriello G., Santeusano F., Brunetti P., Bolli G. B. (1993): Improved insulin action and glycemic control after long-term angiotensin-converting enzyme inhibition in subjects with arterial hypertension and type II diabetes. *Diabetes Care* **16**, 1347–1355
- Tsutsumi K., Saavedra J. M. (1992): Heterogeneity of angiotensin II AT₂ receptors in the rat brain. *Mol. Pharmacol.* **41**, 290–297
- Tsutsumi K., Zorad S., Saavedra J. M. (1992): The AT₂ subtype of angiotensin II receptors has differential sensitivity to dithiothreitol in specific brain nuclei of young rats. *Eur. J. Pharmacol.* **226**, 169–173
- Vassaux G., Gaillard D., Ailhaud G., Negrel R. (1992): Prostacyclin is a specific effector of adipose cell differentiation: its dual role as a cAMP and Ca²⁺ elevating agent. *J. Biol. Chem.* **267**, 11092–11097
- Whitebread S., Mele M., Kamber B., de Gasparo M. (1989): Preliminary characterization of two angiotensin II receptor subtypes. *Biochem. Biophys. Res. Commun.* **163**, 284–291

- Zelezna B , Richards E M, Tang W , Lu D , Sumners C , Raizada M K (1992) Characterization of a polyclonal anti-peptide antibody to the angiotensin II type-1 (AT₁) receptor *Biochem Biophys Res Commun* **183**, 781—788
- Zorad S , Xu N , Heemskerk F M J , Gutkind S J, Saavedra J M (1994) Characterization of AT₂ receptor expression in NIH 3T3 fibroblast cell line *Cell Biol Int* **18**, 491

Final version accepted October 24, 1995